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RE: New C.I.P. Patent Application in U.S.
Dana FOWLKES et al.
Atty's Docket: FOWLKES=4B

Sir:

Transmitted herewith is a continuation-in-part patent application entitled **IDENTIFICATION OF DRUGS USING COMPLEMENTARY COMBINATORIAL LIBRARIES**. The inventors are Dana M. FOWLKES, a U.S. citizen of 2013 Damascus Church Road, Chapel Hill, North Carolina, 27516, Brian K. KAY, a U.S. citizen of 18 Wysteria Way, Chapel Hill, North Carolina, 27514, Jeffrey A. FRELINGER, a U.S. citizen of 1111 Monterey Valley Road, Chapel Hill, North Carolina, 27516, and Robin Parish HYDE-DERUYSCHEER, a U.S. citizen of 205 Portsmouth Place, Chapel Hill, North Carolina, 27516. This application is a continuation-in-part of PCT application PCT/US97/19638, filed October 31, 1997, now pending, which is a continuation-in-part of U.S. Serial Number 08/740,671, filed on October 31, 1996, now pending.

Attached are the specification (143 pages), including 24 claims (3 independent) and an abstract, plus 19 sheets of drawings (Figures 1-24), and a return postcard. In accordance with 37 CFR 1.53(a) and (b), it is respectfully requested that a serial number and filing date be assigned to this application as of the date of receipt of the present papers. In accordance with the present procedures of the U.S. Patent and Trademark Office, an executed Declaration and the filing fee for the present application will be filed in due course.¹

¹ No authorization is given for charging the filing fee at the present time. However, at such time that the declaration is filed, but not before, you are authorized to charge whatever excess fees are necessary (including the filing fee and any

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In re of FOWLKES=4B

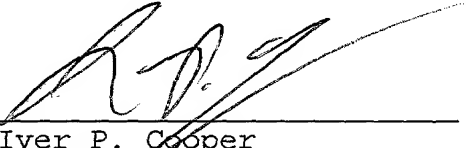
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Respectfully submitted,
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extension of time fees then due) to Deposit Account 02-4035, if any such fees due are not fully covered by check filed at that time.

**IDENTIFICATION OF DRUGS USING COMPLEMENTARY
COMBINATORIAL LIBRARIES**

This application is a continuation-in-part of PCT/US97/19638, filed October 31, 1997, now pending, which is
5 a continuation-in-part of Serial No. 08/740,671, filed October 31, 1996, now pending, which applications are hereby incorporated by reference in their entirety.

BACKGROUND OF THE INVENTION

Field of the Invention

10 This invention relates to methods of identifying drugs which can mediate the biological activity of a target protein.

Protein Binding and Biological Activity

Many of the biological activities of the proteins are attributable to their ability to bind specifically to one or
15 more binding partners (ligands), which may themselves be proteins, or other biomolecules.

When the binding partner of a protein is known, it is relatively straightforward to study how the interaction of the binding protein and its binding partner affects biological
20 activity. Moreover, one may screen compounds for the ability of the compound to competitively inhibit the formation of the complex, or to dissociate an already formed complex. Such inhibitors are likely to affect the biological activity of the protein, at least if they can be delivered in vivo to the site
25 of the interaction.

If the binding protein is a receptor, and the binding partner an effector of the biological activity, then the inhibitor will antagonize the biological activity. If the binding partner is one which, through binding, blocks a
30 biological activity, then an inhibitor of that interaction will, in effect, be an agonist.

The residues whose functional groups participate in the ligand-binding interactions together form the ligand binding site, or paratope, of the protein. Similarly, the functional
35 groups of the ligand which participate in these interactions

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In the case of a protein, the binding sites are typically relatively small surface patches. The binding characteristics of the protein may often be altered by local modifications at these sites, without denaturing the protein.

What is called the hydrophobic interaction is actually the absence of hydrogen bonding between nonpolar groups and water, rather than a favorable interaction between the nonpolar groups themselves. Hydrophobic interactions are important in stabilizing the conformation of a protein and thus indirectly affect ligand binding, although hydrophobic residues are usually buried and thus not part of the binding site.

Peptides have been found to bind proteins at the same sites as those by which the proteins interact with other proteins, macromolecules and biologically significant substances e.g. nucleic acids, lipids and enzyme substrates. The first examples of this property were in the publications of several groups who showed that there is a single peptide binding site on the biotin binding protein streptavidin. This is the same site responsible for biotin binding and these peptides compete with biotin for binding to this site (Biochemistry 34: 15430-15435 (1995) Screening of cyclic peptide phage libraries identifies ligands that bind streptavidin with high affinities, L. B. Giebel, R. T. Cass, D. L. Milligan, D. C. Young, R. Arze & C. R. Johnson; Gene 128: 59-65 (1993) An M13 phage library displaying random 38-amino-acid peptides as a source of novel sequences with affinity to selected targets, B. K. Kay, N. B. Adey, Y. S. He, J. P. Manfredi, A. H. Mataragnon & D. M. Fowlkes; Nature 354: 82-4 (1991) A new type of synthetic peptide library for identifying ligand-binding activity K. S. Lam, S. E. Salmon, E. M. Hersh, V. J. Hruby, W. M. Kazmierski & R. J. Knapp; Proc Natl Acad Sci

U S A 92: 5426-5430 (1995) Library of libraries: approach to synthetic combinatorial library design and screening of "pharmacophore" motifs, N. F. Sepetov, V. Krchnak, M. Stankova, S. Wade, K. S. Lam & M. Lebl; Biochem J 293 (Pt 3): 613-6 (1993) Biotin binders selected from a random peptide library expressed on phage, I. Saggio & R. Laufer). Many other examples exist, for instance Smith demonstrated that peptides displayed on phage which bound to ribonuclease S had a specific consensus motif and that these PLs were antagonistic to ribonuclease activity, implying that the peptides and the RNA were bound by the same ligand binding site (Gene 128: 37-42 (1993) A ribonuclease S-peptide antagonist discovered with a bacteriophage display library, G. P. Smith, D. A. Schultz & J. E. Ladbury). Another example is from the binding of peptide ligands to cell surface integrins (Biochemistry 34: 3948-3955 (1995) Peptide ligands for integrin alpha v beta 3 selected from random phage display libraries, J. M. Healy, O. Murayama, T. Maeda, K. Yoshino, K. Sekiguchi & M. Kikuchi; J Cell Biol 124: 373-80 (1994) Isolation of a highly specific ligand for the alpha 5 beta 1 integrin from a phage display library, E. Koivunen, B. Wang & E. Ruoslahti). Peptides obtained in this way clearly mimic natural protein:protein interactions as in the case for the proteins MDM2 and p53 (Bottger et al. Identification of novel mdm2 binding peptides by phage display, Oncogene, 13:2141-7 (1996)). However, it has not hitherto been appreciated that this phenomenon is sufficiently common so that it might be exploited in identifying inhibitors of the interaction of a protein with an unknowing binding partner. Nor have others explained just how to take advantage of this phenomenon for that purpose.

Traditional Drug Screening

In traditional drug screening, natural products (especially those used in folk remedies) were tested for biological activity. The active ingredients of these products were purified and characterized, and then synthetic analogues of these "drug leads" were designed, prepared and tested for activity. The best of these analogues became the next

generation of "drug leads", and new analogs were made and evaluated.

Both natural products and synthetic compounds could be tested for just a single activity, or tested exhaustively for any biological activity of the interest to the tester. Testing was originally carried out in animals, later, less expensive and more convenient model systems, employing isolated organ, tissue, or cell cultures, membrane extracts or purified receptors, were developed for some pharmacological evaluations.

These methods have many disadvantages. Many of these approaches require large amounts of chemical compound to test, especially testing in whole animals and isolated organs. Since the quantity of a given compound within a collection of potential medicinal compounds is limited, this requires one to limit the number of screens executed.

Also, it is inherently difficult to establish structure/activity relationships (SAR) among compounds tested using whole animals, isolated organs and cultured cells. This is because the actual molecular target of any given compound's action may be quite different from that of other compounds scoring positive in the assay. By testing a battery of compounds on a very specific target, one can correlate the action of various chemical residues with the quantitative activity and use that information to focus ones search for active compounds among certain classes of compounds or even direct the synthesis of novel compounds having a composite of the properties shared by the active compounds tested.

Another disadvantage to whole animal, organ and cell based screening is that certain limitations may prevent an active compound from being scored as such. For instance, an inability to pass through the cellular membrane may prevent a potent inhibitor, within a tested compound library, from acting on the activated oncogene ras and giving a spurious negative score in a cell proliferation assay. However, if it were possible to test ras in an isolated system, that potent inhibitor would be scored as a positive compound and contribute to the establishment of a relevant SAR. Subsequent, chemical modifications could then be carried out to optimize the

compound structure for membrane permeability.

The overwhelming disadvantage to the receptor based methods for screening compounds is that they require a *priori* knowledge about the activity of receptor and its biological ligand. If through genetic mapping of a disease loci one determines that a particular gene product is responsible for the disease; and one lacks knowledge about the gene biochemical function because it is not a previously known receptor or enzyme, then it is very difficult to establish an assay with the methods previously known.

The present invention circumvents all these problems.

Combinatorial Libraries

Libraries of thousands, even millions, of random oligopeptides have been prepared by chemical synthesis (Houghten et al., Nature, 354:84-6(1991)), or gene expression (Marks et al., J Mol Biol, 222:581-97(1991)), displayed on chromatographic supports (Lam et al., Nature, 354:82-4(1991)), inside bacterial cells (Colas et al., Nature, 380:548-550(1996)), on bacterial pili (Lu, Bio/Technology, 13:366-372(1990)), or phage (Smith, Science, 228:1315-7(1985)), and screened for binding to a variety of targets including antibodies (Valadon et al., J Mol Biol, 261:11-22(1996)), cellular proteins (Schmitz et al., J Mol Biol, 260:664-677(1996)), viral proteins (Hong and Boulanger, Embo J, 14:4714-4727(1995)), bacterial proteins (Jacobsson and Frykberg, Biotechniques, 18:878-885(1995)), nucleic acids (Cheng et al., Gene, 171:1-8(1996)), and plastic (Siani et al., J Chem Inf Comput Sci, 34:588-593(1994)).

Libraries of proteins (Ladner, USP 4,664,989), peptoids (Simon et al., Proc Natl Acad Sci U S A, 89:9367-71(1992)), nucleic acids (Ellington and JW, Nature, 246:818(1990)), carbohydrates, and small organic molecules (Eichler et al., Med Res Rev, 15:481-96(1995)) have also been prepared or suggested for drug screening purposes.

Sparks, et al., Nature Biotechnology, 14:741 (June 1996) used an SH3 domain-binding peptide isolated from a phage-displayed random peptide library to screen a 16-day mouse

Patented by the University of California, San Diego

5 libraries for additional SH3-domain-binding ligands.

10 In addition, the variants are composed of linear polymers such
that each actually represents a sliding window of many
differing chemical constituents. For instance, if a given
macromolecular interaction is based on the side chains of four
amino acids within a binding peptide, then a 13 amino acid
15 peptide has 10 potential combinations of residues which may
bind; therefore a library of 10^8 members has about 10^9 4-mer
permutations. This, combined with ease of producing and
screening exceptionally large and diverse peptide libraries,
provides the incentive to use peptide combinatorial libraries
20 for the initial identification and probing of protein
functional domains.

25 transit cellular membranes. Other classes of compounds have better properties for drug candidates. However, historically, acquiring chemical compound libraries has been a barrier to the entry of smaller firms into the drug discovery arena. Due to the large quantity of chemical required for testing on whole
30 animals and even on cells in culture, it was a given that whenever a compound was synthesized it should be done in fairly large quantity. Thus, there was a synthesis and purification throughput of less than 50 compounds per chemist per year. Large companies maintained their immensely valuable collections
35 as trade barriers. However, with the downsizing of targets to the molecular level and the automation of screens, the quantity of a given compound necessary for an assay has been reduced to very small amounts. These changes have opened the door for the

utilization of so-called combinatorial chemistry libraries in lieu of the traditional chemical libraries. Combinatorial chemistry permits the rapid and relatively inexpensive synthesis of large numbers of compounds in the small quantities suitable for automated assays directed at molecular targets. Numerous small companies and academic laboratories have successfully engineered combinatorial chemical libraries with a significant range of diversity (reviewed in Doyle, 1995, Gordon et al, 1994a, Gordon et al, 1994b).

We have developed a systematic means for development of drug discovery screens for numerous targets. One of the special advantages of this system is that the high throughput screens are essentially identical for similar and dissimilar targets, bypassing the need to develop distinct assays for biochemically diverse targets. This is desirable for several reasons. First and foremost, one is never certain how useful a specific target is for therapeutic intervention. It is not until active compounds have been isolated and tested that one can truly "validate" a molecular target. Thus it makes sense to choose as many targets as practical, establish screens for each and then validate each target pharmacologically using the identified compounds. Second, for many potential targets one may not be aware of a biochemical activity that can be used to establish molecular assays. Many potential targets can be proposed based upon the results of genetic experimentation rather than biochemical data. This has been the case for viruses due to ease of subcloning and mutagenic analysis and, now, with the outpouring of human genetic data, shall be true in many other disease areas. The challenge is to go from genetic data to development of useful drug screens.

All references, including any patents or patent applications, cited in this specification are hereby incorporated by reference. No admission is made that any reference constitutes prior art. The discussion of the references states what their authors assert and applicants reserve the right to challenge the accuracy and pertinency of the cited documents.

SUMMARY OF THE INVENTION

The present invention relates to a method of identifying drugs which can mediate the biological activity of a target protein via inhibition of binding of the target protein to a binding partner. Unlike prior methods, it does not require that the natural binding partner be used as a reagent, or even that it have been characterized. The need for the natural binding partner is obviated by the use of complementary combinatorial libraries.

The term "library" generally refers to a collection of chemical or biological entities which can be screened simultaneously for a property of interest. (They may be screened sequentially, if desired, but simultaneous screening is more efficient.) Typically, they are related in origin, structure, and/or function.

The term "combinatorial library" refers to a library in which the individual members are either systematic or random combinations of a limited set of basic elements, the properties of each member being dependent on the choice and location of the elements incorporated into it. Typically, the members of the library are at least capable of being screened simultaneously. Randomization may be complete or partial; some positions may be randomized and others predetermined, and at random positions, the choices may be limited in a predetermined manner. The members of a combinatorial library may be oligomers or polymers of some kind, in which the variation occurs through the choice of monomeric building block at one or more positions of the oligomer or polymer, and possibly in terms of the connecting linkage, or the length of the oligomer or polymer, too. Or the members may be nonoligomeric molecules with a standard core structure, like the 1,4-benzodiazepine structure, with the variation being introduced by the choice of substituents at particular variable sites on the core structure.

The ability of one or more members of such a library to recognize a target molecule is termed "Combinatorial Recognition".

In a "simple combinatorial library", all of the members

belong to the same class of compounds (e.g., peptides) and can be synthesized simultaneously. A "composite combinatorial library" is a mixture of two or more simple libraries, e.g., DNAs and peptides. The number of component simple libraries in a composite library will, of course, normally be smaller than the average number of members in each simple library, as otherwise the advantage of a library over individual synthesis is small.

A biased combinatorial library is one in which, at one or more positions in the library member, only one of the possible basic elements is allowed for all members of the library, i.e., the biased positions are invariant.

The term "amplifiable combinatorial library" refers to a library in which the individual members, after found to bind to a target, may be amplified in vivo or in vitro, using elements already present in the library as starting materials. There are two classes of amplifiable members. First, nucleic acids may be amplified in vivo through natural replicative processes, or in vitro through techniques such as polymerase chain reaction (PCR). Second, peptides, when presented on phage, or otherwise associated with an encoding nucleic acid, may be amplified indirectly by in vivo or in vitro amplification of the associated nucleic acid encoding the peptide, the amplified nucleic acid being expressed to produce the peptide.

The term "biopolymeric library" refers to a library composed of peptides (together with peptoids), nucleic acids, and/or oligosaccharides. (It is not necessary that they be composed of naturally occurring amino acids, bases, or sugars, respectively.) However, because of the greater complexity of carbohydrate synthesis, peptides and nucleic acids are of greater interest.

A "panel of combinatorial libraries" is a collection of different (although possibly overlapping) and separately screenable simple or composite combinatorial libraries. A "panel" differs from a composite library in that the component simple libraries have not been mixed together, that is, they may still be screened separately.

A "structural panel" is a panel as defined above where there is some structural relationship between the member libraries. For example, one could have a panel of 20 different biased peptide libraries where, in each library, the middle
5 residue is held constant as a given amino acid, but, in each library the constant residue is different, so, collectively, all 20 possible genetically encoded amino acids are explored by the panel.

A "scanning residue library" refers to the preparation of
10 panel of biased combinatorial peptide libraries such that the position of the constant residue shifts from one library to the next. For example, in library 1, residue 1 is held constant as a particular residue AA, in library, residue 2 is, and so forth through two or more (usually all) positions of the
15 peptide.

One may have structured panels of libraries in which one may define subpanels, too. For example, in one subpanel, the middle residue AA₁ may be the same for all libraries, but the libraries also have a constant residue AA₂ which is scanned
20 through all other residue positions.

A library screening program is a program in which one or more libraries (e.g., a structured panel of biased peptide libraries) are screened for activity. The libraries may be screened in parallel, in series, or both. In serial screening,
25 the results of one screening may be used to guide the design of a subsequent library in the series.

Applicants screen a first combinatorial library for binding to the target protein. Applicants then screen a second (complementary) library (preferably combinatorial in nature)
30 for the ability to inhibit the binding of one or more of the target binding ligands in the first library to the target protein.

The successful inhibitors are candidate antagonists of one or more of the biological activities of the target protein.

35 Applicants believe that those members of a combinatorial library, especially a biopolymer library, which bind to a target protein having a biologically significant binding activity will bind preferentially to the sites at which the

target protein interacts with the natural binding partners which mediate its biological activity, as opposed to randomly, with equal probability, over the entire surface of the target protein. If so, then the target-binding members of the library in question can be used as surrogates for an unknown or unavailable natural binding partner in screening a second combinatorial library (the "complementary library"), which need not be a biopolymeric library, for members which can inhibit the complexing of target protein to its natural binding partner. The surrogate library is preferably a peptide or nucleic acid library.

The size of a library is the total number of molecules in it, whether they be the same or different. The diversity of a library as the number of different molecules in it. "Diversity" does not measure how different the structures of the library; the degree of difference between two structures is referred to here as "disparity" or "dispersion". The "disparity" is quantifiable in some respects, e.g., size, hydrophilicity, polarity, thermostability, etc. The average sampling frequency of a library is the ratio of size to diversity. The sampling frequency should be over the detection limit of the assay in order to assure that all members are screened.

The combinatorial libraries usually will have a diversity of at least 10^3 different structures. Preferably, the initial, surrogate-generating library is of high diversity, e.g., preferably at least about 10^6 , more preferably at least about 10^9 different members. While a peptide library is preferred, a library composed of a different class of compounds (e.g., peptoids or nucleic acids) is acceptable if there would be a detectable preference for binding the activity-mediating binding sites of the target protein.

The complementary library need not be, and preferably is not, a peptide library and it may be of lower overall diversity. It may be screened against all of the surrogate peptides; or only against selected ones. The screenings may be individual or collective. Often, the members of the complementary library will be less specific in their binding

to the paratopes of the target protein than are the members of the first library, possibly because their surface area is smaller and offers fewer opportunities for favorable (or unfavorable) interactions with other molecules. A preferred
5 complementary library is a benzodiazepine library.

The degree of complex-inhibitory activity of the members of the complementary library may be quantified by means of a labeled surrogate peptide and an insolubilized target protein. Either the amount of labeled surrogate peptide is fixed, and
10 the amount of complementary compound varied, or, more preferably, the amount of labeled surrogate peptide is varied and the amount of complementary compound held constant. The greater the activity of the complementary compound, the less labeled surrogate peptide will be in the solid phase (i.e.,
15 complexed to the target protein) and the more will be in the liquid phase (i.e., uncomplexed). The amount of label in either phase is then measured and correlated with the amount of the variable component. Conventional method of screening libraries for binding molecules do not lend themselves to
20 quantification of the degree of affinity.

It is possible that some of the target protein binding members of the first library will not bind the target protein at the site bound by the natural binding partner which mediates the biological activity of interest, or bind it that site but
25 still do not have an effect similar to that of the natural binding partner, i.e., that these nominal surrogates are not true surrogates for the natural binding partner. However, as long as one or more of the identified members are true surrogates, if all of the nominal surrogates are used in
30 screening the complementary library, then one necessarily will screen for inhibitors of the binding of the true surrogates to the target protein, too.

To reduce the number of "false hits" generated (i.e., compounds which inhibit the binding of a false surrogate to the
35 target protein, or which inhibit binding of a true surrogate but at the wrong site), one may first test the nominal surrogates in a suitable biological system, for the ability to interact with the target protein so as to mediate its

biological activity of interest (or at least a related activity that is evaluatable in that biological system). Then only those nominal surrogates which are active in this model system are used in screening the complementary library.

5 It is expected that most of the compounds of the complementary library which inhibit the complexing of the surrogate peptide to the target protein will achieve this inhibition by binding to the target protein in such a manner as to block its interaction with the surrogate peptide. While
10 it is theoretically possible that the complementary compound will bind to the surrogate peptide instead of the target protein, this interaction is likely to be weak, since most oligopeptides do not have a stable conformation.

It is, of course, a simple matter to distinguish
15 inhibitory compounds which bind the target protein from those which bind the surrogate peptide by use of either the target protein or surrogate peptide alone, in labeled or immobilized form, as an assay or affinity separation reagent.

BRIEF DESCRIPTION OF THE DRAWINGS

20 Figure 1. Benzodiazepine scaffold used to create a combinatorial library, and a synthetic pathway leading to that structure.

Figure 2. Vector pFLAG-ATS-BAP for expression of peptide ligands fused to bacterial alkaline phosphatase. This vector
25 has been used to successfully express both large and small domains of signaling proteins. In most instances the protein is secreted and one simply concentrates the fusion protein by ammonium sulfated precipitation from the media. In rare instances the protein is not secreted, but accumulates within
30 the cells. In this case, we wash the cells in tris buffered saline then sonicate to release active fusion protein. In either instance, sufficient amounts of the material are obtained from 100ml cultures. The FLAG epitope is disclosed, as a tag for a fusion protein, in Grihalde, et al., Gene,
35 166:187-95 (1995).

Figure 3. Occurrence of Amino Acids (AA) in Binding Peptides Identified by screening from Phage Display Libraries.

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For each of 16 targets, the sequences of the binding peptides identified by screening phage display libraries were analyzed. First, for peptides binding a given target, a consensus sequence was determined, and from this, a core binding region. The amino acids in the core region of all the peptides binding that target were tallied, and the tallies were divided by the number of peptides in question, to obtain a subtotal for each target. The target subtotals were then added and divided by the number of targets. The final totals were converted into percentages. If all of the residues were represented equally their values would be 5%.

Figure 4. Occurrence of amino acid residues in binding peptides identified by screening phage display libraries, after correcting for codon usage. The numbers in figure 3 were divided by the number of codons available for each residue using an NNK coding scheme and the resulting numbers normalized to 100%.

Figure 5. Enrichment of Phage Binding to CMV UL44. The proportion of phage binding to UL44 was monitored after 2 and three rounds of selection as described in the text. The ratio represents the number of blue plaques/# of white plaques. The library letters represent the residue held constant in each library.

Figure 6. Individual phage ELISAs for CMV UL44. Individual clones were picked after three rounds of selection on UL44 and tested for binding in a phage ELISA format. Letters under the graph represent the libraries that individual clones were isolated from. Letters A-H in the legend represent the row designation on the microtiter plate.

Figure 7. Competition for binding between phage and substrates. Competitive binding experiments were carried out as described in the text using GSTUL44 immobilized on plates as a target. Various concentrations of (a) glutathione (Fig. 7a) or (b) DNA (Fig. 7b) was added to each well and the binding of phage was monitored using a phage ELISA. Square boxes represent phage which bind to the GST portion of the fusion and diamonds represent phage which bind to the UL44 portion of the

fusion protein.

Figure 8. ELS Assay of UL44 BioKey (surrogate ligand). The peptide isolated from affinity selection against UL44 was synthesized and tested for specific binding as described in the text. (ELS=Enzyme Linked Spectrophometric). The targets tested were streptavidin, GST-UL44, GST, and GST-src SH3. The amount of biotinylated peptide binding was monitored using a streptavidin-HRP conjugate.

Figure 9. Time Course of 50 pmol UL44 BioKey (surrogate ligand) Binding to GST-UL44. The binding of the UL44 Surrogate ligand was monitored as a function of time. The signal increases in a linear fashion for at least 4 hours.

Figure 10. Titration Curves for Binding of UL44 BioKey (surrogate ligand) to GST-UL44. The binding of biotinylated peptide was monitored as a function of target concentration immobilized on the plate (from 0.125 micrograms to 2 micrograms) and as a function of peptide concentration (from 0.1 to 0.5 micrograms).

Figure 11. UL44 BioKey (surrogate ligand): Self-Competition between biotinylated UL44 surrogate ligand and non-biotinylated surrogate ligand was determined. Competitor is the concentration on non-biotinylated UL44 peptide added. The signals are presented as a percent of binding compared to no inhibitor added.

Figure 12. Titration of PKC binding phage. A serial dilution of phage were incubated with immobilized PKC and the amount of phage binding monitored using a phage ELISA.

Figure 13a. Specific Binding of the ProRS BioKey (surrogate ligand). (a) The peptide isolated from affinity selection against ProRS was synthesized and tested for specific binding as described in the text. The targets tested were ProRS, GST-src SH3, GST, GST-UL44, TyrRS and GST-MDM2. The amount of biotinylated peptide binding was monitored using a streptavidin-HRP conjugate. Fig. 13b BioKeys (Surrogate ligands) for other targets do not bind to ProRS. Surrogate ligands for src SH3, UL44, and MDM2 were tested for binding to ProRS as described in the text.

Figure 14. Self competition for ProRS BioKey. Self-

Competition between biotinylated ProRS surrogate ligand and non-biotinylated surrogate ligand was determined. Competitor is the concentration on non-biotinylated ProRS peptide added. BioKey concentration is the concentration of biotinylated ProRS surrogate ligand used. The signals are presented as a percent of binding compared to no inhibitor added and are graphed in two formats: (a) linear (Fig. 14a) and (b) log (Fig. 14b) scales of the competitor concentrations.

Figure 15. Concentration and time dependence of ProRS surrogate ligand binding. (a) The binding of biotinylated peptide was monitored as a function of peptide concentration (from 0.1 to 0.5 micrograms) (Fig. 15a). (b) The binding of biotinylated peptide was monitored as a function of time (Fig. 15b).

Figure 16. Other BioKeys (Surrogate ligands) for other targets do not bind to TyrRS. Surrogate ligands for src SH3, UL44, MDM2 and ProRS were tested for binding to ProRS as described in the text.

Figure 17. Binding Specificity of the TyrRS BioKey (surrogate ligand). The peptide isolated from affinity selection against TyrRS was synthesized and tested for specific binding as described in the text. The targets tested were TyrRS, GST-src SH3, GST, GST-UL44, GST-MDM2 and ProRS. The amount of biotinylated peptide binding was monitored using a streptavidin-HRP conjugate.

Figure 18. Concentration dependent binding of the TyrRS BioKey (surrogate ligand). The binding of biotinylated peptide was monitored as a function of peptide concentration (from 0.1 to 0.5 micrograms).

Figure 19. Time Course for Binding of TyrRS BioKey (biotinylated peptide).

Figure 20. Competition between HiTryRs BioKey and CB239 and CB16914 for binding between surrogate ligand and inhibitor. Competitive binding experiments were carried out as described in the text using TyrRS immobilized on plates as a target. Various concentrations of inhibitor CB239 was added to each well and the binding of surrogate ligand was monitored using a ELSA. Diamonds represent competition with a specific

inhibitor (CB239) which binds to the active site of the enzyme. Square boxes represent competition with a related inhibitor (CB16914) which has no effect on TyrRS activity.

Figure 21. Competition for binding between phage and an inhibitor. Competitive binding experiments were carried out as described in the text using beta-glucosidase immobilized on plates as a target. The binding of phage was monitored using a phage ELISA. Solid bars represent phage binding in the absence of inhibitor and hatched bars represent phage binding in the presence of inhibitor.

Figure 22. Structural and Functional Domains of the Estrogen Receptor. Structural domains are labeled A-F. Functional domains for hormone binding, dimerization, DNA binding, and transactivation are indicated.

Figure 23. Competition of estrogen receptor (ER) binding phage with estradiol and tamoxifen in a phage ELISA-format.

Figure 24A. Binding Activity of phage with LXXLL motif to Estrogen Receptor, in presence of estradiol, estriol, tamoxifen, nafoxidene, clomiphene, or without a competitor.

Figure 24B. Binding Activity of phage without LXXLL motif to Estrogen Receptor, in presence of estradiol, estriol, tamoxifen, nafoxidene, clomiphene, or without a competitor.

In both Figs. 24A and 24B, the phage were selected for binding to ER in the presence of estradiol.

25 DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is directed to the identification of compounds in a compound library which can mediate the biological activity of a target receptor protein, even when the ligands which mediate that activity through binding to that receptor are not already known. Such compounds can then be used as "drug leads", i.e., used as a starting point for the design of analogues which can in turn be tested for activity.

The method of the invention comprises three steps:

- (1) Screen at least one potential surrogate combinatorial library (as previously defined) for members (preferably peptides or nucleic acids) binding to the target protein (TP) and hence capable

(2) Screen at least one complementary library, preferably a combinatorial library, (which is not limited to, and may not even include, peptides, or nucleic acids and hence is referred to on occasion as a "compound library") for compounds which inhibit the binding of one or more surrogates (peptides or nucleic acid which bind TP to TP.

10 (3) Determine whether the inhibitory compound mediates
the biological activity of the said TP.

A library screening program may include multiple rounds of surrogate library screening, with the first library sampling a broad "sequence space", and later libraries focusing on sequences related to those previously found to bind the target. Or the different libraries may sample different sequence spaces, such as peptides and nucleic acids, or peptides of different length or composition. Similarly, it may include multiple rounds of screening of complementary library. The later rounds may sample the same class of compounds, a different class of compounds, or a subclass of those previously screened. Different rounds may use different surrogates in the screen. The screenings may occur in any rational order, e.g., surrogate/complementary/surrogate/complementary, or surrogate/surrogate/complementary/complementary. The compounds of the complementary library may be screened simultaneously or sequentially.

If the invention is described below with specific reference to peptide surrogates, such description applies mutatis mutandis to nucleic acids, too.

In effect, since the natural ligands of the receptor protein are not known (or for some other reason are not desirable for use in screening) a peptide library is searched to identify surrogate ligands. Of course, even if a given peptide does bind the receptor protein, there is no guarantee that it binds it at the correct site. However, by screening a diverse peptide library, many TP-binding peptides can be identified. It is likely that at least one of these peptides

will bind the TP at the same site as it is bound by a natural ligand. The peptide library "samples" the surface of the TP for sites having a high interaction potential. It is likely that the TP evolved to offer one or more such sites and that
5 such a site evolved to permit binding by the natural ligand (or that the natural ligand evolved to bind such a site on the TP). Preferably, the peptide library is a biased library as hereafter defined.

If desired, once TP-binding peptides have been identified,
10 they may be screened for ability to act as a surrogate for the natural ligand, i.e., to mediate the biological activity of the TP. If so, then only the TP-binding peptides with this mediative property need be used in the second step.

In the second step, a compound library is screened for the
15 ability of the compounds to inhibit the binding of the TP-binding peptides of step (1) with the TP. The compounds need not be peptides, and may be screened sequentially or simultaneously.

The affinity range of the TP-binding (surrogate) peptides
20 for the TP must be such that binding is detectable, and that inhibition of such binding by the compounds sought to be found is also detectable. Initially, the compounds screened are likely to have low inhibitory activity. As lead optimization progresses, compounds of higher inhibitory activity are more
25 likely to be present. Different surrogate peptides may be preferred for initial and subsequent rounds of library construction and screening. Generally speaking, affinity in the range of 10^{-5} to 10^{-11} M is desirable. The affinity of the surrogate ligand for the TP and the affinity range of compounds
30 sought for in the first round of will determine the concentrations of surrogate ligand and drugs used in the screen. It is preferable to keep the surrogate ligand concentration below its binding constant and the drug lead concentration above its binding constant. For example, if a
35 peptide surrogate ligand has a binding constant of 1×10^{-7} M, it should be used at concentrations of less than this in the assay. Conversely, if one wished to find compounds which bound to the TP with affinities around 1×10^{-6} M, then the compounds

should be present at higher concentrations than this in the assay. This does place limits on useful ligands and compounds. The ligand must be used at a concentration where its binding can still be detected. Thus with present detection methods, it is not desirable to use surrogate ligands with binding constants less (affinity greater) than 1×10^{-12} M, and compounds with binding constants greater (affinities poorer) than 10^{-6} M are not likely to be detected. As detection limits improve, the desired affinities will change.

The foregoing is not to be construed as limiting the invention to a particular affinity range.

The compounds may inhibit the binding of the TP-binding peptides to the TP either by binding to the peptide, or to the TP. Those that actually bind to the TP are more likely to mediate the biological activity of the TP. Those that bind to the TP-binding peptide alone are less likely to be useful, unless the TP-binding peptide is a true surrogate for the natural ligand, i.e., the compound cross-inhibits the natural ligand.

Finally, we determine whether the inhibitory compounds in fact mediate the biological activity of the TP.

In theory, it is possible to screen the compound library directly for ability to interact with the TP. For instance one can label the TP directly (see below) and test binding directly to a library of diverse compounds which are themselves immobilized on pins (Proc Natl Acad Sci U S A 91: 4708-12 (1994) The combinatorial synthesis and chemical and biological evaluation of a 1,4-benzodiazepine library, B. A. Bunin, M. J. Plunkett & J. A. Ellman) or beads (Proc Natl Acad Sci U S A 90: 10922-6 (1993) Complex synthetic chemical libraries indexed with molecular tags, M. H. Ohlmeyer, R. N. Swanson, L. W. Dillard, J. C. Reader, G. Asouline, R. Kobayashi, M. Wigler & W. C. Still). However, such screening by direct binding severely limits the selection of compound libraries that can be screened. For instance, many large pharmaceutical companies have accumulated many thousands of compounds over the years. Such compounds are stored either as dry powders or in solution and hence can not be assayed for direct interaction with a

putative protein drug target since they are not immobilized to any sort of solid matrix.

Furthermore, binding of the TP to immobilized compounds may be sterically hindered due to its orientation to the pin or bead matrix. Thus, screening assays carried out in solution, as in the preferred method of screening of the complementary library in the present invention, are preferred.

Another advantage this method provides is focusing drug leads to sites on the TP which are biologically relevant. Assays which only require the compound binding to the target do not select for compounds which affect the activity of the TP, e.g., an interaction (e.g., enzymatic) of the TP with a biological ligand (for example another protein).

In another aspect, the present invention is used to identify the biological activity of a target protein whose biological function is not known and perhaps cannot be determined directly.

In this method, compounds of known (or determinable) biological activity are screened for their ability to inhibit the binding of a peptide which binds the target protein. If such a compound inhibits such binding, it is hypothesized that the target protein mediates one or more of the biological activities of the compound.

The complementary library need not be a combinatorial library, provided that it is a library of substantial (e.g., 100 compound) structural diversity, e.g., a library of isolated natural products from various plant or animal sources, or a library of analogues previously made in various drug development programs.

Target Protein

The target protein may be a naturally occurring protein, or a subunit or domain thereof, from any natural source, including a virus, a microorganism (including bacterial, fungi, algae, and protozoa), an invertebrate (including insects and worms), or the normal or cancerous cells of a vertebrate (especially a mammal, bird or fish and, among mammals, particularly humans, apes, monkeys, cows, pigs, goats, llamas,

sheep, rats, mice, rabbits, guinea pigs, cats and dogs). Alternatively, the target protein may be a mutant of a natural protein. Mutations may be introduced to facilitate the labeling or immobilization of the target protein, or to alter its biological activity (An inhibitor of a mutant protein may be useful to selectively inhibit an undesired activity of the mutant protein and leave other activities substantially intact).

The target protein may be, inter alia, a glyco-, lipo-, phospho-, or metalloprotein. It may be a nuclear, cytoplasmic, membrane, or secreted protein. It may, but need not, be an enzyme. The known binding partners (if any) of the target protein may be, inter alia, other proteins, oligo- or polypeptides, nucleic acids, carbohydrates, lipids, or small organic or inorganic molecules or ions. The biological activity or function of the target protein may be, but is not limited to, being a

kinase

protein kinase
 tyrosine kinase
 Threonine kinase
 Serine Kinase
 nucleotide kinase
 polynucleotide kinase

Phosphatase

Protein phosphatase
 nucleotide phosphatase
 acid phosphatase
 alkaline phosphatase
 pyrophosphatase

deaminase

protease

endoprotease
 exoprotease
 metalloprotease
 serine endopeptidase
 cysteine endopeptidase

nuclease

Deoxyribonuclease
 ribonuclease
 endonulcease
 exonuclease

REF ID: A55050

polymerase

- DNA Dependent RNA polymerase
- DNA Dependent DNA polymerase
- telomerase
- 5 primase

Helicase**Dehydrogenase****transferase**

- 10 peptidyl transferase
- transaminase
- glycosyltransferase
- ribosyltransferase
- acetyltransferase

Hydrolase

- 15 urease

carboxylase**isomerase**

- 20 dismutase
- rotase
- topoisomerase

glycosidase

- endoglycosidase
- exoglycosidase

- 25 deaminase

lipase**esterase****sulfatase****cellulase**

- 30 lyase

reductase**synthetase****Ion Channel****DNA Binding**

- 35 RNA Binding

Ligase

- RNA ligase
- DNA ligase

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F epsilon Receptor

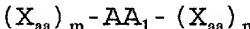
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different by more than 2; most preferably m and n are equal. Even if the chosen AA_1 is required (or at least permissive) of the TP binding activity, one may need particular flanking residues to assure that it is properly positioned. If AA_1 is more or less centrally located, the library presents numerous alternative choices for the flanking residues. If AA_1 is at an end, this flexibility is diminished.

The most preferred libraries are those in which AA_1 is tryptophan, proline or tyrosine. Second most preferred are those in which AA_1 is phenylalanine, histidine, arginine, aspartate, leucine or isoleucine. Third most preferred are those in which AA_1 is asparagine, serine, alanine or methionine. The least preferred choices are cysteine and glycine. These preferences are based on evaluation of the results of screening random peptide libraries for binding to many different TPs.

The effect of fixing one position in a library is to increase the occurrence of that particular residue from 1 in 20 to 20 in 20, an increase of 20 fold. Thus in theory if a particular residue is required for binding in the middle of the peptide, the rate of finding clones would be 20 fold higher than if a random residue were used. Therefore by using 20 libraries with one fixed residue the chances of finding members that bind to the target protein would be increased $[20 \times (\# \text{ of residues conserved for binding})]$ when compared to using completely random libraries. These 20 libraries (or at least a subset of them) would be effective against any target and no prior knowledge of the sequence for the peptide ligand would be required.

Ligands that bind to functional domains tend to have both constant as well as unique features. Therefore, by using "biased" peptide libraries, one can ease the burden of finding ligands.

For example, HPQ occurs in most streptavidin-binding peptides, which bind with the HPQ side chains oriented inward so as to interact with the biotin-binding site of the TP streptavidin. Some of the residues that participate in binding biotin also interact with the peptides; however, the peptides

adopt an alternate method of utilizing binding determinants (Biochemistry 31: 9350-4 (1992) [93003082], Crystal structure and ligand-binding studies of a screened peptide complexed with streptavidin, P. C. Weber, M. W. Pantoliano & L. D. Thompson).

5 Therefore, if one starts off with a biased library e.g. X(6)-H-X(6), then one finds many binding peptides in a short period of time because that library will be rich in peptides having the cognate binding site.

The example above showed a biased library with one residue
10 held constant. The net effect of this is to increase the number of peptides with the constant residue in that position. If this residue at this position is helpful for binding, then the number of individuals per library that will bind to the target protein will be increased. If all the amino acids are
15 represented equally, then the number of potential binding peptides is increased 20 fold in a library made up of the 20 naturally occurring amino acids. Libraries using different ratios of amino acids will be enriched according to the proportion of each residue in the starting library.

20 Of course, if the library is biased with a constant residue which happens to disrupt binding, the screening results will be negative. Therefore, it may be advantageous to screen a plurality of different biased peptide libraries in parallel. One could have a constant Trp, another, a constant Glu, etc.

25 If two residues were held constant and both were required for binding, then the incidence of binders would be increased by a much larger amount. The incidence of occurrence is independent at each position, therefore holding two residues constant is multiplicative: in a simple case of equal
30 representation, 20 fold for each site or 400 fold overall. Evidence supporting this was found in the use of a two residue biased library to enrich for peptides which bind to src homology 3 domains (SH3) (Proc. Natl. Acad. Sci. USA. 93:1540-1544 (1996) Distinct ligand preferences of Src homology 3
35 domains from Src, Yes, Abl, Cortactin, p53bp2, PLCgamma, Crk, and Grb2. A. Sparks, J. Rider, N. Hoffman, D. Fowlkes, L. Quilliam, and B. Kay). The authors found an increase in the titers of SH3-binding phage approximately 100 fold over random

libraries of the same size and complexity. This is close to the theoretical increase for these libraries ((2 codons for P divided by 31 possible codons)² = 240 fold increase).

The use of libraries biased at two positions known to be required for binding is an extremely powerful tool. However, to make parallel biased libraries which collectively include all eleven amino acid peptides, with, in each individual biased library, two constant residues, would require 44,000 libraries. (11 positions for fixed residue 1 x 20 amino acids x 10 positions for fixed residue 2 x 20 amino acids). Even if one of the constant residues were always the middle residue, there would be 4,000 libraries. While screening this number of libraries may be possible, the increase in the number of binding peptides would probably not justify the complexities of the task. However, if the number of libraries was limited to less than 100, these could be screened with relative ease.

The task can be reduced to this practical level if the following assumptions and approaches are used. First, some residues have similar functional groups and are often interchangeable for peptide target interactions. In addition, due to the degeneracy in the genetic code, many amino acids can be encoded by allowing one base in a codon to be degenerate. Amino acids can be grouped and coded for in the following manner:

25	1	W	TGG
	2	F, Y	T(T,A)T
	3	P	CCT
	4	H	CAT
	5	D, E	GAX
30	6	K, R	A(G,A)A
	7	N	AAT
	8	Q	CAA
	9	L, I, V	(G,A,C)TT
	10	M	ATG
35	11	S, T	A(G,C)T
	12	A, G	G(G,C)T
	13	C	TGT

This reduces the number of variables to 13, however this would still require 1690 (13 x 13 x 10) libraries to represent all the possible combinations in a 11 mer with the central residue held constant and one additional constant residue scanning the remaining 10 positions. This number can be reduced

10 fold by constructing the libraries using mixed oligonucleotides in the following way. In this example a central residue is held constant as a W (encoded by TTG), and D, E (encoded by GAX) are scanned through the remaining 10 positions. To construct the library, 10 oligonucleotides are synthesized:

	1	GAX	NNK	NNK	NNK	NNK	TGG	NNK	NNK	NNK	NNK	NNK
	2	NNK	GAX	NNK	NNK	NNK	TGG	NNK	NNK	NNK	NNK	NNK
	3	NNK	NNK	GAX	NNK	NNK	TGG	NNK	NNK	NNK	NNK	NNK
10	4	NNK	NNK	NNK	GAX	NNK	TGG	NNK	NNK	NNK	NNK	NNK
	5	NNK	NNK	NNK	NNK	GAX	TGG	NNK	NNK	NNK	NNK	NNK
	6	NNK	NNK	NNK	NNK	NNK	TGG	GAX	NNK	NNK	NNK	NNK
	7	NNK	NNK	NNK	NNK	NNK	TGG	NNK	GAX	NNK	NNK	NNK
	8	NNK	NNK	NNK	NNK	NNK	TGG	NNK	NNK	GAX	NNK	NNK
15	9	NNK	NNK	NNK	NNK	NNK	TGG	NNK	NNK	NNK	GAX	NNK
	10	NNK	NNK	NNK	NNK	NNK	TGG	NNK	NNK	NNK	NNK	GAX

These are converted to double stranded oligonucleotides, mixed together and cloned as a group into a phage display vector by conventional methods. The end result is a library that displays peptides with a central W residue and enriched for either D or E in each of the 10 flanking positions. The enrichment in the flanking residues occurs only once per peptide, thereby allowing the rest of the peptide to be random. Thus if the enriched residues (in this case D or E) increase binding in position 1 but are deleterious for binding in other positions, these libraries would still code for a higher number of peptides with the ability to bind to the target protein. This is an advantage over constructing libraries from triplet codon subunits and varying the ratio of each codon represented at each position and constructing the library from a single codon.

One additional complexity added to this calculation is the use of an NNK coding scheme to represent all 20 amino acids. Residues L, R, and S are overrepresented 3 fold in this scheme while V, T, A, G, and P are overrepresented 2 fold. These residues are present in higher amounts using the NNK coding scheme and the increase in the number of surrogate ligands isolated by enriching for these residues will be less than those residues that are coded for only once. The degree of overrepresentation/underrepresentation may be reduced by using

non-equimolar mixtures of bases at each position. This problem could be avoided altogether if the nucleic acid were synthesized triplet by triplet, rather than base by base, so at each step one added one out of 20 possible trinucleotides, each encoding a different amino acid.

It is desirable to enrich for residues that are important for protein-peptide interactions. These residues contain side chains that can interact with other amino acids and are less likely to pack tightly, allowing a greater degree of freedom for interaction with other ligands. A study of residues at protein binding sites showed an overrepresentation of R, H, W, and Y (Villar and Kauvar, FEBS Letters 349: 125-130 (1994) Amino acid preferences at protein binding sites). A compilation of peptide sequences derived from the phage display against a series of proteins reveals that the amino acids are not found in equal amounts, that is to say that some amino acids appear in peptides that bind to various targets more frequently than other amino acids. A graph which shows the raw incidence of residue occurrence in peptides binding to any of 16 proteins is shown in figure 3; figure 4 shows the effect of correcting for codon usage. There is a clear overrepresentation of aromatic residues, proline, cysteine and aspartic acid. Biased libraries with these residues fixed or scanning through the displayed peptide are preferred, whereas biased libraries with residues that are under represented (such as alanine, methionine, and lysine) are less preferred with libraries containing the remaining residues fixed or scanning of intermediate usefulness. As new peptides are described for additional targets this data set should be updated and reevaluated, however, the trends are quite clear.

An empirical way of determining which residues are preferred would be to take a representative mixture of proteins and bind to them a random synthetic peptide library. After washing away the peptides that did not bind, the remaining peptides could be eluted and the molar ratio of residues remaining bound could be determined. The profile should tell which residues result in peptides which would bind to the original mixture of proteins. This approach would also work

on an individual target, providing initial information on residues important for binding. An alternative method for determining which residues are preferred would be to take the mixture of proteins and use a set of phage display libraries in which one residue of the displayed peptide is fixed to select for binding phage. After several rounds of affinity selection, the libraries with the greatest number of binding phage should be those where the fixed residue is contributing to the binding of the displayed peptides.

Using the information from above, the number of libraries can be reduced greatly if the central residue that is held constant is from group 1 through 7 and 13. The C residue is a special case where the C held constant will be placed at the end of the peptide, not in the middle. However, there are special uses for libraries having cysteine constrained loops (see examples 1, 2, and 4) and this method would be especially useful to increase the number of binding peptides from these libraries. These 8 constant residues can be combined with residues from groups 1-7, giving a structured panel of 56 libraries, a practical number to construct and manipulate.

While certain synthetic strategies have been discussed above, the present invention is not limited to any particular method of synthesizing a combinatorial peptide library with one or more predetermined positions held constant, or with a particular mixture of amino acids at a given position.

Another way of thinking about the biased libraries is that they represent a more efficient way of generating complexity. If the structural properties of the binding site on the TP is such that only PLs with a certain residue, say W can bind, then a library with a single fixed, centrally positioned W will have more potential binding motifs than will a totally random library with the same numbers of unique members. However, it is readily apparent that using a library with a fixed W will in fact be a hindrance if the TPs binding site does not (and especially if they cannot) bind PLs with a W.

Therefore, it is important to take note of any apparent motifs shared in common between the diverse PLs identified

which bind to a specific TP. This has two purposes. Firstly, if the TP has more than one functional domain (FD), then it may bind to two distinct populations of PLs. It is desirable to discern this before actual setting up the drug screens themselves. If there is more than a single FD in a TP, then the likelihood of finding a suitable drug lead by creating and carrying out screens for all FDs is increased. The more domains on a target for a potential drug to interact, the more likely one is of identifying compounds that act on that particular TP. However, there are no assurances that one can detect multiple FDs if they actually exist. For example, a TP might have two very similar FDs (such as two SH3 domains within the adaptor protein Grb-2) and thus the PLs may be of a single class even though the TP has two FDs.

Secondly, the chemical nature of the residue side chains composing the motifs provide very useful information about the nature of the LP/FD interaction. This information is quite helpful in directing one's attention to particular compound libraries, or certain subsets of compound libraries for the screening process. Each amino acid has characteristic side chains, with particular characteristics of size, charge and hydrophilicity. If particular amino acids are favored at certain positions in the peptide library it suggests what substituents might be favored in the compound libraries. Some examples are given below:

<u>Residues</u>	<u>Substituents to consider</u>
Ser, Thr	hydroxylated species, especially those aliphatic in nature. Also thiolated species.
Asp, Glu	carboxylated species, especially those aliphatic in nature. Esters and alcohols.
Asn, Gln	carboxylated species; amides.

	Lys, Arg	aminated species; derivatives of the delta-guanido group.
	His	aromatic compounds with heterocyclic nitrogen especially imidazole derivatives.
5	Phe	aromatic compounds, substituted or not.
	Tyr	as for Phe, but also hydroxylated species.
10	Trp	aromatic structures, especially with two or three fused rings; the rings may include heterocyclic nitrogen; indole derivatives are of particular interest.
	Cys	thiolated and hydroxylated species.
	Met	thiolated species; sulfonium salts; sulfoxides; sulfones.
	Ala, Val,	aliphatic hydrocarbons
15	Leu, Ile	
	Pro	pyrrolidine derivatives.

It must be emphasized that these are suggestions, not requirements.

Giving priority to the libraries likely to provide the most "hits" is time and cost effective. The process is akin to "rational drug design" but is probably better called "rational library selection" herein.

If more than one peptide ligand is identified, they can be tested against each other. Competitive inhibition implies that they bind to the same site; its absence, that they bind

to different sites. If the functional domains of the TP are known, these domains can be used separately as screening targets to identify which ligands bind which domains.

A peptide library may be prepared by biological or nonbiological synthesis methods. In a biological synthesis method, a gene encoding the peptides of interest is expressed in a host cell so that the peptides are displayed either on the surface of the cell or on the outer coat of phage produced by the cell. Of course, to achieve diversity, the gene must be randomized at those codons corresponding to variable residues of the peptide. It thus is not a single DNA, but rather a DNA mixture, which is introduced into the host cell culture, so that each cell has the potential, depending on which DNA it receives, of expressing any of the many possible peptide sequences of the library. (On average, each cell will express only one of the sequences of the mixture.) The gene may be randomized by, in the course of synthesis, using a mixture of nucleotides rather than a pure nucleotide during appropriate synthetic cycles. The synthesis cycles may add one base at a time, or an entire codon.

The peptide library may also be prepared nonbiologically by stepwise addition of amino acids. During the cycles which incorporate variable residues, the activated AA is chosen randomly from an AA mixture. Preferably, the synthesis is carried out on a solid surface, such as a pin or bead. See (Proc Natl Acad Sci U S A 81: 3998-4002 (1984) [84248046], Use of peptide synthesis to probe viral antigens for epitopes to a resolution of a single amino acid, H. M. Geysen, R. H. Meloen & S. J. Barteling) or bead (Nature 354: 82-4 (1991) [92049760]) A new type of synthetic peptide library for identifying ligand-binding activity K. S. Lam, S. E. Salmon, E. M. Hersh, V. J. Hruby, W. M. Kazmierski & R. J. Knapp). If the peptide library is on a solid phase, then it is usual for the target protein to be labeled. The peptide "marked" by the labeled TP can then be sequenced.

Preferably the tags are such that only a minimal number of steps are necessary and that special handling such as that

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eluted from the TP and the whole mixture sequenced by automated techniques. Useful information is most readily achieved by using biased peptide library pools wherein one amino acid resident is "fixed", for example, x-x-x-x-Y-x-x-x-x-x. If a tyrosine (Y) is in fact contained with a binding motif for a domain contained within the TP, then other residues within the motif will be in register with the tyrosine. Thus, if cognate binding peptides must have a leucine following the tyrosine, then the sequencing of the pool will yield a sequence of x-x-x-x-Y-L-x-x-x-x. Without a *a priori* knowledge of the natural binding partner, this makes it preferable to use up to 19 (all amino acids except for cysteine) library pools. This is not really so overwhelming as the costs of the random peptide pools is not prohibitive. It is preferable to avoid cysteine as a component as peptides with an odd number of cysteines do not bind well.

In screening phage libraries, it is also routine to immobilize the TP on a solid support, since nonbinding phage can be removed. (Science 249: 404-6 (1990) [90333257], Random peptide libraries: a source of specific protein binding molecules, J. J. Devlin, L. C. Panganiban & P. E. Devlin; Science 249: 386-90 (1990) [90333256], Searching for peptide ligands with an epitope library, J. K. Scott & G. P. Smith; Gene 128: 59-65 (1993) [93285470], An M13 phage library displaying random 38-amino-acid peptides as a source of novel sequences with affinity to selected targets, B. K. Kay, N. B. Adey, Y. S. He, J. P. Manfredi, A. H. Mataragnon & D. M. Fowlkes).

The advantage of identifying peptides using the chemical synthesis-on-a-support (e.g., pins or beads) approach over peptide from phage display libraries are mostly due to the ease of identification of D-amino acid containing peptides which gives one a significant additional source of diversity. Conversely, the advantage of utilizing phage display libraries is that one can screen greater numbers of phage (perhaps 10^9 or 10^{10}) than pins (perhaps 10^4 or 10^5) or even possibly beads (perhaps 10^6).

Peptoid Libraries

Peptoids are oligomers, similar to peptides, in which the peptide bond (-NHCO-) is replaced by an analogous linkage. For example, the -NH- may be replaced by -NR-, where R is a functional group other than H. It may be, for example, an alkyl or aryl group. The -NH- may also be replaced by, e.g., an isostere, such as -O-, -S-, or -CH=CH-. The -CO- may be replaced by, e.g., -SiO-, -CS-, -SO₂-, -PO(OH)-, or -COR-.

It is not necessary that all of the linkages in a peptoid be identical; e.g., the R in each -NRCO- may be different, as an additional (or alternative) source of diversity. A peptoid may include one or more peptidyl bonds, provided that at least one of the main chain linkages between units is not a peptidyl bond.

It is likely that peptoids will enjoy advantages similar to those of peptides for screening purposes, except that peptoid libraries cannot be prepared biologically.

Compound Library

The compound library is a combinatorial library whose members are suitable for use as drugs if, indeed, they have the ability to mediate a biological activity of the target protein.

Peptides have certain disadvantages as drugs. These include susceptibility to degradation by serum proteases, and difficulty in penetrating cell membranes. Preferably, all or most of the compounds of the compound library avoid, or at least do not suffer to the same degree, one or more of the pharmaceutical disadvantages of peptides.

In designing a compound library, it is helpful to bear in mind the methods of molecular modification typically used to obtain new drugs. Three basic kinds of modification may be identified: disjunction, in which a lead drug is simplified to identify its component pharmacophoric moieties; conjunction, in which two or more known pharmacophoric moieties, which may be the same or different, are associated, covalently or noncovalently, to form a new drug; and alteration, in which one

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moiety is replaced by another which may be similar or different, but which is not in effect a disjunction or conjunction. The use of the terms "disjunction", "conjunction" and "alteration" is intended only to connote the structural relationship of the end product to the original leads, and not how the new drugs are actually synthesized, although it is possible that the two are the same.

The process of disjunction is illustrated by the evolution of neostigmine (1931) and edrophonium (1952) from physostigmine (1925). Subsequent conjunction is illustrated by demecarium (1956) and ambenonium (1956).

Alterations may modify the size, polarity, or electron distribution of an original moiety. Alterations include ring closing or opening, formation of lower or higher homologues, introduction or saturation of double bands, introduction of optically active centers, introduction, removal or replacement of bulky groups, isosteric or bioisosteric substitution, changes in the position or orientation of a group, introduction of alkylating groups, and introduction, removal or replacement of groups with a view toward inhibiting or promoting inductive (electrostatic or conjugative (resonance) effects).

Thus, the substituents may include electron acceptors and/or electron donors. Typical electron donors (+I) include -CH_3 , $\text{-CH}_2\text{R}$, -CHR_2 , -CR_3 and -COO^- . Typical electron acceptors (-I) include -NH_3^+ , -NR_3^+ , -NO_2 , -CN , -COOH , -COOR , -CHO , -COR , -COR , -F , -Cl , -Br , -OH , -OR , -SH , -SR , -CH=CH_2 , -CR=CR_2 , and -C=CH .

The substituents may also include those which increase or decrease electronic density in conjugated systems. The former (+R) groups include -CH_3 , -CR_3 , -F , -Cl , -Br , -I , -OH , -OR , -OCOR , -SH , -SR , -NH_2 , -NR_2 , and -NHCOR . The later (-R) groups include -NO_2 , -CN , -CHC , -COR , -COOH , -COOR , -CONH_2 , $\text{-SO}_2\text{R}$ and -CF_3 .

Synthetically speaking, the modifications may be achieved by a variety of unit processes, including nucleophilic and electrophilic substitution, reduction and oxidation, addition elimination, double bond cleavage, and cyclization.

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For the purpose of constructing a library, a compound, or a family of compounds, having one or more pharmacological activities (which need not be related to the known or suspected activities of the target protein), may be disjoined into two or more known or potential pharmacophoric moieties. Analogues of each of these moieties may be identified, and mixtures of these analogues reacted so as to reassemble compounds which have some similarity to the original lead compound. It is not necessary that all members of the library possess moieties analogous to all of the moieties of the lead compound.

The design of a library may be illustrated by the example of the benzodiazepines. Several benzodiazepine drugs, including chlordiazepoxide, diazepam and oxazepam, have been used on anti-anxiety drugs. Derivatives of benzodiazepines have widespread biological activities; derivatives have been reported to act not only as anxiolytics, but also as anticonvulsants, cholecystokin (CCK) receptor subtype A or B, kappa opioid receptor, platelet activating factor, and HIV transactivator Tat antagonists, and GPIIbIIIa, reverse transcriptase and ras farnesyltransferase inhibitors.

The benzodiazepine structure has been disjoined into a 2-aminobenzophenone, an amino acid, and an alkylating agent. See Bunin, et al., Proc. Nat. Acad. Sci. USA, 91:4708 (1994). Since only a few 2-aminobenzophenone derivatives are commercially available, it was later disjoined into 2-aminoarylstannane, an acid chloride, an amino acid, and an alkylating agent. Bunin, et al., Meth. Enzymol., 267:448 (1996). The arylstannane may be considered the core structure upon which the other moieties are substituted, or all four may be considered equals which are conjoined to make each library member.

The basic library synthesis plan and member structure is shown in Figure 1. The acid chloride building block introduces variability at the R^1 site. The R^2 site is introduced by the amino acid, and the R^3 site by the alkylating agent. The R^4 site is inherent in the arylstannane. Bunin, et al. generated a 1, 4-benzodiazepine library of 11,200 different derivatives

prepared from 20 acid chlorides, 35 amino acids, and 16 alkylating agents. (No diversity was introduced at R⁴; this group was used to couple the molecule to a solid phase.) According to the Available Chemicals Directory (HDL Information Systems, San Leandro CA), over 300 acid chlorides, 80 Fmoc-protected amino acids and 800 alkylating agents were available for purchase (and more, of course, could be synthesized). The particular moieties used were chosen to maximize structural dispersion, while limiting the numbers to those conveniently synthesized in the wells of a microtiter plate. In choosing between structurally similar compounds, preference was given to the least substituted compound.

The variable elements included both aliphatic and aromatic groups. Among the aliphatic groups, both acyclic and cyclic (mono- or poly-) structures, substituted or not, were tested. (While all of the acyclic groups were linear, it would have been feasible to introduce a branched aliphatic). The aromatic groups featured either single and multiple rings, fused or not, substituted or not, and with heteroatoms or not. The secondary substituents included -NH₂, -OH, -OMe, -CN, -Cl, -F, and -COOH. While not used, spacer moieties, such as -O-, -S-, -OO-, -CS-, -NH-, and -NR-, could have been incorporated.

Bunin et al. suggest that instead of using a 1, 4-benzodiazepine as a core structure, one may instead use a 1, 4-benzodiazepine-2, 5-dione structure.

As noted by Bunin et al., it is advantageous, although not necessary, to use a linkage strategy which leaves no trace of the linking functionality, as this permits construction of a more diverse library.

Other combinatorial nonoligomeric compound libraries known or suggested in the art have been based on carbamates, mercaptoacylated pyrrolidines, phenolic agents, aminimides, N-acylamino ethers (made from amino alcohols, aromatic hydroxy acids, and carboxylic acids), N-alkylamino ethers (made from aromatic hydroxy acids, amino alcohols and aldehydes) 1, 4-piperazines, and 1, 4-piperazine-6-ones.

DeWitt, et al., Proc. Nat. Acad. Sci. (USA), 90:6909-13

(1993) describes the simultaneous but separate, synthesis of 40 discrete hydantoins and 40 discrete benzodiazepines. They carry out their synthesis on a solid support (inside a gas dispersion tube), in an array format, as opposed to other
5 conventional simultaneous synthesis techniques (e.g., in a well, or on a pin). The hydantoins were synthesized by first simultaneously deprotecting and then treating each of five amino acid resins with each of eight isocyanates. The benzodiazepines were synthesized by treating each of five
10 deprotected amino acid resins with each of eight 2-amino benzophenone imines.

Chen, et al., J. Am. Chem. Soc., 116:2661-62 (1994) described the preparation of a pilot (9 member) combinatorial library of formate esters. A polymerbead-bound aldehyde
15 preparation was "split" into three aliquots, each reacted with one of three different ylide reagents. The reaction products were combined, and then divided into three new aliquots, each of which was reacted with a different Michael donor. Compound identity was found to be determinable on a single bead basis
20 by gas chromatography/mass spectroscopy analysis.

Holmes, USP 5,549,974 (1996) sets forth methodologies for the combinatorial synthesis of libraries of thiazolidinones and metathiazanones. These libraries are made by combination of amines, carbonyl compounds, and thiols under cyclization
25 conditions.

Ellman, USP 5,545,568 (1996) describes combinatorial synthesis of benzodiazepines, prostaglandins, beta-turn mimetics, and glycerol-based compounds. See also Ellman, USP 5,288,514.

30 Summerton, USP 5,506,337 (1996) discloses methods of preparing a combinatorial library formed predominantly of morpholino subunit structures.

Heterocyclic combinatorial libraries are reviewed generally in Nefzi, et al., Chem. Rev. 97:449-72 (1997).

35 Examples of candidate simple libraries which might be evaluated include derivatives of the following:

Cyclic Compounds Containing One Hetero Atom

Heteronitrogen

		pyrroles	
		pentasubstituted pyrroles	
5		pyrrolidines	
		pyrrolines	
		prolines	
		indoles	
		beta-carbolines	
10		pyridines	
		dihydropyridines	
		1,4-dihydropyridines	
		pyrido[2,3-d]pyrimidines	
		tetrahydro-3H-imidazo[4,5-c]	
15	pyridines		
		Isoquinolines	
		tetrahydroisoquinolines	
		quinolones	
		beta-lactams	
20		azabicyclo[4.3.0]nonen-8-one	amino
	acid		

Heterooxygen

		furans	
		tetrahydrofurans	
25		2,5-disubstituted	
	tetrahydrofurans		
		pyrans	
		hydroxypyranones	
		tetrahydroxypyranones	
30		gamma-butyrolactones	

Heterosulfur

sulfolenes

Cyclic Compounds with Two or More Hetero atoms

Multiple heteronitrogens

imidazoles

5

pyrazoles

piperazines

diketopiperazines

arylpiperazines

benzylpiperazines

10

benzodiazepines

1,4-benzodiazepine-2,5-diones

hydantoins

5-alkoxyhydantoins

dihydropyrimidines

15

1,3-disubstituted-5,6-

dihydropyrimidine-

2,4-diones

cyclic ureas

cyclic thioureas

20

quinazolines

chiral 3-substituted-quinazoline-

2,4-diones

triazotes

1,2,3-triazoles

25

purines

Heteronitrogen and Heterooxygen

dikelomorpholines

isoxazoles

isoxazolines

30

Heteronitrogen and Heterosulfur

thiazolidines

dihydrothiazoles

2-aminothiazoles

3-amino thiophenes

4-melathiazanones

10 For details on synthesis of libraries, see Nefzi, et al.,
Chem. Rev., 97:449-72 (1997), and references cited therein.

15 acetals

alcohols

amides

amidines

20 amines

amino acids

amino alcohols

amino ethers

amino ketenes

25 ammonium compounds

azo compounds

enols

ethers

guanidines

hydrocarbons

ketones

lactams

lactones

10 mustards

nitro compounds

nitroso compounds

organo minerals

phenones

15 quinones

semicarbazones

stilbenes

sulfonamides

sulfones

20 thiols

thioamides

thioureas

ureas

urethans

hypnotics

anticonvulsants

narcotic analgesics

analgesics, antipyretics, antirheumatics

5

10

neuroleptics

arylacetic acid (indomethacin)

15

phenothiazine (chlorpromazine)

reserpine

anxiolytics

benzodiazepines (chlordiazepoxide, diazepam, oxazepam)

antidipressants

muscle/relaxants

20

CNS stimulants

phenylalkylamines (amphetamine)

25

oxazolidinones (pemoline)

cholinergics

choline esters (acetylcholine)

N,N-dimethylcarbamates

adrenergics

5 aromatic amines (epinephrine, isoproterenol,
phenylephrine)

alicyclic amines (cyclopentamine)

aliphatic amines (methylhexaneamine)

imidazolines (naphazoline)

10 anti-adrenergics

indolethylamine alkaloids (dihydroergotamine)

imidazoles (tolazoline)

benzodioxans (piperoxan)

beta-haloalkylamines (phenoxybenzamine)

15 dibenzazepines (azapetine)

hydrazinophthalazines (hydralazine)

antihistamines

ethanolamines (diphenhydramine)

ethylenediamines (tripelennomine)

20 alkylamines (chlorpheniramine)

piperazines (cyclizine)

phenothiazines (promethazine)

local anesthetics

benzoic acid

25 esters (procaine, isobucaine, cyclomethycaine)

basic amides (dibucaïne)

anilides, toluidides, 2, 6-xylydides (lidocaine)

tertiary amides (oxetacaine)

vasodilators

polyol nitrates (nitroglycerin)

diuretics

5 xanthines

thiazides (chlorothiazide)

sulfonamides (chlorthalidone)

antihelmintics

cyanine dyes

10 antimalarials

4-aminoquinolines

8-aminoquinolines

pyrimidines

biguanides

15 acridines

dihydrotriazines

sulfonamides

sulfones

antibacterials

20 antibiotics

penicillins

cephalosporins

octahydronaphthacenes (tetracycline)

sulfonamides

25 nitrofurans

cyclic amines

naphthyridines

xlenols

antitumor

alkylating agents

5 nitrogen mustards

aziridines

methanesulfonate esters

epoxides

amino acid antagonists

10 folic acid antagonists

pyrimidine antagonists

purine antagonists

antiviral

adamantanes

15 nucleosides

thiosemicarbazones

inosines

amidines and guanidines

isoquinolines

20 benzimidazoles

piperazines

For pharmacological classes, see, e.g., Goth, Medical Pharmacology: Principles and Concepts (C.V. Mosby Co.: 8th ed. 1976); Korolkovas and Burckhalter, Essentials of Medicinal Chemistry (John Wiley & Sons, Inc.: 1976). For synthetic methods, see, e.g., Warren, Organic Synthesis: The Disconnection Approach (John Wiley & Sons, Ltd.: 1982); Fuson, Reactions of Organic Compounds (John Wiley & Sons: 1966); Payne and Payne, How to do an Organic Synthesis (Allyn and Bacon, Inc.: 1969); Greene, Protective Groups in Organic Synthesis

(Wiley-Interscience). For selection of substituents, see e.g., Hansch and Leo, Substituent Constants for Correlation Analysis in Chemistry and Biology (John Wiley & Sons: 1979).

The library is preferably synthesized so that the individual members remain identifiable so that, if a member is shown to be active, it is not necessary to analyze it. Several methods of identification have been proposed, including:

- (1) encoding, i.e., the attachment to each member of an identifier moiety which is more readily identified than the member proper. This has the disadvantage that the tag may itself influence the activity of the conjugate.
- (2) spatial addressing, i.e., each member is synthesized only at a particular coordinate on or in a matrix. This might be, for example, the location of a particular pin, or a particular well on a microtiter plate.

The present invention is not limited to any particular form of identification.

However, it is possible to simply characterize those members of the library which are found to be active, based on the characteristic spectroscopic indicia of the various building blocks.

Solid phase synthesis permits greater control over which derivatives are formed. However, the solid phase could interfere with activity. To overcome this problem, some or all of the molecules of each member could be liberated, after synthesis but before screening.

Screening of Compound Libraries

There are many suitable formats for carrying out efficient screens. The choice of format, in part, is dependent on the nature of the compound library being screened. Solid phase libraries consisting of compounds immobilized on pins or beads must be screened differently than compound libraries consisting of soluble compounds.

In general, assays for soluble compounds are composed of 1) an immobilized target and 2) a soluble, labeled ligand. The design of the assay is such that one measures the formation of PL/TP complex. Compounds that bind to the PL's cognate TP binding site (i.e. the FD) and prevent the binding of the PL and hence the formation of a PL/TP complex. Such interactions generally follow first order kinetics, that is, the amount of complex formed decreases with increasing amounts of compound. Compounds with potential utility shall display elements of "specificity. This means that useful compounds will inhibit the formation of one or only some PL/TP complexes, but not all.

It is necessary to establish assay kits which contain controls for the inhibition of the readout *per se*. For instance, if the readout of the amount of PL/TP complex formed is via an Elisa assay utilizing alkaline phosphatase, then it is necessary to be able to distinguish between a test compound's inhibition of the phosphatase versus inhibition of the PL/TP complex. NOTE: Because all screens established using this invention involve generically the same reagents i.e. PL and a protein target, there is an inherent advantage of the subject process since many functionally similar and dissimilar PL/TP complexes can be tested virtually simultaneously, so that the specificity of a given compound's activity can be determined with great easy.

In the specific instance of this invention, the target is the TP of interest and its cognate PL. Because the PL is a peptide, there are numerous means of labeling the PL. One very convenient means of labeling the peptide is to have it chemically synthesized with a biotin moiety attached to its amino or carboxy terminus. Preferable, the biotin moiety is separated from the peptide sequence identified above by the addition of one or more residues common in protein loops (reverse turns), or in interdomain spacers in natural multidomain proteins, e.g., glycine and serine residues. However, many permutations work well e.g. two glycines, G-S-G, S-G-S etc. The purpose is to have the biotin flexibly linked some distance away from the residues on the peptide that bind to the FD within the TP. Once the PL/TP complex is formed, it

can be detected by a procedure using commercially available alkaline phosphatase-conjugated streptavidin to form a tertiary complex which is then detected through the use of soluble colorimetric substrates for alkaline phosphatase. Since the amount of substrate hydrolyzed per unit of time is a direct reflection on the amount of initial PL/TP complex present one can conveniently use a microtiter plate reader with kinetic capacity to estimate the inhibition of complex formation by a given compound without the requirement for testing multiple dilutions of a compound in the initial screen.

It is desirable to pre-complex the biotinylated peptide to the streptavidin complex. This reduces the number of steps required in the assay *per se* and by increasing the valency of the PL reagent, thus increasing the apparent avidity leading to a stronger signal to noise ratio. If the affinity of the PL/TP complex is especially low, it is preferable to use complexes with greater avidity e.g. the peptide can be complexed to dextran polymers (DEX) which have been pre-labeled with biotin as described by Dwyer (Nature Biotechnology, 14:348-351, Detection of Low Affinity Interactions between Peptides and Heat Shock Proteins by Chemiluminescence of Enhanced Avidity reactions (CLEAR), Leslie D. Causey and Donard S. Dwyer). This very nicely circumvents any need to laboriously seek out PL of high affinity for the TP.

An example of the results expected are shown below. In this experiment, About 100 ul of peptides conjugated to DEX (at a molar ratio of 5:1 to 10:1, diluted to 1.0 ug/ml) are brought into contact with recombinant GST fusions of the Target Proteins which have been coated onto standard low-background "ELISA" grade microtiter plates using the standard methods for binding antibodies. After 15 minutes, 100 ul of the diluted test compounds (10 micromolar in 5% DMSO in PBS, pH 7.4) are added to wells in the same position in each plate. After 15 minutes, the wells are washed several times with PBS and the amount of complex formed evaluated using colorimetric reagents. In the hypothetical examples below, numerical output from the microtiter plate reader has been scored as, full binding (+++), less binding (++) , some binding (+) and no detectable binding

(-).

Table I. Peptide Ligand One/Protein Target One tested against 18 compounds

	A1: +++	A2: +++	A3: +++	A4: +++	A5: +++
5	B1: +++	B2:+++	B3: -	B4: +++	B5: +++
	C1: +++	C2: -	C3: +++	C4: +++	C5: +++
	D1: +++	D2: +++	D3: +++	D4: +++	D5: -
	+++: Complete complex formed. Wells A1 and D5 contain compound diluent but no test compound. No PL ligand is added to D5, thus any signal generated here is the result of non-specific background and is subtracted from the values for all wells.				
10					

Table II. Peptide Ligand Two/Protein Target Two tested against 18 compounds

15	A1: +++	A2: +++	A3: +++	A4: +++	A5: +++
	B1: +++	B2:+++	B3: +++	B4: +++	B5: +++
	C1: +++	C2: -	C3: +++	C4: -	C5: +++
	D1: +++	D2: +++	D3: +++	D4: +++	D5: -
	+++: Complete complex formed. Wells A1 and D5 contain compound diluent but no test compound. No PL ligand is added to D5, thus any signal generated here is the result of non-specific background and is subtracted from the values for all wells.				
20					

Table III. Peptide Ligand Three/Protein Target Three tested against 18 compounds

25	A1: +++	A2: +++	A3: +++	A4: +++	A5: +++
	B1: +++	B2:+++	B3: +++	B4: +++	B5: +++
	C1: +++	C2: -	C3: +++	C4: +++	C5: +++
	D1: +++	D2: +++	D3: +++	D4: +++	D5: -
	+++: Complete complex formed. Wells A1 and D5 contain compound diluent but no test compound. No PL ligand is added to D5, thus any signal generated here is the result of non-specific background and is subtracted from the values for all wells.				
30					

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wells.

Interpretation of Results: Since the compound added to well C2 prevented complex formation and/or blocked phosphatase label in all cases, it appears that it has non-specific effects. The compound added to wells B3 specifically blocks the formation of PL One /TP One complex; therefore, compound B3 appears to be a specific inhibitor of the binding site in the FD on TP One responsible for binding to PL One. Similarly, the compound in wells C4 specifically blocks the binding of PL Two to its cognate binding site in TP Two. In this assay, no compounds were observed to specifically block the interaction between PL Three and PT Three.

Compounds that interact with FDs on PTs can be identified directly by testing for direct PT binding to compound immobilized on a pin or bead. However, this is not advantageous. Firstly, it does not work well for "sticky" proteins. If the protein of interest binds to substrata e.g. plastic or amide resin with a high background, then it is difficult to detect specific binding. More importantly, because the compounds are synthesized on the pins or beads at a rather high density, then the protein target may bind due to a very high apparent avidity between the compound and the PT, thus the signal background is too high for reliable detection of suitable lead compounds. Finally, there is no convenient means to estimate affinity between the immobilized compound and the PT.

One can significantly improve upon direct binding assays, by using a labeled (to detect binding to the bead or pin) TP and test the specificity and affinity of the interaction of the TP and a given compound by specific inhibition by the appropriate PL. Here too, it is desirable to complex the peptide with dextran (average molecular weight ca. 6,000-8000) (about 10 peptides/mol). The dextran/PL complex is premixed with the labeled PT which are together brought into contact with the immobilized compound. The presence of the dextran *per se* minimizes the formation of non-specific complexes with the compound's plastic support and the dextran/PL complex modulates

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specific, high avidity binding of the labeled TP to the immobilized compound.

While compounds may be synthesized and evaluated immobilized to a matrix, their ultimate use will be in solution. Thus, having prepared PL cognates for the PT, one can evaluate putative inhibitory compounds for selectivity and specificity as described in the first example above.

Compounds can also be evaluated after synthesis on sheets of matrix e.g. derivatized plastic, nylon or nitro-cellulose sheets using pre-labeled TP in a mixture of dextran/PL complex. The benefits to this method are that it lends itself to miniaturization and simple record keeping by having many samples on a single sheet. Once putative inhibitory compounds are identified, they can be synthesized in larger quantities for solution based studies such as those described above.

REF ID: A505060

EXAMPLES

The following examples are given to further illustrate, but not limit, the invention.

Reference Example A

5 **The Effect of Single Residue Biased Phage Display Libraries
on the Selection of Binding Peptides**

Introduction

Phage Libraries have been used as a means of displaying large collections of peptides while at the same time carrying the genetic information which encode them. This is a powerful tool for the discovery of peptide ligands for various molecules, however, it does have some limitations. At the present time it is possible to make libraries with a complexity on the order of $1-5 \times 10^9$ clones. It is technically feasible to make libraries with random stretches of 25 residues, however, in order to represent all possible combinations of amino acids, the length of the displayed peptide is limited to about 7 amino acids (20^7 or 1.28×10^9 possibilities). This may be sufficient for some interactions, however, some protein:peptide interactions may require more than a 7 amino acid peptide to accurately imitate the interaction of two proteins. We are thus forced to choose between using a library which represents all possible amino acid combinations with a peptide which may be sub optimal for binding or using a library which has the possibility of displaying a more optimal peptide at the cost of not having any of the binding peptides being represented.

To circumvent this problem we have taken a novel approach. The consensus sequences for ligands have at least one amino acid residue which is highly conserved. Purpose built libraries (i.e. a X-X-X-P-P-X-X-P-X-X library (Yu H, Chen JK, Feng S, Dalgarno DC, Brauer AW and Schreiber SL (1994) Cell 76:933-945)) based on consensus ligand sequences have been successfully used to isolate large numbers of phage displaying binding peptides using proteins in the same family. We theorized that libraries which have a single fixed residue

flanked by regions coding for a random peptide should work in a similar fashion for targets for which no information on peptide ligands is available. If the fixed residue is important in the formation of a ligand, the number of phage which will display a peptide with binding characteristics will be enriched. Conversely, if the fixed residue is deleterious for the formation of a binding peptide, the number of phage in this library which display binding characteristics will be reduced.

We have tested this hypothesis using the SH3 domains from Abl and Src as targets, two domains for which a body of information exists on peptides which will bind. Ligands were screened for in libraries which display:

- 1) a completely random 12 amino acid peptide (random library),
- 2) four libraries having a peptide in which one fixed amino acid is flanked on each side by a random 5 mer (biased library) and
- 3) a $X_6PXPPXPX_2$ motif which conforms to the consensus for ligands that bind SH3 domain containing proteins (Class I SH3 purpose built library).

The four different biased libraries used contained a fixed proline, arginine, aspartate or phenylalanine residue. The predicted effect of the fixed proline was to be advantageous, the arginine and aspartate residues neutral, and the phenylalanine deleterious. The results indicate that the biased proline library was much more efficient than the random library and was almost as good as the purpose built library for isolating phage that bind specifically. The biased arginine library was more efficient than the random library and the biased aspartate and phenylalanine libraries were less efficient than the random library.

Methods

Phage were panned. Briefly, Src and Abl GST fusion proteins were immobilized on ELISA plates and blocked with BSA. Approximately 2×10^{11} phage from each library was added to each

5
10

Results

15

Abl Binding phage

20

25

Table A-1: Phage isolated with Abl SH3 domain data

Library	Strong Binding >1.75 OD	Weak Binding 0.5-1.74 OD	non-specific <0.5 OD
Random X ₁₂	0	10	2
X ₅ FX ₅	0	0	12
X ₅ DX ₅	0	2	10
X ₅ RX ₅	2	4	6
X ₅ PX ₅	7	0	5
X ₆ PXPPXPX ₂ (Class I SH3)	7	3	2

Table A-2: Phage isolated with the Src SH3 domain

Library	Strong Binding >1.75 OD	Weak Binding 0.5-1.74 OD	non-specific <0.5 OD
Random X ₁₂	3	0	2
X ₅ FX ₅	1	1	3
X ₅ DX ₅	4	1	0
X ₅ RX ₅	0	0	5
X ₅ PX ₅	4	1	0
X ₆ PXPPXPX ₂ (Class I SH3)	3	0	2

Conclusions

The use of single residue biased libraries greatly enhances the isolation of phage displayed peptides which have desirable binding characteristics. In both cases, single residue biased libraries yielded a larger number of high affinity binders than the random library. This enhancement is both in the number of phage isolated and in the affinity that the phage have for the target. Furthermore, in each instance, the biased libraries were as good as the so-called purpose built library. The purpose built library was made using a *priori* knowledge of the residues necessary for ligand affinity of the SH3 domain containing target.

This technique will be extremely valuable in the isolation of peptide ligands to

Example 1**Identification of Drugs for Treatment of Human Cytomegalovirus Infections.**

Human cytomegalovirus: disease associations. Human
5 cytomegalovirus (HCMV) is a ubiquitous human pathogen (for
recent reviews see Huang and Kowalik, 1993; Britt and Alford,
1996). HCMV is highly species specific. Humans are the only
reservoir for the virus, and transmission occurs by direct or
indirect contact among individuals. HCMV infections are
10 generally asymptomatic in healthy children and adults.
However, HCMV is responsible for about 8% of mononucleosis
cases (Klemola et al., 1970; Horwitz et al., 1979) and for
transfusion disease (Reyman, 1966) in some individuals that
receive blood products contaminated with the virus. HCMV can
15 cause serious disease in unborn children and in
immunocompromised people.

HCMV is the most common congenital infection in humans
(Britt and Alford, 1996); about 40,000 infected children are
born each year in the United States. Given estimates that
20 about 10-15% of these infected infants exhibit long-term
neurological pathology, then HCMV is the leading infectious
cause of central nervous system maldevelopment in newborn
children (Fowler et al., 1992). Damage to perceptual organs
is the most common outcome of intrauterine infection, and
25 congenital HCMV infection appears to be the most common non-
genetic cause of childhood hearing loss in the United States
(Hicks et al., 1993).

HCMV disease is a common posttransplantation complication
in solid organ allograft recipients. In general, the degree
30 of immunosuppression in the allograft recipient correlates with
the probability of clinically significant HCMV disease. More
than 60% of heart, kidney and liver allograft recipients
develop active HCMV infections (reviewed in Pollard, 1988;
Britt and Alford, 1996). Solid organ allograft recipients
35 exhibit a range of clinical syndromes resulting from HCMV
infection, such as prolonged fever, leukopenia,
thrombocytopenia, atypical lymphocytosis and elevated hepatic

transaminases (Hofflin et al., 1987; Singh et al., 1988; Smyth et al., 1991) with life threatening complications that include severe infections of the gastro-intestinal tract with perforations of abdominal viscera, hepatitis, and pneumonia (reviewed in Dummer, 1990; Smyth et al., 1991). Superinfection with fungal, protozoal and bacterial pathogens occurs late in the progression of HCMV disease (Chatterjee et al., 1978; Rand et al., 1978), possibly as a result of the ability of HCMV to suppress natural killer cell activity and T cell proliferation (Schrier et al., 1986).

HCMV infection is seen in about 40-50% of allogeneic bone marrow transplant recipients, and pneumonia is the most common clinical syndrome that results (reviewed in Wingard et al., 1990). Even with the availability of gancyclovir, the mortality rate following bone marrow transplant remains at 10-20% for patients with HCMV pneumonia (Goodrich et al., 1991; Schmidt et al., 1991; Yau et al., 1991; Enright et al., 1993; Winston et al., 1993).

HCMV might be a cofactor in the pathogenesis of HIV. Epidemiological studies have suggested that HCMV infection is associated with increased risk for the development of AIDS in HIV-infected individuals (Webster et al., 1989; Webster, 1991; Webster et al., 1992). Although there is no direct evidence for such a role, there have been clear demonstrations that HCMV can influence HIV gene expression and growth in cultured cells (Barry et al., 1990; Rando et al., 1990; Biegelke et al., 1991; Koval et al., 1991; Peterson et al., 1992). In addition to its uncertain role as a cofactor in HIV progression, HCMV is a major life-threatening opportunistic infection in AIDS. Although HCMV disease in AIDS has been reported to affect almost every organ system, clinically significant HCMV infections have been reported most frequently in the central nervous system (principally retinitis which is found in 20-25% of long lived AIDS patients), the gastro-intestinal system and the lung (Britt and Alford, 1996).

Prevention and treatment of human cytomegalovirus disease. Several recent reviews have described the different approaches

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to the prevention and treatment of HCMV disease (Coen, 1992; Britt and Alford, 1996).

Passive immunoprophylaxis using HCMV-specific immunoglobulin generally has not met with success as a therapy for established HCMV disease (Britt and Alford, 1996), and active immunoprophylaxis is also problematic. Attenuated HCMV vaccines have failed to provide protection in human trials (Stern, 1984; Plotkin et al., 1989). Furthermore, the use of live virus vaccines has met with safety concerns because of limited understanding of HCMV pathogenesis and virulence determinants and concerns about inoculation of women of childbearing age with a possibly teratogenic virus (Plotkin et al., 1990). Most recently there has been interest in developing subunit vaccines using individual HCMV proteins such as gB (Plotkin et al., 1990; Spaete, 1991).

Two well studied drugs have proven useful in the treatment of HCMV disease. Ganciclovir, a congener of acyclovir, and foscarnet both exhibit potent anti-HCMV activity in cell culture assays. Ganciclovir is preferentially phosphorylated in HCMV-infected cells by a viral gene product, UL97, and its subsequent incorporation into growing DNA chains by viral DNA polymerase encourages chain termination (Frank et al., 1984; Reid et al., 1988). Mutations in the viral polymerase or in UL97 confer resistance to ganciclovir (Sullivan et al., 1993; Baldanti, et al., 1995; Hanson, et al. 1995). Foscarnet inhibits the HCMV DNA polymerase directly (Snoeck et al., 1993). Numerous studies have demonstrated the efficacy of ganciclovir and foscarnet in the control of HCMV disease in both allograft recipients and AIDS patients (reviewed in Britt and Alford, 1996), although the utility of foscarnet is limited since its chronic use often leads to renal toxicity (Chrisp and Clissold, 1991; Reusser et al., 1992). In the case of AIDS, anti-HCMV drug therapy often must be continued on a long term basis since viral replication resumes shortly after withdrawal of the drug, and, as a consequence, viral variants that are resistant to one or both drugs are becoming increasingly common (Drew et al., 1991; Balfour, 1992), underscoring the need for

additional drugs with anti-HCMV activity. Recently, a new drug with anti-HCMV activity was approved by the FDA for the treatment of retinitis in patients with AIDS. Vistide is a nucleotide analog which has performed well in clinical trials, although it can cause renal impairment.

One can envision many HCMV gene products that might be targets for the development of antiviral drugs. However, two HCMV proteins are most often cited as potential targets. The first is a proteinase encoded by the HCMV UL80 open reading frame and termed assemblin (Welch et al., 1991a and b). This proteinase functions during assembly of the virion particle. A temperature-sensitive allele of the homologous herpes simplex virus gene is defective for virus assembly at the nonpermissive temperature (Preston et al., 1983), predicting that a drug that interfered with the activity of assemblin, would block viral assembly and spread. The second target is the UL44-coded polymerase accessory protein. This protein, together with the UL54-coded polymerase, is the topic of this example.

Human cytomegalovirus DNA replication as a target for drug discovery. Currently available anti-HCMV drugs target the viral polymerase either directly or indirectly, validating HCMV DNA replication as a target for drug intervention. We propose to develop powerful screening assays for drugs that directly interfere with the processivity function of the HCMV-encoded DNA polymerase.

The approximately 230,000 base-pair HCMV genome is packaged into virions as a unit length, linear, double-stranded DNA molecule (reviewed in Mocarski, 1996). When the viral DNA enters a newly infected cell, it is believed to circularize (LaFemina and Hayward, 1983) and then replicate in the nucleus, producing huge concatemeric molecules. HCMV DNA replication requires both a *cis*-acting DNA origin of replication, termed *oriLyt* (Hamzeh et al., 1990; Anders et al., 1992; Masse et al., 1992), and a set of *trans*-acting viral proteins. Eleven *trans*-acting viral replication proteins have been identified (Pari and Anders, 1993; Pari et al., 1993) using a transient replication assay originally developed to study herpes simplex

5 Replicated reporter plasmid DNA that is resistant to Dpn I is then monitored by DNA blot assay. The eleven HCMV products required for replication of the reporter are listed in Table 101. Five of the HCMV replication genes were originally identified on the basis of their sequence homology to known
10 herpes simplex virus type 1 (HSV-1) replication proteins and on the basis of biochemical studies that confirmed predicted functions. These include a DNA polymerase, encoded by the UL54 HCMV open reading frame (Huang, 1975; Ye and Huang, 1993), and a polymerase accessory protein, UL44 (Ertl et al., 1991; Ertl
15 and Powell, 1992); a single-stranded DNA-binding protein, UL57 (Anders et al., 1986; Kemble et al., 1987; Anders and Gibson, 1988); and putative helicase/primase proteins, UL105 and UL70 (Martignetti and Barrell, 1991; Chee et al., 1990). Further, it was noted that HCMV UL102 was located on the viral
20 chromosome relative to other viral genes at a similar position as the HSV-1 UL8 gene encoding a primase associated factor (Chee et al., 1990), i.e., these genes were positional homologs. These proteins probably act directly at the replication fork to mediate HCMV DNA replication. Candidate
25 homologs of this set of proteins have been found in all herpes viruses sequenced to date (EBV: Baer et al., 1984; VZV: Davison and Scott, 1986; HSV-1: McGeoch et al., 1988; HCMV: Chee et al., 1990; HVS: Albrecht et al., 1992; HHV6:), suggesting that they might represent a characteristic set of herpesvirus
30 replication machinery proteins.

Table 101. Genes that are required for HCMV DNA replication when assayed by the transfection assay of Challberg (1986)

	HCMV Gene	Function	HSV Homolog	EBV Homolog
5	UL54	Polymerase	UL30	BALF5
	UL44	Polymerase accessory factory	UL42	BMRF1
	UL57	ssDNA-binding protein	UL29	BALF2
10	UL105	DNA helicase	UL5	BBLF4
	UL70	Primase	UL52	BSLF1
	UL101-102	Primase-associated factor	UL8	BBLF2/3
	IE1/IE2	Regulatory	none	none
	UL84	Binds IE2	none	none
15	UL36-38	Regulatory	none	none
	UL112-113	Regulatory	none	none
	IRS1	Regulatory	none	none

Modified from Pari and Anders (1993). The indicated gene products were expressed under the control of their native promoters. The five regulatory components listed in the bottom portion of the table might serve entirely to activate expression of the five components listed at the top of the table that function directly in DNA replication (Iskenderian *et al.*, 1996). Some of the genes listed in the table encode multiple polypeptides, e.g. UL112-113 encodes four polypeptides (Wright *et al.*, 1988), and it is not yet known which members of the set are important for replication function. The Epstein-Barr virus (EBV) homologs listed have also been shown to be essential for complementation of lytic EBV replication (Fixman *et al.*, 1992).

The IE1/IE2, UL36-38, and IRS1-TRS1 HCMV genes encode immediate early gene products have been shown to regulate the expression of viral and cellular genes at the level of transcription (e.g. Pizzorno et al., 1988; Cherrington and Mocarski, 1989; Depto and Stenberg, 1989; Stenberg et al., 1990; Colberg-Poley et al., 1992; Stasiak and Mocarski, 1992). The UL84 and 112-113 genes encode early viral proteins with unknown functions (Wright et al., 1988; He et al., 1992). Although no effect on transcriptional regulation has been observed, the UL84 protein has been shown to bind to the IE2 protein (Spector and Tevethia, 1994) and therefore might somehow modulate its transcriptional regulatory function. Recently, Iskenderian et al. (1996) demonstrated that the combination of IE1/IE2, UL36-38, IRS1, and UL112-113 cooperated as a group to more effectively activate expression from several HCMV promoters controlling HCMV replication proteins. Thus, these proteins as well as UL84 might serve to facilitate expression of the set of HCMV proteins that act at the replication fork.

The studies on HCMV replication failed to identify a homolog of the HSV-1 UL9 protein which is required for HSV-1 DNA replication. This is not surprising since this protein binds in a sequence-specific fashion to the HSV-1 origin of DNA replication (Elias et al., 1986; Koff and Tegtmeyer, 1988). The HSV-1 UL9 protein together with the six HSV proteins identified in Table 101 are necessary and sufficient to reconstitute HSV-1 DNA replication in the transfection assay (Challberg, 1986) described above. Thus, it is highly likely that all HCMV proteins that participate directly in viral replication, with the exception of an origin-binding protein, have been identified. If HCMV utilizes an origin-binding protein, it presumably is one of the 11 proteins identified in Table 101.

Of particular interest to this example is the HCMV UL44 protein. As mentioned above, this protein is generally referred to as the DNA polymerase accessory protein. The HCMV UL44 protein has some homology to the HSV-1 UL42 protein (data not shown). Both of these proteins exist in a 1:1 complex with

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viral DNA polymerase (UL42: Powell and Purifoy, 1977; Gallo *et al.*, 1988, Crute and Lehman, 1989; UL44: Huang, E.-S., 1975; Ertl and Powell, 1992). The HSV-1 UL42 protein has been shown to be essential for viral DNA replication by analysis of viral mutants (Johnson *et al.*, 1991; Digard *et al.*, 1993), and the HCMV UL44 protein has been shown to be essential for viral DNA replication in antisense experiments (Ripalti *et al.*, 1995). Mutations that disrupt the HSV-1 polymerase-processivity factor interaction block virus DNA replication, arguing that the interaction is required for DNA replication (Digard *et al.*, 1993a and b; Reddig *et al.*, 1994), reinforcing the conclusion that the polymerase-processivity factor interaction is essential for viral growth. Amino acids at the extreme C-terminus of the polymerase (Digard and Coen, 1990; Digard *et al.*, 1993a; Stow, 1993; Tenny *et al.*, 1993; Digard *et al.*, 1995) and two distinct regions within the HSV-1 processivity factor (Monahan *et al.*, 1993) have been shown to be important for the interaction.

Biochemical studies have established that the HSV-1 UL42 and HCMV UL44 proteins dramatically enhance the processivity of the viral polymerases by discouraging dissociation of the enzyme from the growing DNA strand's terminus (Gottlieb *et al.*, 1990; Hernandez and Lehman, 1990; Weiland *et al.*, 1994). Therefore, agents that destabilize the interaction of the processivity factor with the polymerase, interfere with the interaction of the processivity factor with DNA (the processivity factor was first identified as a DNA-binding protein), or otherwise perturb the function of the processivity factor are very likely to interfere with viral DNA replication. As a result, such agents could likely interfere with viral replication; and, therefore, would be candidate anti-viral drugs.

The goal of this example is the development of a high throughput screen for the identification of small molecule anti-HCMV drug candidates that target the viral UL44 processivity factor. We will focus on the DNA synthesis processivity factor, UL44; however, the Combinatorial Recognition system can be readily applied to other viral

What advantages are provided by using Combinatorial Recognition to identify inhibitors of HCMV replication? Assays for viral factors such as the UL44 processivity factor are difficult and expensive to automate. Presently, the action of the processivity factor is measured by running a sample for each time point on an agarose gel. High throughput assays for other viral regulatory proteins are virtually impossible to design. However, the process we describe below can provide high throughput screens for many viral proteins.

Cloning and Expression of UL44

Like most HCMV mRNAs, the mRNA encoding UL44 is not spliced. Therefore, it is possible to isolate the complete UL44 coding sequence as a functional unit from HCMV genomic DNA prepared from purified virions. We have used the following oligonucleotide primers and the PCR to amplify the coding region of UL44 from CMV DNA: 5'-CTGTGCGGATCCATGGATCGCAAGACG-3' and 5'-CTGTGCGAATTCCTAGCCGCACTTTTG-3'. The resulting 1.3 kb product was purified using a Wizard PCR Preps clean up resin, blunted with T4 DNA Polymerase (NEB), cut with BamHI repurified on an agarose gel and cloned into the vector pGex2T digested with BamHI and SmaI. Individual clones were tested for the correct insert by restriction enzyme digests and two clones were sequenced in entirety to insure the correct protein was

encoded by the clone. E. coli DH5aF' carrying the plasmid were induced with IPTG to produce the fusion protein and the product was purified by affinity chromatography using glutathione sepharose as recommended by the manufacturer (Pharmacia). The resulting protein was used as a fusion protein or was cleaved from the glutathione sepharose using the protease thrombin (which cleaves between the GST portion of the fusion and the UL44 protein) by treating 1 mg of fusion protein on beads with 50 Units of Thrombin (Pharmacia) for 2 hours at room temperature. The resulting UL44 protein was analyzed by SDS gel electrophoresis and contained two major cleavage products, one of full length and the other roughly 5 kdaltons smaller.

Phage Libraries

Phage libraries were made using published protocols (Construction of Random Peptide Libraries in Bacteriophage M13 in Phage Display of Peptides and Proteins: A Laboratory Manual. Edited by B. Kay, J. Winter and J. McCafferty. Academic Press 1996.). Briefly, oligonucleotides which encoded the random peptide with one residue fixed were converted to double stranded DNA by extending a complementary primer using Sequenase (USB). The resulting fragments were digested with XhoI and XbaI, gel purified and ligated into previously digested mBAX vector. The ligation was introduced into bacteria by ten successive electroporations and the transformed bacteria were amplified overnight. The supernatant containing phage was harvested and the phage precipitated using PEG/NaCl, resuspended in 1X PBS containing 10% glycerol and frozen at -80° C. Ten of the oligonucleotides encoded peptides with the following structure: X_3UX_3 where X is any amino acid and U is a fixed residue. The following residues were fixed, one in each library: D(GAT), F(TTC), H(CAC), K(AAA), L(CTG), M(ATG), N(AAT), P(CCG), R(CGT), and W(TGG). For convenience, the X_3DX_3 library is referred to as a "D" library, X_3FX_3 as an "F" library, etc. The oligonucleotide sequence for each was 5'-GACTGTGCCTCGAGK(NNK)_{xxx}(NNK)₅TCTAGACGTGTCAGT-3' where xxx is the codon shown above for each residue fixed. In addition, a library with 10 random residues followed by a fixed C (TGC) was

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5 The Carolina Workshop Library (CWL) is a 12-a.a.
"unbiased" (no constant residue) peptide phage library, with
each amino acid encoded by NNK.

UL44 or GSTUL44 was immobilized on microtiter plates (Costar) by incubating 1 μ g of protein in 200 μ l of 0.1 M NaHCO₃, pH 8.5 overnight at 4° C. The remaining protein binding sites on the plate were blocked by adding 150 μ l of 1% BSA in 0.1 M NaHCO₃ and incubating the plate at room temperature for 1 hour. The plate was then washed 5 times with 300 μ l of TBST (100 mM Tris-Cl, pH 7.5, 150 mM NaCl, 0.1 % tween-20). Phage libraries were then added to the wells in 200 μ l of TBST and allowed to incubate at room temperature for 5 hours. The wells were washed 5X with TBST and the phage were eluted by incubating with 200 μ l of 50 mM Glycine, pH 2.0 for 10 minutes. The eluant was removed to a tube at the pH neutralized with 200 μ l of 200 mM NaHPO₄ buffer, pH 7.0. the phage were then amplified by adding the eluted phage to 5 ml of 2XYT broth containing 1:100 dilution of an overnight culture of E. coli DH5 α F'. The cultures were grown with agitation overnight at 37° C. The next morning the bacteria were removed by centrifugation at 3000 xg for 10 minutes in a SS-34 rotor. 100 μ l of the supernatant containing the amplified phage were then used in the next round of affinity purification.

Enrichment for phage that bind to UL44 were monitored by including a non-specific phage which formed white plaques on DH5 α F' in the affinity selection process starting at round 2. Upon plating each round of selection, the ratio of blue:white plaques was monitored by plating the phage in the presence of X-gal and IPTG. Phage from our libraries appear blue and the non-specific control phage appear as white plaques. If a particular library has been enriched for phage which bind to the target, then they should be selectively retained in higher

numbers than the control phage and this is reflected in the ratio of blue to white plaques (Fig. 5). Based on this data we tested individual isolates from the D, F, N, W, X₁₀C, and CWL (Carolina Workshop Library) libraries when GSTUL44 was used as the target protein. 87 out of 96 phage bound to GSTUL44, and of these 86 bound to other GST fusion proteins while one was specific for the GSTUL44 fusion. Thus it is relatively easy to obtain phage expressing peptides which recognize the GST moiety of the fusion protein. However, we are also able to isolate phage specific for the UL44 portion of the protein, showing that we are targeting multiple domains on a single polypeptide.

To increase the number of phage isolated which bind to the UL44 portion of the fusion protein, we cleaved the fusion with thrombin and used the free UL44 as a target for our selection process. To assess the distribution of binders across all of the libraries, we tested 16 individual phage from each library. Most of the phage exhibited specific binding to UL44, however, phage from the X₁₀C library gave much stronger signals in the phage ELISAs (Fig. 6).

To determine how specific the phage were for UL44 we assessed the binding of 23 of the isolates against a variety of other targets. Two populations of phage were found: the first recognized thrombin cleaved UL44 and GSTUL44 while the second group recognized only thrombin cleaved UL44. All 23 of these phage were subjected to automated DNA sequencing analysis and the results are shown below.

Isolated from panning against GSTUL44 and thrombin cleaved UL44 on Immulon4^R plates as well as thrombin cleaved GSTUL44 on Covalink^R plates

X ₁₀ C	E	H	V	C	S	W	G	W	G	R	C
others not from X ₁₀ C:											
17	P	T	S	D	L	W	R	N	L	G	G
18	W	G	E	T	M	W	D	N	R	K	V
35 11 many clones	A	G	L	T	P	W	S	L	L	V	D
8H	D	T	G	T	W	W	H	S	Y	V	L
2A	R	A	P	L	A	D	R	L	L	E	G
2E	K	L	W	S	A	D	M	S	S	I	V
2H	F	I	V	G	N	D	Y	R	L	G	K

8B	E	G	Y	P	S	W	V	Y	M	G	M
1E	A	R	D	F	E	D	V	Q	Q	C	C

Two important points are illustrated in the sequence analysis. First, regardless of how we immobilized the target we continually isolated phage from the X₁₀C library expressing the exact same peptide. These phage have identical DNA sequences and due to the complexity of the library and the sequences which are theoretically possible, it is quite likely that we continually isolated the exact same phage. Phage expressing this peptide bind to both GSTUL44 and thrombin cleaved UL44. Second, phage in from the second group represent a diverse population which only recognize the thrombin cleaved GSTUL44. None of these peptide sequences appear related to any known proteins in Genbank or the SWISS-PROT databases.

To demonstrate that the phage isolated in this screen were specific for a biologically relevant site and that they are targeting distinct sites on the protein, we carried out competition experiments between phage which recognize the GST portion of the protein and glutathione (GSH). To map the binding site for the UL44 specific phage displaying the constrained peptide from the X₁₀C library, we competed this phage for binding with linear double stranded DNA. To show the specificity of binding for the substrates and the phage, both phage were competed with both substrates. Microtiter plates were coated with GSTUL44 protein and blocked as described above. Phage specific for either the GST portion or the UL44 portion of the fusion were then added to separate wells and at the same time various concentrations of either GSH or DNA were added. The results are shown in Figure 7. It is important to keep in mind that these phage were isolated from the same affinity selection run. This clearly shows that phage specific for GST bind at the active site of the enzyme because the binding of phage is competed in a dose dependent fashion with GSH. The concentrations of GSH used here had no effect on the binding of the UL44 specific phage to the same GSTUL44 protein. Conversely, the phage specific for UL44 are binding in the DNA binding pocket of UL44 and are competed away by the addition of DNA in a dose dependent manner. The same concentrations of

DNA have little effect on the binding of the GST specific phage to the same GSTUL44 fusion protein.

Enzyme Linked Spectrophotometric Assay (ELSA) for CMV UL44 Using Peptides derived from Phage Display

5 Immulon 4 (cat.# 011-010-3855) 96-well plates were purchased from Dynatech. Bovine serum, albumin (BSA) (A2153), Streptavidin alkaline phosphatase (SA-AP) (S2890), Tween-20 (P1379), and *p*-nitrophenyl phosphate tablets (pNPP) (N-1891, N-2770) were purchased from Sigma. Phosphate buffered saline
10 (PBS) (21600-010) was obtained from Gibco-BRL. Ultrapure glycerol (#16374) was purchased from USB. Biotinylated surrogate ligands were prepared as a 1 mM stock solution in the appropriate solvent (H₂O or 10% acetonitrile). SA-AP was prepared as a 1 mg/ml stock solution in PBS containing 10%
15 glycerol and stored in aliquots at -80 °C. Peptides corresponding to peptides displayed on the surface of binding phage (H-Ser-Gly-Ser-Gly-Glu-His-Val-Cys-Ser-Trp-Gly-Trp-Gly-Arg-Cys-OH and Biotin- H-Ser-Gly-Ser-Gly-Glu-His-Val-Cys-Ser-Trp-Gly-Trp-Gly-Arg-Cys-OH) (the underlined residues were from
20 the X₁₀C peptide listed above; the Ser-Gly-Ser-Gly unit is a linker) were synthesized by AnaSpec, Inc.

Target protein was immobilized in microtiter wells by incubating 0.5-2.0 µg per well in 100 µl of 0.1 M NaHCO₃ overnight at 4 °C. (Studies have indicated that maximal
25 protein binding can be attained with 0.5 µg of target protein per well.) The target protein was removed and the wells were blocked with 200 µl of 1% BSA prepared in 0.1 M NaHCO₃ for 1 hour at room temperature. During the 1 hour blocking period, the SA-AP:surrogate ligand conjugate was prepared by mixing 2
30 µg SA-AP and 50 pmol biotinylated surrogate ligand for each well of target protein. (This corresponds to a 1:1 ratio of biotinylated peptide to biotin binding sites.) The mixture was incubated at room temperature for 15-20 minutes and then diluted with Tris-buffered saline-Tween-20 (TBST) 10 mM Tris-
35 HCl, pH 8.0, 150 mM NaCl, 0.05% Tween -20 such that there was 100 µl of conjugate per well. For lower concentrations of surrogate ligand, an unrelated biotinylated peptide was

titrated in to keep the total amount of biotinylated peptide at 50 pmol/well and total SA-AP at 2 μ g/well.

After blocking, the wells were washed once with 200 μ l TBST. The surrogate ligand:SA-AP conjugate was then added (100 μ l/well) and incubated at room temperature for 2 hours. The wells were then washed 5 times with 300 μ l TBST using a BioRad 1575 Immunowash plate washer. The assay was developed by adding 100 μ l of pNPP and incubating at room temperature for 5-10 minutes. The absorbance was read at 405 nm.

Figure 8 shows that a peptide derived from phage display binds specifically to UL44. The only significant binding is to GST-UL44 and not to streptavidin, GST or a GST-SrcSH3 fusion protein, indicating that the structure recognized by this peptide is UL44 and not the GST tag. This binding is time and dosage dependent (Figures 9 and 10). The binding is dependent on the concentration of the surrogate ligand and the concentration of target protein on the plate. Saturation of the target protein binding to the plate occurs at about 0.5 μ g of protein per well. The binding of this peptide to UL44 can be competed specifically with an identical peptide that is not biotinylated (Figure 11).

Conclusion:

We have isolated phage displaying peptides from biased peptide libraries which bind specifically to a GSTUL44 fusion protein. These phage bind to one of two domains: the GSH binding site on the GST portion of the fusion or the DNA binding site on the UL44 fusion. The phage are specific for each site and they can be competed for binding with their respective ligands. A peptide of the same sequence as the phage which binds to the DNA binding domain of UL44 also binds to UL44 specifically. This binding can be demonstrated by a competition assay between the peptide and the parent phage or by using a biotinylated peptide which is then detected using a streptavidin-alkaline phosphatase conjugate (SA-AP).

This conjugate could be used to screen through large numbers of low molecular weight compounds to find those that bind at the DNA binding site of UL44. This could be done

several ways. The first approach is using the assay shown here. First the target protein, in this case UL44 or a UL44 fusion protein would be immobilized on a solid surface (i.e. a microtiter plate). The peptide would be complexed with SA-AP and this complex added to a well containing the immobilized target at the same time that a small molecular weight compound is added. After allowing time for binding, the wells would be washed with buffer and the substrate for the conjugate (PNPP) could be added and allow for any conjugate in the well to convert the clear substrate to the colored product. The amount of color is proportional to the amount of SA-SP in the well. If a small molecular weight compound bound to the target at the same place as the surrogate ligand:SA-AP complex, then a reduced amount of color would be detected because the compound and the peptide:SA-SP complex can not occupy the same space. This would be the identification of a potential drug lead. This method could be used in an automated format to screen large numbers (from 1,000 to 1,000,000) of compounds for those which have the property of binding to a target in the same place as the surrogate ligand and preventing the labeled surrogate ligand from binding to the target. This may be done in such a way that the compounds are added first and allowed to bind before the addition of surrogate ligand or conversely the surrogate ligand may be bound to the target first and the compounds added after and the displacement of surrogate ligand monitored. The peptides here could be used in a precomplexed format (as shown above) or they may be used as monomeric ligands and then detected in a second step.

This example illustrates the use of phage display to isolate surrogate ligands targeted to a biologically relevant site on a viral protein and the utility of using this ligand in a high throughput screen of small molecules for the discovery of potential therapeutic drug leads. Similar experiments can be done with any viral protein which is a target for antiviral therapy. This would include proteins from CMV as well as other herpes viruses, but also proteins from any other virus regardless of classification or mode of propagation. All that is required is a source of the viral

protein. In this case the protein was produced by cloning and expressing the target in bacteria. The cloned protein could be expressed in any system including but not limited to bacteria, yeast, baculovirus, vaccinia virus, CHO cells (chinese hamster
 5 ovary cells), HeLa, fibroblasts, adenovirus or any other expression system in which the target protein is produced in such a way that it presents an active conformation to the potential surrogate ligands. The protein could also be made *in vitro* by transcription and translation using any of a variety
 10 of RNA polymerases in conjunction with lysates from reticulocytes, wheat germ or any other source of enzymatic machinery for the translation of RNA to protein. If the target protein were small enough or if a synthetic scheme could be devised to produce it *de novo*, the target could also be a
 15 completely synthetic molecule.

Biological Activity Testing (Prophetic)

As discussed above, peptides generally do not cross the plasma membrane of cells. Although technology is available to deliver peptides into cells with high efficiency using
 20 liposomes, electroporation, micro-injection, etc., we plan use an assay that does not depend on the intracellular delivery of peptides as our initial method to evaluate the physiological effect of the peptides that interact with the processivity factor. This *in vitro* assay was originally developed in the
 25 HSV-1 system (Hernandez and Lehman, 1990), and we will adapt and validate the assay for the HCMV replication system. The *in vitro* HSV-1 system utilizes three virus-coded proteins: the DNA polymerase (HSV UL30), the processivity factor (HSV UL42), and the single-stranded DNA-binding protein (HSV UL29). The
 30 template used for the *in vitro* replication system is single-stranded M13mp18 DNA to which an oligonucleotide primer (5'-GTTTTCCAGTCACGAC-3') is annealed. The oligonucleotide is generally used for DNA sequencing and is commercially available (New England BioLabs). Thus, the replication assay is an
 35 origin-independent primer extension reaction. The HSV-1 polymerase and single-stranded DNA-binding protein extend the primer annealed to M13 DNA, producing chains of variable length

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affinity that the polymerase-processivity factor or DNA-processivity factor interaction, but we can use a vast excess of the peptide to drive the peptide interaction to search for effects. We can also vary the order of addition of reactants to give the peptide an opportunity to interact with the processivity factor before the addition of other factors. We anticipate that some of the peptides will interfere with the essential interactions of the processivity factor; and, as a result, will inhibit processivity in the primer extension reaction. The specificity of inhibitory effects will be assayed by testing the ability of active peptides to inhibit the activity of mammalian DNA polymerase alpha (partially purified from HeLa cells by sequential chromatography on Q-Sepharose and double-stranded DNA cellulose, Owsianka et al., 1993) or the *E. coli* Klenow polymerase (commercially available) on the M13-primer complex. There is precedent for this type of assay in the HSV-1 system. Owsianka et al. (1993) assayed a series of 15-mer peptides corresponding to segments of the HSV-1 processivity factor (the opposite 'sense' to our peptides that will bind to the processivity factor), and identified one peptide that inhibited a primer extension reaction and exhibited some specificity for the viral as compared to the cellular DNA polymerase. The peptides that inhibit the HCMV replication reaction would then be used in high throughput peptide displacement screens for identification of small molecules from combinatorial libraries with the potential to interfere with processivity in phase II of this proposal.

We are confident that the *in vitro* origin-independent primer extension assay can be established to monitor processivity of the HCMV polymerase given the analogous nature of the HSV-1 and HCMV polymerase/processivity factor complexes and the reports of Ertl and Powell (1992) and Weiland et al. (1994) who used very similar systems. If, however, the recombinant proteins do not appear to be active, we would initially suspect that the 5' epitope tag is interfering with function. To address the potential problem, we would remove the epitope tags with enterokinase cleavage, and if that does not yield functional products, we would purify untagged

proteins from insect cell extracts using standard chromatographic procedures, as was done for the HSV-1 proteins (Hernandez and Lehman, 1990) and subsequently for the HCMV proteins (Ertl et al., 1991; Ertl and Powell, 1992). In the unlikely event that we are unable to produce active HCMV proteins, we will consider two alternative approaches. It has been shown in the HSV-1 system that extracts of insect cells can sponsor origin-independent replication of circular plasmid templates when the insect cells have been infected with a mixture of baculoviruses expressing polymerase, processivity factor, single-stranded DNA-binding protein and helicase/primase complex (Skaliter and Lehman, 1994). To implement this assay, we would clone the HCMV helicase/primase subunits (UL105, UL 70, UL 101-102) into baculovirus vectors, infect with the mixture of baculoviruses expressing the HCMV proteins equivalent to the HSV-1 proteins listed above, produce extracts and test their activity. Peptides could then be tested for their ability to inhibit origin-independent replication in the extracts. Alternatively, cells could be infected with a set of plasmids expressing the complete set of eleven factors needed to reconstitute HCMV oriLyf-dependent replication (Pari and Anders, 1993; Pari et al., 1993). Peptides would be introduced into cells by electroporation, which has been used to introduce functional small polypeptides into cells with high efficiency (Kashanchi et al., 1992) and tested for their ability to inhibit the reaction.

Screening small molecule, combinatorial chemical libraries for compounds that bind specific functional domains of HCMV UL44. (prophetic)

Combinatorial chemical libraries will be screened to identify compounds that block macromolecular interactions of UL44. We will screen the benzodiazepine library described above and other chemical diversity libraries.

The compound libraries will be screened by placing individual compounds in microtiter plates wells that have been coated with GST-UL44 fusions and adding the molecular probe. While one can use biotinylated peptides as probes within

screening assays, we have found that this can be made much more "user friendly" for compound screening by using an expression vector pMY to which peptides or protein modules can be fused to bacterial alkaline phosphatase (BAP) (described above). The BAP fusion protein retains the activity of both the enzyme and the peptide or domain. We will fuse a representative member of each class of UL44 peptide ligands to the BAP protein (by inserting fragments of DNA assembled from oligonucleotides) and then use the fusion protein to assemble a high-throughput screening assay. If the compound competitively inhibits the binding of the BAP-ligand fusion to the immobilized UL44 protein the well will remain clear during color development; on the other hand, if the compound fails to bind to the domain then the BAP-ligand fusion protein will bind to the immobilized target leading to the development of an intense yellow color.

Because the Ligand/BAP fusion protein also encodes the FLAG epitope, we include as a control well for each compound examined a test for any apparent non-specific activities by examining the effects of the compound upon binding the Ligand/BAP fusion to immobilized anti-FLAG mAb M1.

Once specific compounds with binding activity have been identified they will be characterized in extensive biochemical and viral growth studies. Among our goals is the identification of lead compounds that act at each of the UL44 functional domains. If we achieve this, then we shall test combinations for synergistic activity.

Subsequently the program will progress to the development of second-generation combinatorial chemical mini-libraries based upon the structure of the identified compounds. We seek to identify compounds that are active in the nanomolar range.

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Example 2: Isolation of Phage Expressing Peptides which Bind to Protein Kinase C β II.

Protein Kinases are important regulators of intracellular processes and play a central role in signal transduction. One of the most important controlling points is the regulation of protein kinase C. Its various isozymes are involved in many processes which regulate cell growth and hence it is an attractive target for an antitumor agent. To explore this possibility we have isolated surrogate ligands specific for human protein kinase C beta II.

Baculovirus produced Protein Kinase C β_{II} was obtained from L. Ballus (Sphinx Pharmaceutical Corp.) and was immobilized as described in Example 1 and affinity selection of phage was carried out as in example 1 with several slight modifications. Elution was found to be more efficient if the initial pH 2.0 glycine treatment was carried out with buffer prewarmed to 50° C and was immediately followed by an elution at pH 12 with prewarmed 100 mM ethanolamine for 10 minutes. The eluted phage was pooled, neutralized, and amplified in 1 ml of 2X YT containing E. coli DHFaF' in a 2 ml Megatiter plate in which each 2 ml well is arranged in a 96 well array.

Results:

We tested 95 individual phage (8 from each library except 7 from the CWL) and found potential binding phage in the D, F and X₁₀C libraries. Additional phage from these libraries were tested and we found 23 potential binders. We further confirmed the specificity of binding by testing the phage against 7 other proteins. A strong signal and good specificity was found on 13 clones and DNA was isolated from these and was sequenced using an automated sequencing protocols. The DNA sequences were compared and translated and an alignment and consensus of the peptides encoded by the phage are shown below.

Frequency									Sequence			
35	8	G	K	G	W	K	C	F	G	A	L	C
	2	S	T	T	F	Q	C	V	G	L	L	C
	1	A	N	G	W	E	C	I	G	Q	F	C
	1	K	P	V	W	K	C	T	G	L	F	C
	1	S	A	Q	W	Q	C	V	G	E	F	C
<hr/>												
1	Consensus				W	Phi	C	Pho	G	x	F/L	C

Phi=hydrophilic
Pho=hydrophobic

2		L	P	M	A	R	W	T	C	V	N	C
1		A	V	D	R	G	W	T	C	V	N	C
5	1	Q	I	T	P	Q	W	T	C	I	N	C
1	Consensus						W	T	C	V/I	N	C
1		G	V	C	Q	S	S	D	H	R	E	C
1		G	W	Q	E	R	F	Q	Q	E	D	R
1		E	V	P	T	T	K	V	L	W	P	S

10 The peptides fall into 3 classes. The first and largest class are peptides from the X₁₀C library and are characterized by a loop of 4 amino acids constrained by two cysteine residues. There are conserved residues both inside the loop and flanking the N terminal cysteine. The second group of peptides
15 also came from the X₁₀C library and is characterized by 2 residue loop. the conserved sequence is a hexapeptide in which the only variance between residues is a valine vs. isoleucine substitution on one of the peptides. The third group consists of three peptides with little primary sequence conservation.

20 To place the different clones in a relative order based on the affinity they have for the target, we carried out a set of phage ELISA with decreasing concentrations of phage. The results are shown in Figure 12. The phage exhibit a fairly wide range of binding affinities, needing anywhere from 0.5 μ l to
25 10 μ l of phage supernatant to exhibit half maximal binding.

 This example uses a eukaryotic cellular protein kinase as a target for which we have isolated artificial ligands. The peptide sequences shown above could easily be used to set up a screen for small molecules which bind at the same site. The
30 artificial ligand could be used in any of the ways discussed in example 1. We could also use any other cellular enzyme as a target. These selections may also be done in the presence of one or more cofactors or regulators of the enzymes function. In the case of PKC, we could have carried out the selection in
35 the presence of diacylglycerol or phorbol esters to activate the enzyme. This would result in the enzyme taking on a different conformation and may alter the ligands that are obtained. This

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15 This approach could also be used if a compound was available which one wished to find alternatives to. In this case, the phage could be eluted with the compound at each round of selection or the target could be blocked by the compound before phage from the final round of selection were allowed to
20 bind. Both of these approaches would give rise to artificial ligands which bound to the site blocked by the compound. These ligands could then be used in a high throughput screen to find additional compounds which bind to the same or overlapping sites.

**Example 3. Isolation of Peptides which bind to human MDM2.
Subcloning of human MDM2 cDNA into GST expression vector.**

The MDM2 (mouse double minute protein) is involved in the control of cell growth by interacting with the protein product of the tumor suppressor gene p53. In normal cells p53 acts as a sensor of DNA damage and uncontrolled cellular proliferation and prevents cell growth by activating a number of gene products which cause either cell cycle arrest or programmed cell death (apoptosis). MDM2 interacts with the N terminus of p53 and prevents it from activating these genes. Thus overexpression of MDM2 leads to uncontrolled cell growth. One potential place for pharmacological intervention in this system is the disruption of the MDM2 - p53 interaction. We set out to isolate surrogate ligands of the p53 - MDM2 interaction using phage display with the goal of using these surrogate ligands to format a screen designed to find small molecule inhibitors of this interaction.

In order to produce a fusion protein for affinity selection we subcloned the MDM2 cDNA from pQE11-hMDS by digestion with BamHI and EcoRV and ligating the cDNA into pGEX5X-1 (Pharmacia) cut with BamHI and SmaI to produce pGSThMDM2. The deletion construct expressing GST fused to the first 139 amino acids of MDM2 was made by digesting pGSThMDM2 with BsrGI and XhoI, blunting the ends with Klenow and religating. All clones were verified by restriction enzyme mapping and DNA sequence analysis. The fusion proteins were produced according to standard protocols from the manufacturer and were used as is or cleaved with thrombin.

Affinity Selection for phage displaying MDM2 binding peptides.

Selection for phage used the same techniques used in examples 1 and 2 except that enrichment was monitored by performing phage ELISAs on the pools of phage isolated after rounds 2 and 3 using dilutions of phage equivalent to 1 to 100 μ l of the phage supernatant in a 200 μ l binding reaction.

Enrichment for binders monitored by pool ELISA. After 3 rounds of selection, the only libraries which showed an enrichment for binding phage were the H and W libraries. 95 clones were tested,

48 from the H library and 47 from the W library. Strong binding was observed on 5 from the H library and 28 from the W library. Testing the specificity of these 33 phage against a variety of proteins showed that 17 of them bound to give strong signals and were very specific. The DNA sequences for the displayed phage were determined and are shown below:

10 11 clones PFQDYWEELLN
 PFHSWWQDLTD
 NFWDEWQTFMD
 SFTDYWRDLEQ

 Consensus xFxDyWqdLxx

 MEEPQSDPSVEPPLSQETFSDLWKLLPENNVL human p53
 MTAMEESQSDISLELPLSQETFSGLWKLLPPEDIL mouse p53
 surface contacts: * ** **

15 Other peptides which do not fit the consensus:

GAPWNWEKKEL
 ADPRLPVEREL
 MDGSGGERNSMW
 PMRTEWAVGSES

20 These peptides sequences can be placed in two groups. The first group align between themselves to form the consensus sequence FxDyWqdL where the upper case residues are completely conserved. This sequence aligns perfectly with a sequence with the human or mouse p53 protein that has been shown to interact with the
 25 N terminal portion of hMDM2 by biochemical studies and crystallography (Leng et al 1995 and Kussie et al 1996). The other peptides have limited homology to each other and do not align with peptide sequences from p53 or any other protein in genbank.

30 Conclusions:

 These experiments show the use of biased combinatorial peptide libraries to identify ligands which mimic a biological interaction, in this case a protein-protein interaction between MDM2 and p53. The peptides obtained in these experiments clearly
 35 mirror the native sequence of p53 and are identical to both the mouse and human p53 at the residues which are in contact with MDM2.

 Leng, P., D.R. Brown, C.V. Shivakumar, S. Deb and S.P. Deb. (1995). N-Terminal 130 amino acids of MDM2 are sufficient to

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Example 4. Isolation of Peptides which bind to E. coli Proline tRNA Synthetase (ProRS).

Transfer RNA synthetases catalyze the ATP dependent charging of a tRNA molecule with a specific amino acid. These charged tRNAs are then utilized in translation for the production of new proteins. These enzymes are required for growth in all organisms and are quite different from bacteria to humans. They thus represent an attractive target for antimicrobial compounds. We set out to isolate surrogate ligands to the E. coli proline synthetase and test to see if these surrogate ligands were targeted to the active site of the enzyme.

Affinity Selection for phage displaying E. coli ProRS binding peptides. Selection for phage used the same techniques used in example 3.

Enrichment for binders monitored by pool ELISA. After 3 rounds of selection, the only library which showed significant enrichment for binding phage was the X₁₀C library. We screened 95 individual clones for binding to the target and carried 34 individual clones through for screening against a variety of proteins. Sequence was obtained from 30 of these and is shown below.

We carried out a second affinity selection to determine our efficiency at isolating phage which bound to the target, the only difference from the first selection in that the time allowed for binding was overnight at 4° C. The phage were then amplified during the day and the next round of binding was carried out at night. We observed enrichment in the X₁₀C library again and also a slight enrichment in the P library. We tested individual isolates from the P and X₁₀C and analyzed 20 by sequence analysis. All 9 sequences from the X₁₀C library were the same as the most abundant clone isolated in the first screen. All of the 11 clones from the P library were identical and encoded a peptide shown below. A third selection was also carried out identical to the first and phage from the X10C and P libraries were tested, this time giving rise to several new sequences as well as several found in earlier selections:

frequency	Screen	Library	Sequence (SR linker-random
-----------	--------	---------	----------------------------

peptide-SR linker)

22+9+2	1,2,3	X ₁₀ C	S R V C A I W P D L D G C S R
6	1	X ₁₀ C	S R W C S L R P Q D E G C S R
2	1	X ₁₀ C	S R W C E L W S Q D I G C S R
5 11+6	2,3	P	S R W C E L W P E G S G C S R
1	3	X ₁₀ C	S R L C E V W P Q T A G C S R
1	3	P	S R W C D I W P D T G S C S R
1	3	P	S R L C D I M P Q T V G C S R
1	3	P	S R W C E V W P D K R W C S R
10	CONSENSUS		S R W C _{acidPho} W P P _{Phi} x x G C S R

X₁₀C TCG AGG GTG TGT GCT ATT TGG CCG GAT CTG GAT GGT TGC TCT AGA
S R V C A I W P D L D G C S R

P TCG AGG TGG TGT GAG TTG TGG CCG GAG GGT TCT GGT TGT TCT AGA
S R W C E L W P E G S G C S R

15 Each of the isolated phage display peptides constrained by
two cysteines with an intervening loop of 8 residues with
several conserved residues both inside and outside the loop. The
one clone isolated from the P library also contained a peptide
with identical constraints. The codon structure of the DNA
20 encoding the displayed peptide shows that this phage is not a
contaminant from the X₁₀C library. The fixed C residue in the
X₁₀C library uses the codon TGC, however, in the phage isolated
from the P library this C residue is encoded by a TGT codon,
clearly demonstrating that this clone did indeed arise from the
25 P library.

A peptide corresponding to the most often isolated phage
was synthesized and used in an ELSA as described in example 1.
Figure 13 shows that the peptide is specific for E. coli ProRS
and that the TP does not bind other peptides. This interaction
30 can be disrupted in a dose dependent fashion by a non-
biotinylated peptide of the same sequence (Figure 14). In
addition, the binding of this peptide to ProRS is dependent on
time and peptide concentration (Figure 15).

One question that could be answered with peptides designed
35 from these phage is whether or not they are directed to random
regions on the target protein or if they target active sites and
regions of other biological interactions. The peptides used in
the above ELSAs were added to a charging assay to assess its
affects on enzymatic activity of the target. Although the time

5

10

30

Cluster analysis of the selected peptides suggests that there is selective binding of peptides to the target protein. If peptides were capable of binding to any surface of the

protein, each of the peptides would be different and there would not be any clustering of peptide sequences. Clearly all of the peptides isolated here are binding to the same site. In addition, they are binding to the active site of the enzyme, which is likely the only available biologically active site on the protein. Therefore the peptides isolated by this process are targeting biologically relevant sites on the target protein.

A fourth and intriguing point is illustrated by virtue of isolating only disulfide containing peptides. This particular protein was from *E. coli*. Inside of prokaryotic cells, the environment is such that disulfide bonds are reduced. For the peptide libraries this means that the displayed peptides are likely to be linear inside of the cell and then circularize after being exposed on the outside of the cell. This means that the peptide would not take on the same conformation inside the cell as it is displayed on mature phage that are used in panning. If a peptide expressed inside of *E. coli* bound to an *E. coli* protein and inhibited its function and this protein were essential, then that peptide would be selected against during the process of making and propagating the phage display library (because these phage are grown on an *E. coli* host). In contrast to all of the linear libraries used in this example, the X₁₀C library must display a cyclized peptide which exhibits a different conformation inside of the cell and therefore would not bind to the protein in question, in this case the *E. coli* proline tRNA synthetase.

Thus, if one were trying to isolate peptides which bind to *E. coli* proteins, the peptides themselves will most likely have to be constrained or they will be selected against during library construction and amplification. Furthermore, using libraries with two, four or six fixed cysteines as suggested by Ladner (USP 5,223,409) would only be successful if the spacing between the cysteine residues were fortuitiously chosen. In this case the spacing required was 8 residues, but this was not known before hand. To make and screen many different libraries with all the combinations would be cumbersome. In these cases it would be advantageous to fix one cysteine and enrich for a second at each position as described in the beginning of this document. In this way the number of peptides expressing a

cysteine at each position would be increased, but each clone would have only a pair of cysteines and all other residues would be random.

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Example 5. *H. influenzae* Tyrosine tRNA synthetase

As discussed above for the proline tRNA synthetase, the tyrosyl tRNA synthetase is an attractive target for new antibiotics. We set out to isolate surrogate ligands for this synthetase and to determine where these peptides were targeted on the enzyme.

Affinity Selection for phage displaying *E. coli* TyrRS binding peptides. Selection for phage used the same techniques used in example 3.

10 Results:

After 3 rounds of selection, phage ELISAs showed enrichment for binding phage in the D, F, W, N, P, CWL, PHD7 (random 7-mer, New England Biolabs) and PHD12 (random 12-mer New England Biolabs) libraries. Individual clones from each of these libraries was tested for specific binding and the peptide displayed were deduced from the DNA sequence. The results are shown below:

# of Clones		Library	Sequence
1		CWL	L Y S W P D E Q Y E R P (TyrRS1)
1		W	F G F Y G W P D D Q Y
20	1	PHD12	M Y T W P G S P Y L Q M
	2	PHD12	M Y S W P G E H Y T V H
	2	CWL	M Y A W P D S S E L E K
	1	PHD7	M Y S W P G V
	2	PHD7	Y Y G W P S E
25	1	W	D R V Y G W P P F E E
	1	W	A Y H W P W V E S E W
	1	W	G Y S W P W P D D N A S R
	1	W	I Y S W P W P S N E N
	1	PHD7	Q Y T W P W P
30	1	P	Y S W P W P D F N E T
	1	W	A Y S W P W H D T V D (TyrRS2)
	5	W	W D G F A W P M H Q T
	3	F	W P W G G F E W P K L
35	1	D	R Y W W P D W G S R E
	1	W	L W W P E W G V Y T G
	3	D	Y F W W P D W G S S A
	1	W	D R G W W W P S W G V S R
	7	D	G Y W W P D W G S G Q
	1	P	A E Y W W P D W G F F (TyrRS3)
40	1	W	R L Q Y W W P D W G P
	4	N	M Y W W P N W G S Q E (TyrRS4)
	1	P	W L D G L P L Y H E V (TyrRS5)
	1	CWL	D T V R K D L L L E R E (TyrRS6)

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Peptides corresponding to TyrRS1 through TryRS6 (shown above) were synthesized and tested in several assays. Peptide TyrRS1 was biotinylated and was used for a standard ELISA as described above. This peptide binds to TyrRS specifically (Fig. 16), other peptides do not bind to the target TyrRS (Fig 17), and the binding is dependent on time and concentration (Figs 18-19).

		Peptide					
phage		1	2	3	4	5	6
35	1	+	+	+	+	-	-
	2	+	+	+	+	-	-
	3	-	-/+	+	+	-	-
	4	+	-	+	+	-	-
	5	-	+	+	-	-/+	-/+

6 + + + + - -/+

The clustering of sequences strongly suggests that the peptides are interacting at specific sites on the target. Is this a functionally relevant site? This question was addressed in two ways. First the peptides were added to a standard charging assay to see if they inhibited enzyme activity. Peptides from group 1 and 2 effectively inhibited enzymatic activity and accomplished this in a competitive manner with ATP and amino acid. Thus the peptides from the clusters both are targeted to the enzyme active site and are themselves effective inhibitors.

Clearly these peptides are directed to a potential area of target drug interaction. If this peptide can be displaced by a small molecular weight compound, the compound is likely to be a drug lead candidate. Effective inhibitors of this target are available for our use and they mimic an intermediate in the reaction of charging Tyr tRNA. These inhibitors are tyrosyladenylate compounds. One such inhibitor was tested for its ability to disrupt the phage:target interaction with all of the phage isolated. We determined that the inhibitor is an effective competitor for phage binding on a majority of group 1 phage.

The ability of the compound to inhibit the binding of the TyrRS1B peptide to TyrRS was determined. A standard ELSA was carried out except that various concentrations of the compound were added to surrogate ligand before adding the mixture to the immobilized target in a well of a microtiter dish. The results are shown in Figure 20. It is clear that the compound inhibits the binding of the surrogate ligand to the target in a dose dependent manner. A related compound, a prolyladenylate that inhibits another synthetase (ProRS) had no effect on the binding of the surrogate ligand.

This example clearly shows several important points. First, a majority of the peptides displayed on the surface of phage are targeted to active regions of the protein. In addition, these peptides are themselves effective inhibitors of protein function. These experiments prove the utility of using peptide based surrogate ligands to detect the binding of a small

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Example 6. Targeting of Transmembrane receptors (prophetic)

Cellular receptors which span the membrane often need to be in a membrane to take on the correct conformation for a biologically active protein. This presents a problem for conventional techniques designed to find artificial ligands targeted to the native form of the protein. One possible solution to this problem is the use of live cells to express the receptor of choice and then use the whole cell as the way to present the target to the library of artificial ligands. One system in which to do this is the oocyte from *Xenopus laevis*. I would first clone the receptor of interest into a vector from which RNA could be produced *in vitro* using bacterial or phage RNA polymerases. This RNA would then be injected into oocytes and the oocytes then incubated to allow the production of protein. The oocytes (probably 1-10 per binding reaction), now with the receptor of interest on the cell surface would be mixed with the library of artificial ligands and binding allowed to occur. The oocytes would be washed to remove the non-specific binding ligands and then the ligands would be eluted using a change in pH, salt concentration or another treatment which would break the interaction. The ligands would then be amplified and subjected to further rounds of selection.

The positive selection described above may give rise to ligands specific to the receptor of interest, however, it may be necessary to use a negative selection to remove the ligands binding to proteins on the surface of the oocyte. This would be accomplished by binding the pool of artificial ligands to an oocyte which has not been injected with any RNA and thus is presenting only native oocyte proteins on its surface. Any ligands which bind to the oocyte would be removed and the ligands remaining in the supernatant would be used for subsequent rounds of positive selection. This negative selection may be carried out before or after each round of positive selection or it may only be required once during the selection process. In any case the concept is to remove artificial ligands which are binding to oocyte proteins and leave the pool of ligands enriched for those which bind to the receptor of interest.

The enrichment for ligands binding to the receptor can be

Other expression systems are likely to work as well. These include bacteria, yeast, baculovirus, vaccinia virus, CHO cells (chinese hamster ovary cells), HeLa, fibroblasts, adenovirus or any other expression system in which the target protein is produced in such a way that it presents an active conformation to the potential surrogate ligands. The protein could also be made *in vitro* by transcription and translation using any of a variety of RNA polymerases in conjunction with lysates from reticulocytes, wheat germ or any other source of enzymatic machinery for the translation of RNA to protein. It may be advantageous to produce and isolate the protein in an environment that promotes proper protein folding. One example of this would be to include canine pancreatic microsomes in the *in vitro* transcription/translation reaction. If the target protein were small enough or if a synthetic scheme could be devised to produce it *de novo*, the target could also be a completely synthetic molecule.

Example 7. Isolation and Use of nucleic acids as surrogate ligands (prophetic)

The use of peptide based artificial ligands is a powerful approach in the paradigm of drug screening discussed here. However, there may be some targets for which it will be difficult to isolate peptide ligands. In this case it may be preferable to use a DNA or RNA based aptamer as the ligand, especially as one can work with very large (complexities of $> 10^{14}$) DNA or RNA libraries. Ligands could be isolated by several different screening methodologies (US patents 5,270,163; 5,475,096; 5,567,588; 5,595,877; and 5,637,459). For example, the starting libraries for a DNA library would have defined sequences on each end of 10 to 30 bases flanking a random core of 10 to 100 bases. Primers complementary to the defined sequences on each end would be used to amplify the library and one would have a tag (such as biotin). After amplification the double stranded DNA would be bound to a matrix (streptavidin agarose) and denatured to release ssDNA. To isolate the ligand, the target protein would be incubated with a starting library of single stranded DNA (ssDNA) and the aptamers allowed to bind. Protein:aptamer complexes would then be collected by filtration through nitrocellulose or nylon membranes which will bind protein with a very high capacity but have a low affinity for ssDNA. The unbound aptamers would be washed away through the filter with an excess of buffer leaving only the aptamers which bound to the original target protein. These aptamers would be eluted by one of several methods (pH shock, phenol extraction, SDS treatment or heat), precipitated with ethanol and then amplified by PCR to synthesize a new pool for use in the next round of selection.

This process would be repeated from once to 20 times. The number of times this would be carried out is determined by monitoring the enrichment for binders after each round or after every other round of selection. This could be accomplished in several ways. The most often used approach is to radioactively label a small percentage of the library and monitor the fraction of the library retained on the filter after each round. An alternative method is to use a primer in the amplification reaction which would allow the aptamer to be detected. Two

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Example 8. Peptides which bind to *Agrobacterium faecalis* β -glucosidase, carboxypeptidase, alcohol dehydrogenase, and *E. coli* Pro RS.

The inventors believe that a majority of the peptide surrogate ligands isolated using phage display will be directed to biologically active sites. These could be the sites of protein protein interactions, protein ligand interactions, the active sites of enzymes and the regulatory sites of enzymes. To demonstrate this phenomena on a range of proteins, we have chosen a diverse set of enzymes whose biological activity can be monitored. If the surrogate ligand binds to the target enzyme at the active site, it is likely that it will act as an effective inhibitor of enzyme activity. In addition, for many of the targets below, a number of active site directed inhibitors are available. Using these we will be able to map the site of interaction between the target and the protein. The enzymes targeted are *Agrobacterium faecalis* beta-glucosidase (this example), carboxypeptidase B from pig pancreas (Boehringer Mannheim cat #103 233) (Ex. 9), Yeast alcohol dehydrogenase (Sigma cat. # A3263) (Ex. 9), and *E. coli* ProRS (Exs. 4 and 9), glycogen phosphorylase a, and yeast hexokinase.

Affinity selections were carried out as in example 3 except that the protein was presented in several ways. At first the protein was immobilized on Immulon 4 plates (Dynex) as in previous examples, however, repeated attempts failed to isolate phage which bound specifically to the target. It was found that the target exhibited greatly reduced activity when bound to Immulon 4 plates, making it likely that it was denatured when bound to plastic. To circumvent this problem, two approaches were used, both of which utilized biotinylated protein. Protein (1 mg) was biotinylated with Sulfo NHS-LC-LC biotin (Pierce, cat. #21338) prepared fresh as a 10 mM stock solution in ddH₂O. All proteins were in phosphate buffered saline. Biotinylation reagent was added to the protein solution in a 17-fold molar excess to protein and the reaction was carried out at room temperature for 30 minutes followed by ice for 30 minutes. Biotinylated proteins were separated from the excess biotinylation reagent on a Sephadex G-50 micro-spin column (Pharmacia Biotech, Cat# 27-5335-01) according to the

manufacturers directions. Protein assays were conducted using BioRad Protein Assay reagent (cat#500-0006). All proteins were stored in 1X PBS+10% glycerol at -80° C.

For the first method of affinity purification, phage libraries were mixed with 1 µg biotinylated target protein in solution and allowed to incubate at room temperature for 4 hours. Phage:protein complexes were then captured on streptavidin paramagnetic beads (Promega, cat# Z5482) by adding beads and incubating the tubes while rotating at room temperature for 30 minutes. The complexes were then drawn to the sides of the tubes using a magnet and the beads were washed with TBST containing 5 mM biotin. Beads were washed once in the first round of panning and 3 times for each additional round. Phage were eluted and amplified using the sequential procedure described in example 3. In the second method of affinity purification, phage libraries were mixed with 1 µg biotinylated target protein that was first captured on streptavidin paramagnetic beads. After the protein was bound, the beads were washed once with TBST containing 5 mM biotin to block the remaining binding sites on streptavidin. The libraries and beads were then mixed at room temperature on a rotator for 4 hours and washed, eluted and amplified as above. Phage ELISAs were carried out by first coating a microtiter plate (Immulon4, Dynex) with 1 µg streptavidin, followed by blocking with BSA. Biotinylated protein was then added to the wells and allowed to bind to the streptavidin for 1 hour. The plates were washed with TBST and used for phage ELISAs.

Results:

The following peptides were found to be displayed on phage that bound specifically to β-glucosidase:

	Displayed Peptide Sequence	Method	Library	Frequency
35	S S Q T D W R K I F Q S L S R	beads	K	3
	S S S T D W L N V W R Q L S R	beads	N	2
	S S A T D W G R V Y S I L S R	beads/sol	R	5
40	S S A S Y A P W P I Y F A S R	beads	W	2
	S S G A F K P W P V Y S F S R	beads	W	1
	S R Q V E V F K P W P V Y S R	beads/sol	K	3
	S S S F K P W P I Y L G S S R	sol	P	1
	S S E P F S V W P I Y K H S R	sol	W	1

S S S V P F A P W P V Y A S R	beads	P	1
S S T S L P F N R W P I Y S R	beads	N	2

The peptides fall into two clusters based on homology. Different methods of immobilizing the protein produced similar results: peptides from both classes were isolated from solution or bead displayed target and identical peptides were isolated from the two methods. Peptides in the second cluster were isolated predominantly from the W and P fixed residue libraries, a finding which is in concordance with the PWP motif in the conserved sequence. Phage presenting peptides in the first group came from the N, K, and R libraries, again consistent with the consensus sequence in this cluster. The only conserved residues in this motif for which a fixed residue library was screened were D and W. In both cases, the consensus sequence extends for greater than 5 residues to the carboxy terminal side making it impossible for these libraries to encode peptides which would bind to the target.

To see if the phage displaying these peptides were directed to the active site of the enzyme, a competition between phage and conduritol, an active site directed irreversible inhibitor of β -glucosidase activity, was carried out. The target was immobilized as above and was incubated with 3 mM conduritol for 3 hours. The plate was aspirated and a standard phage ELISA was carried out. The results are shown in figure 21. The binding of phage from group 2 was inhibited by conduritol, however the binding of phage from group 1 was unaffected. Conduritol is a small molecule (MW = 162.1) and it is possible that phage from group 2 are recognizing a portion of the active site that does not overlap with the conduritol binding site. Therefore at least one of the groups display peptides which bind to the active site of the enzyme.

The following peptides displayed on the surface of phage were found to bind specifically to carboxypeptidase:

	<u>Displayed Peptide Sequence</u>	<u>Library</u>	<u>Frequency</u>
35	S R L L E V S P G W W Q M S R	P	9
	S S F R E L K P G W W S Y S R	P	1
	S S W G D Y F N W R D G L S R	N	2

The following peptides displayed on the surface of phage were found to bind specifically to alcohol dehydrogenase:

	<u>Displayed Peptide Sequence</u>	<u>Library</u>	<u>Frequency</u>
	S R Q V E V F K P W P V Y S R	K	1
5	S S S F K P W P I Y L G S S R	P	1
	S V S V G M K P S P R P	PHD12	2
	S S N Y W W Q S P V L S R H S R	CWL	1
	S S W Q G N V L L G N W I S R	L	3
	S S L L N E S R L Q W S T S R	R	1

- 10 For both of these targets there are peptides which have homology to each other and cluster as well as peptides which do not have any obvious homology to the cluster. Phage displaying these peptides are very specific to the target for which they were selected to bind. Cross reactivity ELISA assays show that they
- 15 give signals at least 10 times as stronger on the specific target vs any other targets tested.

The following peptides displayed on the surface of phage were found to bind specifically to biotinylated ProRS:

	<u>Displayed Peptide Sequence</u>	<u>Library</u>	<u>Frequency</u>
20	S R D W G F W D W G V D R S R	D	5
	S R D W G F W R L P E S M A S R	CWL	3
	S R E W H F W R D Y N P T S R	R	4
	S S E R G S G D R G E K G S R	D	1

- These sequences are markedly different from those in example 4
- 25 above where the target was immobilized directly on Immulon 4 plates. It is likely that upon binding to plastic that the proteins conformation is altered and so may present a distorted binding site. Selection for phage which bind in solution may represent peptides which bind to a more native form of the
- 30 protein and thus are a better indicator of the native conformation of the target.

Similar affinity selection procedures could be carried out using glycogen phosphorylase a and yeast hexokinase. For each of the targets in this example, enzyme activity could be

monitored in the absence and presence of phage displaying peptides or using synthetic peptides to determine if they are effective inhibitors. In addition, competitions between the phage/synthetic peptides and inhibitors or substrates could be carried out to determine if the surrogate ligands identified are binding at the active site of the target. These surrogate ligands could be used to format a competitive binding assay used to search for small molecular weight inhibitors of each of these targets as described in the above examples.

- 10 From the data presented here it is clear that some proteins are very sensitive to inactivation upon binding to plastic. The use of streptavidin coated microtiter plate or beads in conjunction with biotinylated target protein provides one alternative method for the presentation of the target protein.
- 15 Other methods of target presentation such as using an antibody to an epitope tag or a ligand:fusion protein combination, as well as others may be advantageous in the selection of phage and in the screen for small molecule inhibitors.

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Identification of Inhibitors of the B7:CD28 Complex.

There is a large and growing body of biological literature which suggests that expression of B7 on the antigen presenting cell can have a critical role of the outcome of the subsequent immune response. Expression of B7 is a potent adjuvant for the priming of T cell *in vitro*, and more importantly, *in vivo*.

Peptides as molecular probes. We propose to identify small B7-surrogate ligands for CD28 from enhanced phage displayed peptide libraries to characterize the B7 binding site on CD28 and to establish screens for low molecular weight compounds that block the B7/CD28 interaction. We have chosen to develop CD28 binding peptides for several reasons.

25 The extracellular domain of CD28 will be cloned from the CD28 cDNA by PCR. PCR amplified product (with linkers) will be cloned into the TA plasmid (Invitrogen, San Diego, CA, pCRII vector, "Original TA cloning kit"). We will subsequently insert it the pGEX2T vector (Pharmacia) to generate fusion proteins.

30 Sequences of the inserts will be confirmed by automated sequencing. Subsequently, bacteria harboring the recombinant plasmids will be induced to express the GST-CD28 extracellular domain fusion proteins. The proteins will be purified by simple affinity chromatography with glutathione-agarose (Pharmacia).

35 If we have difficulty producing the external domain as a
GST fusion, we will clone it into an Ig fusion construct as has

2. Screen a bacteriophage M13 random peptide libraries for
5 peptides that bind CD28.

10 The binding of individual isolates to the GST-CD28 fusion
proteins will be evaluated by a simple enzyme linked
immunosorbent assay (ELISA) with a goat anti-phage antibody
conjugated to horseradish peroxidase (Pharmacia).

Consensus sequences of CD28 binding peptides will be determined in a manner similar to that described for UL44 in Example 1. By comparisons of sequences of the phage from the apparent "preferred" library with those from the other libraries we should be able to obtain a definition for an optimal CD28 binding peptide ligand. These consensus sequences are anticipated to be useful in computer searches such as those using the PROSITE system and the SWISS Protein database to identify novel CD28 ligands. We anticipate that some of the CD28 binding peptides may have similarity to the CD28 binding domains within B7.1 and B7.2. Based on published observations [Peach, 1995 #9], we might expect to see some structural similarity to the GFCC'C" beta-sheet face of the IgV fold and the ABED beta-sheet face of the IgC domain in B7.1 and/or B7.2.

35 Actual K_d measurements will be determined for non-biotinylated

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4. Test peptide for agonist/antagonist activity in CD28 dependent T cell-based assays.

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We expect that antagonist peptides will result in a shift in the dose response curve towards higher stimulator cell concentrations for an equivalent T cell response. Peptides with agonist activity would be expected to shift the dose response to lower numbers of stimulator cells. We anticipate that these peptides may function either as agonist, activating CD28, or antagonists, blocking CD28 dependent responses. Any peptides that fail to show blocking activity will be tested for agonist activity using suboptimal doses of anti-CD3 to stimulate T cells. Those peptides which can act as agonists will showed enhanced responses. Anti CD28 antibody will be a control for these responses.

These data will be confirmed and refined using dose response curves of peptides. Peptides will be added to culture at a single concentration stimulators (added to give 50% maximum stimulation) at varying peptide doses. Antagonist peptides should show a diminishing response as the dose increases while agonists will show stimulation above baseline as dose increases. Partial agonists are also possible. This would inhibit at low dose, but stimulate at high doses. MHC binding peptides with this characteristic have been reported (32).

Clearly the largest stumbling block to this program would be a failure to obtain CD28 extracellular domain binding proteins. Based on our previous experience, we believe that this is highly unlikely since that portion of CD28 clearly binds other proteins (B7), even when monomeric. However, if in the unlikely event this is the case, we will make a fusion construct between the extracellular domain of B7.1 and bacterial alkaline phosphatase (BAP). If the affinity of this domain for GST-CD28 is too high (greater than 1 micromolar), we will selectively mutate the B7 sequences to lower the affinity to a range acceptable for use in compound library screens.

5. Establish a high through-put screen of combinatorial chemical ligands for the GST-CD28 fusion proteins.

Combinatorial chemical libraries will be screened to identify B7/CD28 antagonists. These compounds will be used to

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5 Once specific compounds with binding activity have been identified we will test their cross-reactivity with other functional homologues of CD28 as well as families of functional domains e.g. SH3, WW and PTB domains. We can rapidly determine specificity by testing whether the compound prevents the binding
10 of biotinylated peptide ligands to extracellular domains of CTLA4 as well as other control fusion proteins. Compounds that competitively inhibit the binding of peptide ligands to CD28's extracellular domain will be used in biological experiments (see above).

15 If there is cross-reactivity between targets (which is quite possible) then a second-generation combinatorial chemical mini-library based upon the structure of the identified compounds will be generated. It is anticipated that some members of these purpose-built mini-libraries will bind to the
20 CD28 domains with greater specificity and possibly greater affinity than to CTLA4. We seek to identify compounds that are active in the nanomolar range.

References for Example 9

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ACCEPTED MANUSCRIPT

Example 10 (Hypothetical)**Identification of Antagonists of Apoptotic Proteins.**Background

Programmed cell death, also known as apoptosis, is both a
 5 normal and abnormal cellular process. There are several examples
 of normal cell death that occur during embryo development, such
 as formation of digits in limbs, destruction of larval muscles
 during metamorphosis of the moth, sculpting of the mouth,
 removal of misconnected neurons, and removal of thymocytes that
 10 recognize self antigens. Defects in the control of cell death
 have also been described as the basis of certain forms of
 autoimmunity, escape of cancer cells from growth control, and
 immortalization of virally infected cells, to name a few. Thus,
 the molecular basis of apoptosis is considered by many
 15 scientists to be critical in understanding normal and abnormal
 growth control. If the molecules involved in cell death can be
 defined, they will serve as valuable targets with which to
 develop therapeutic drugs that may be used to block or induce
 apoptosis. These drugs may have uses in treating certain forms
 20 of autoimmune disease, fighting bacterial and viral infection,
 promoting survival of damaged neurons, and battling cancer.

Many of the molecules involved in the initiation of
 apoptosis are known. They include such proteins as tumor
 necrosis factor (TNF) receptor, nerve growth factor (NGF)
 25 receptor, Fas receptor (also known as APO-1 and CD95), a variety
 of immune cell receptors, an Rb interacting protein, a nematode
 neuronal guidance protein, and two fly proteins involved in
 embryonic axis formation. The TNF-, NGF-, and Fas-Receptors (R)
 are members of the TNF-R protein family, which consists of 12
 30 members. The extracellular domains of the family all share from
 three to six cysteine-rich domains. While the extracellular
 domains are relatively conserved, the cytoplasmic regions are
 not, with the exception of the TNF-, NGF-R, and Fas-R molecules
 which share a ~80 amino acid domain. Functional analyses (see
 35 below) have shown that this domain plays an essential role in
 the cell death signalling of these receptors and hence the
 domain has been named the death domain (DD).

A consensus sequence has been established for the DD, even though the percent amino acid identity among the different DD sequences is ~30%. An up-to-date comparison of the primary structures of 13 different DD's has been posted on the web (http://ulrec3.unil.ch/domains/dd/index.html) by Dr. Kay Kauffman. A comparison of the specific DD sequences of the TNF, NGF, and Fas receptors can be found in the alignment below. As can be seen, the degree of identity (open boxes) is low and dispersed over the entire length of the DD. Only a few residues (i.e., gly¹⁷, ile²³, asp²⁴, leu⁴², trp⁴⁵, leu⁵⁸, leu⁶², glu⁷²) are conserved (bold faced below) in all three proteins.

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NGF-R  L1NGSAGDTWRHLAGELGYQPEHIDSFTHE-----ACPVRALLA
Fas-R  A1GVMTLSQVKGFVRKNGVNEAKIDEIKNDNVQDTAEQKVQLLR
TNF-R  TDDPATLYWKEFVRRLGLSDHEIDRLELQNGRCLREAQYSMLA

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15      SWATQD--SATLDALLAALRRIQRADLVESLCSESTATSPV
      NWHQLHGKEAYDTLIKDLKKANLCTLAEKIQTILKDITS
      TWRRRTRREATLELLGRVLRDMDLLGCLEDIEEALCAPPLP

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In the case of the TNF and Fas receptors, their DD's have been shown to be critical in activating cell death. When cDNA clones encoding either receptor are introduced into various cell lines, these cells will undergo apoptosis in the presence of TNF or the Fas ligand (Nagata and Golstein, 1995). However, if the transfected receptors have mutations in their DD's, apoptosis fails to occur in the presence of the extracellular signal (Tartaglia et al., 1993). Truncations of as little as 10 amino acids from the N-terminus or three from the C-terminus of the domain are inactivating. Furthermore, when most residues (black boxes) in the TNF-R DD (above alignment) were replaced with alanines, the receptor no longer caused apoptosis in host cells. Interestingly, a naturally occurring mutation in the Fas-R has been observed: in the lpr mouse, which experiences lymphocyte proliferation and an autoimmune syndrome resembling human systemic lupus erythematosus, there is a single amino acid mutation (equivalent to a change at V238; of the human Fas-R protein sequence) in the DD of the Fas receptor (Watanabe-Fukunaga et al., 1992). [The equivalent phenotype occurs in the gld mouse, where the Fas ligand is defective (Takahashi et al., 1994).] Thus, it is clear that proper function of the TNF- and

Fas-R requires intact DD's.

Both the MORT1/FADD and TRADD proteins share an additional motif, termed the death effector domain (DED). This ~80 amino acid long domain is 18-38% identical between examples. The DED plays an essential role in cell death because if the domains are removed from FADD, cell death is blocked (Chinnaiyan et al. 1996). Conversely, if only the first DED (aa 1-117) is expressed in cells, apoptosis is triggered (Chinnaiyan et al. 1996).

10 1. Construct glutathione S-transferase (GST) fusions with cell death domains from several different protein.

For use as targets, we will prepare GST-cell death domain fusion proteins.

To verify that the peptides bind the DD's in a biologically relevant manner, we will test their sensitivity to inactivating mutations of the DD's. One negative control that we will generate will be equivalent of the mouse lpr mutation (Watanabe-Fukunaga et al., 1992) in the human Fas, TNF, and NGF receptor DD's. Other inactivating mutations can be introduced in the DD's according to those described in the literature (Tartaglia et al., 1993). GST fusions to mutant DD's will be useful as negative controls in experiments examining the specificity of peptide-DD interactions (i.e., the peptide ligands should bind poorly, or not at all, to the inactive DD's). The mutations will be engineered by replacing a pair of oligonucleotides during the assembly of the DD's for cloning (above).

2. Identify peptide sequences from a phage-displayed random peptide library that bind to the GST-cell death domain fusion proteins.

30 We will screen phage-displayed random peptide libraries with GST-DD fusion proteins by affinity selection, in a manner analogous to that set forth for UL44 in Hypothetical Example 1.

The libraries will be screened according to standard techniques (Kay et al., 1993; Adey and Kay, 1996). In brief,

Conclusions regarding motifs will be confirmed with
25 synthetic peptides. We will synthesize peptides with biotin
attached and then follow their relative binding by an enzyme
linked assay (Sparks *et al.*, 1994) and dot blots. Actual K_d
measurements will be determined for non-biotinylated peptides
by fluorescence polarization; the DD segment will be released
30 from GST-DD fusion protein by thrombin cleavage and then
fluorescentated at its N-terminus. Alternatively, if the peptide
ligands lack tryptophan, the fluorescence properties of the DD's
own tryptophans will be monitored instead during peptide-DD
complex formation. In addition, dissociation constants will be
35 quantitated on a BIAcore system, see Karlsson, *Anal. Biochem.*,
228:274-280 (1995) and Raghavan, *Structure*, 3:331-3 (1995).
Based on experience with other target molecules, peptides
isolated from phage-displayed libraries are anticipated to have

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228:274-280 (1995) and Raghavan, *Structure*, 3:331-3 (1995).
Based on experience with other target molecules, peptides
isolated from phage-displayed libraries are anticipated to have

10 μ M to 10nM dissociation constants. If desired to determine the importance of individual residues of the peptide ligands, and thereby facilitate the design of more efficient libraries for the target in question, we may prepare a set of alanine-scanned
5 variants and measure their affinities.

The synthetic peptides will be used in competition experiments as well. Soluble peptides will be added to radioactive (35 S-methionine labelled) cell lysates incubated with various GST-DD fusion proteins. A similar experiment was
10 performed with SrcSH3 peptide ligands to demonstrate that they bound in the same way as natural ligands (Sparks et al., 1994). If the peptides bind to the DD, little or no cellular protein should bind to the GST-DD fusion protein when it is recovered by chromatography over glutathione-agarose, resolved by SDS-
15 PAGE, and autoradiographed. Non-binding peptides will serve as negative controls. If the peptides fail to block the interactions, due to their low affinities relative to natural ligands, we will retest them in a multivalent format. Biotinylated peptide ligands will be complexed with streptavidin
20 to raise the affinity of the peptides to the DD's through avidity. In experiments with peptide ligands to SH3 domains, isolated from phage-displayed random peptide libraries, it was shown that raising the valency of the peptides to four is an effective means of increasing their apparent affinities.

25 3. Screen a human λ cDNA expression library by COLT to identify novel cell death domain-containing proteins.

One concern in evaluating a drug lead is its cross-reactivity. That is, a drug may yield undesirable side effects due to its interaction to an unwanted target. Given that DD's
30 share some identity, the most likely cross-reactive targets of a DD specific compound will be other DD containing proteins. At the moment, only nine DD containing proteins are known. Other DD containing proteins may be identified by the COLT technique (Sparks et al., 1996b). COLT has been used to isolate over 20
35 SH3 domain containing proteins, of which half are novel. In unpublished work, we have used COLT to identify members of the calmodulin family as well as proteins containing WW domains.

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Example 11:Isolation of Surrogate Ligands for Estrogen and Other Nuclear Receptor

Nuclear receptors are a family of ligand activated
5 transcription factors which include the receptors for steroid
and thyroid hormones, retinoids and vitamin D. The receptors
are organized into distinct domains for ligand binding,
dimerization, transactivation, and DNA binding. Ligand binding
induces conformational changes in the receptors that allow for
10 dimerization and the binding of co-activating proteins. These
co-activators, in turn, facilitate the binding of the receptors
to DNA and subsequent transcriptional activation.

The steroid receptor family of nuclear receptors is
comprised of receptors for glucocorticoids, mineralocorticoids,
15 androgens, progestins, and estrogens. Unique to the steroid and
thyroid receptor families is a group of receptors known as
orphan receptors. The genes for these receptors were cloned on
the basis of their structural similarity to previously
identified members of the steroid and thyroid receptor family.
20 Certain orphan receptors are found only in specific tissues,
while others are expressed ubiquitously. The common denominator
of the orphan receptors is that they have no known ligand.
Identifying ligands for these receptors would be of considerable
value as there is evidence that members of the group of orphan
25 receptors are important transcriptional regulators during
critical stages of development. Identification of ligands for
these orphan receptors would provide useful information for
understanding regulation of gene expression as well as provide
tools for discovering pharmacological agents. As a first step
30 towards developing a method for the identification of orphan
receptor ligands, current work is centered on a known member of
the steroid family of nuclear receptors, the estrogen receptor
 α (ER). This receptor not only provides a model for the
discovery of orphan receptor ligands, but it is also a current
35 target for drug discovery.

Estrogens exert an influence on many important
physiological functions including cell growth and
differentiation, particularly in the tissues of the female and

male reproductive tract. Estrogens have also been linked to the preservation of bone mass and protection of the cardiovascular system. Despite the positive effects of estrogen, the overexpression of the estrogen receptor (ER) in certain estrogen responsive tissues has been linked to diseases such as breast and ovarian cancer (Gallo and Kaufman, 1997). For this reason, the ER is a target for drug discovery. All of the ER therapeutic agents in use today are molecules that compete with estradiol for binding to the estrogen receptor. These compounds, called antiestrogens, are classified as either antagonists or partial agonists of estrogen receptor function. While effective, these compounds have adverse side effects such as loss of bone density and an increased risk of uterine cancer. Additionally, the cancers eventually become resistant to these agents (Katzenellenbogen, *et.al*, 1997). Because of these shortcomings in the current treatment for ER related disease, new treatment methods are being sought.

Our approach is to identify, via phage display, peptide surrogate ligands or mimics of proteins that interact with the ER. This approach is unique in that not only is the estradiol binding pocket the target for small molecules, but also any site on the ER that is involved with receptor signaling.

As mentioned above, the ER is organized into distinct domains (Figure 22). Several of these regions contain sites predicted to be useful for drug intervention of ER function. For instance, the estrogen receptor and other nuclear receptors interact with one or more steroid receptor co-activators (SRC's) via a conserved alpha-helical domain located within the AF2 region of the receptor. This interaction is ligand dependent and is believed to bridge the interaction between the receptor and transcriptional activation. Many co-activators have been identified and their interactions with several nuclear receptors have been investigated. Mutagenesis experiments have identified the sequence of the binding domain on the co-activator as an LXXLL motif where X is any amino acid. This sequence has been shown to be both necessary and sufficient for receptor binding (Heery *et.al*, 1997; Torchia *et.al*, 1997). The XX portion of this motif and the surrounding residues are believed to impart

specificity to the interaction. Disruption of this interaction would be predicted to disrupt receptor signaling by blocking transcriptional activation. Peptides that mimic this interaction will provide a tool for discovering pharmacological agents that act at the co-activator binding site on the ER. Additional sites on the ER that could serve as targets for drug intervention include the AF1 region, the dimerization domain and the DNA binding domain. Once peptides are obtained for one or more of these sites, they can be used in a competitive displacement assay to screen libraries of compounds.

Methods and Results

The ER (Panvera Corp.) was immobilized on Immulon 4 plates (Dynatech) for the phage affinity selection, after it was determined that the immobilized ER was capable of binding estradiol. Phage display was conducted on the ER, as described in example 1 above, in both the presence and the absence of the natural ligand for the ER, 17- β estradiol (100 μ M).

The sequences obtained in the absence of estradiol are shown in Table 11-1. These sequences possibly represent proteins that interact with the unliganded ER such as HSP90 (90 Kda heat shock protein). Two sequences compete with estradiol (Figure 23). These sequences may bind to the estradiol binding pocket or they may bind to sites that are masked by a change in receptor conformation following binding of estradiol. The antiestrogen 4-hydroxytamoxifen does not inhibit the binding of any of these phage (Figure 23).

Sequences obtained in the presence of estradiol are shown in Tables 11-2 and 11-3. The sequences are overall leucine rich and the majority of them contain the LXXLL motif, which is found in the nuclear receptor co-activators described above.

In Tables 11-1 to 11-3, certain strongly conserved residues are boldfaced or underlined.

The effect of no estradiol, the weak agonist estriol, and the antiestrogens tamoxifen, nafoxidine and clomiphene on the binding of these phage to the ER was investigated. Immobilized ER was incubated with 100 μ M estradiol, estriol, nafoxidine,

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As mentioned above, the ligands for orphan receptors are unknown. This poses an obstacle to receptor activation prior to phage display. Possible sources of ligand for orphan
5 receptors are fetal bovine serum or other enriched medium such as conditioned media from cells grown in culture. Once a peptide is identified that interacts with an orphan receptor in a serum or medium dependent manner, the peptide may be used to identify the natural ligand by fractionating serum, conditioned
10 medium or other appropriate tissue homogenate. The peptides may also be used to screen compound libraries for pharmacologically active agents.

Peptides that bind to the ligand binding domain of orphan
receptors may also be isolated. These peptides may be used in
15 a competitive displacement assay to identify the natural ligand
from a source described above. They may also be used to screen
libraries of compounds to identify pharmacologically active
species.

References for Example 11

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Table 11-1

Cluster Analysis for Estrogen Receptor

		Phage #
	S R T W E S P L G T W E W S R	13
5	S S K Y S Y S R S S E G H S R	29
	S S W V R L S D F P W G V S R	1
	S S W D R L S D F P W G V S R	2
	S S W I R L R D L P W G E S R	3
	S S W V L L R D L P W G S R	31
10	S S C K W Y E K C S G L W S R	7
	S S G I C F F W D G C F E S R	35
	S R N L C F F W D D E Y C S R	41
	H H H R H P A H P H T Y G G	47

0005033 03143
 06T00" 69E05060

Cluster Analysis for ER + Estradiol

[illegible]

Additional Sequences from ER + Estradiol

*binding not inhibited by antiestrogens

*binding not inhibited by antiestrogens

Additional Remarks

While the primary utility of the surrogate peptides disclosed above is in screening for non-peptide antagonists of the biological activity of the receptor to which they bind, the peptide may also themselves be useful as agonists or antagonists of the receptor, or as the binding component of a diagnostic agent which binds the receptor.

Surrogate peptides of particular interest, either in screening, or as therapeutic or diagnostic agents in their own right, include (but are not limited to) those comprising the sequences shown below:

<u>Ex</u>	<u>Target</u>	<u>Peptide</u>
1	HCMV UL44	E-H-V-C-S-W-G-W-G-R-C D R L T K N K I A H Q M G
2	Protein Kinase C β II	W-Phi-C-Pho-G-X- (F/L) -C and W-T-C- (V/I) -N- C
3	human MDM2	S-F-T-D-Y-W-R-D-L-E-Q and conservative mutants thereof.
5	tyrosine tRNA	Y-Phi-W-P-W and Y-Phi-W-P-Phi and (Y/F) - (S/T/G/A/H) -W-P (W/G/D/S/P) and (Y/F/W/L) -W-W-P- (D/E/S/N) -W-G
8	glucosidase	{F-K}-P-W-P- (I/V) -Y { }=optional
	carboxypeptidase	P-G-W-W
	ProRS	S-R-D-W-G-F-W E
11	Estrogen Receptor	W-Pho-R-L-Phi-D-Pho-P-W-G and C-F-F-W-D and L-X-X-L-L

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CLAIMS

We hereby claim:

1. A method of identifying a ligand which can mediate the biological activity of a target protein via inhibition of the binding of a target protein to a binding partner which comprises

(a) screening a first combinatorial library comprising a plurality of first member ligands for binding to the target-binding ligands, thereby identifying one or more target-binding ligands,

(b) screening a second library comprising a plurality of second member ligands for the ability to inhibit the binding of one or more of said target-binding ligands to said target protein, thereby obtaining one or more inhibitory ligands, and

(c) determining which of the inhibitory ligands can mediate a biological activity of the target protein.

2. The method of claim 1 in which the first combinatorial library is composed of peptides and/or peptoids.

3. The method of claim 1 wherein the first combinatorial library is composed of nucleic acids.

4. The method of claim 1 in which the first combinatorial library is composed of peptides, peptoids and/or nucleic acids, and the second library is not.

5. The method of claim 1 in which the first combinatorial library has a greater diversity than the second library.

6. The method of claim 1 in which the second library is a combinatorial library.

7. The method of claim 1 in which the target-binding ligands obtained in step (a) are tested in a suitable biological system for the ability to interact with the target protein so as to mediate its biological activity and only the effective ligands are used in screening step (b).

8. The method of claim 1 in which the inhibitory ligands obtained in step (b) are tested to determine whether their inhibitory action is attributable to their binding the target

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9. The method of claim 1 in which the first library is a peptide library and the second library is a benzodiazepine library.

21. A structured panel of biased combinatorial peptide
35 libraries, each library having one or two constant residues,

wherein, in each component library, at a first fixed position within the middle 50% of the peptide, the amino acid assigned to said first position is constant within said component library, is not the same in all libraries of said panel, and
 5 as a result of such library-to-library variation, said panel collectively presents all possible genetically encoded peptides of a predetermined length.

22. The panel of claim 21 wherein said peptides are of the form

10 $(\text{Xaa})_m\text{-R1-(Xaa)}_n,$

where R1 is the amino acid at said first fixed position, and m and n do not differ by more than two.

23. The panel of claim 21, said structured panel further characterized in that in each library, a second position is
 15 held constant, but the location of said second position is varied so that said second position scans all residue position except for said first position, whereby the panel is composed of subpanels in which said first and second positions are fixed, and where, in each subpanel, the amino acid assigned to
 20 said second position is constant within said component library, but varies from library to library within said subpanel.

24. A biased combinatorial peptide library of the form

$(\text{Xaa})_m\text{-Cys},$

where m is greater than or equal to 5.

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ABSTRACT OF THE DISCLOSURE

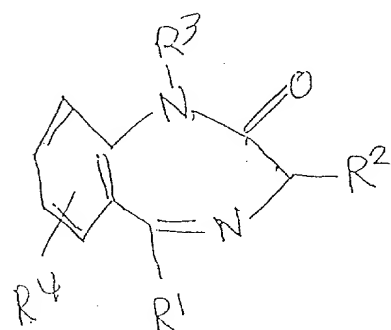
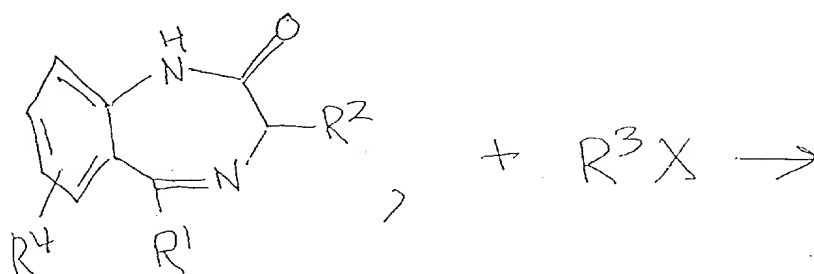
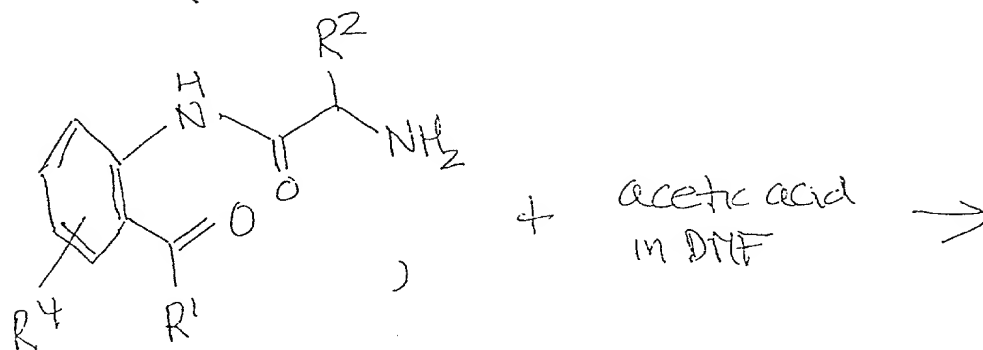
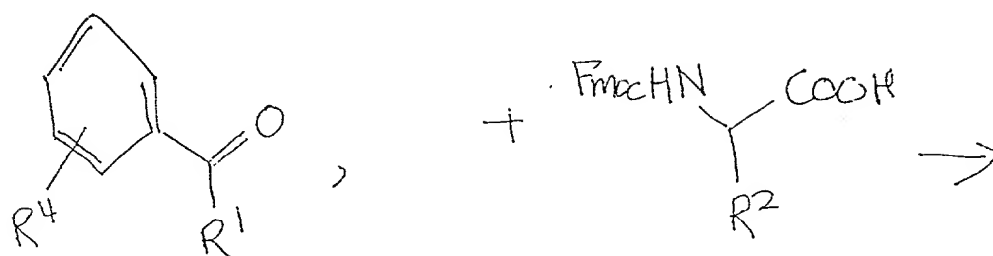
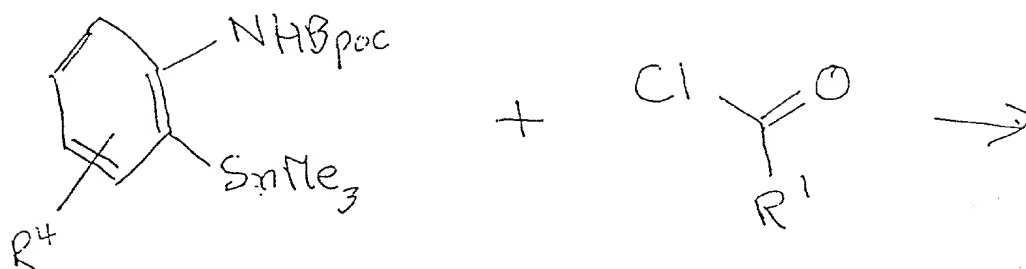
The present invention is directed to the identification of compounds in a compound library which can mediate the biological activity of a target receptor protein, even when the
5 ligands which mediate that activity through binding to that receptor are not already known. Such compounds can then be used as "drug leads", i.e., used as a starting point for the design of analogues which can in turn be tested for activity.

The method of the invention comprises three steps:

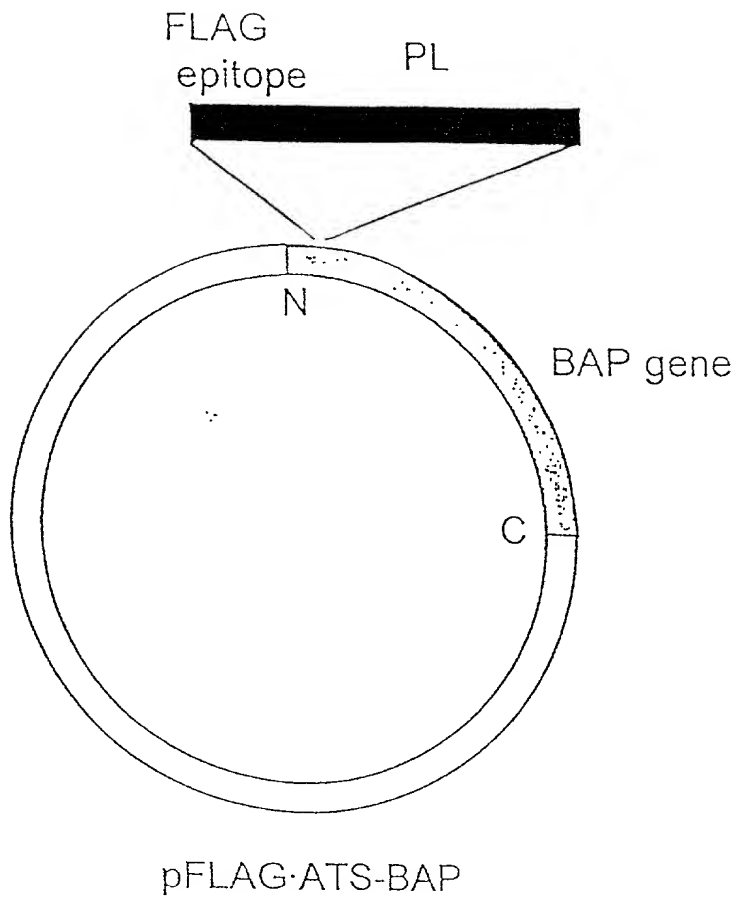
- 10 (1) Screen at least one potential surrogate combinatorial library for members (preferably peptides or nucleic acids) binding to the target protein (TP) and hence capable of use as surrogates for the unknown ligand in steps (2) and (3).
- 15 (2) Screen at least one complementary library, preferably a combinatorial library, (which is not limited to, and may not even include, peptides, or nucleic acids and hence is referred to on occasion
20 as a "compound library") for compounds which inhibit the binding of one or more surrogates from step (1) to TP, and, optionally.
- (3) Determine whether the inhibitory compound mediates the biological activity of the said TP.

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Fig. 1



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Fig. 2

Fig.3 Occurrence of Amino Acids (AA) in Binding Peptides Identified by screening from Phage Display Libraries

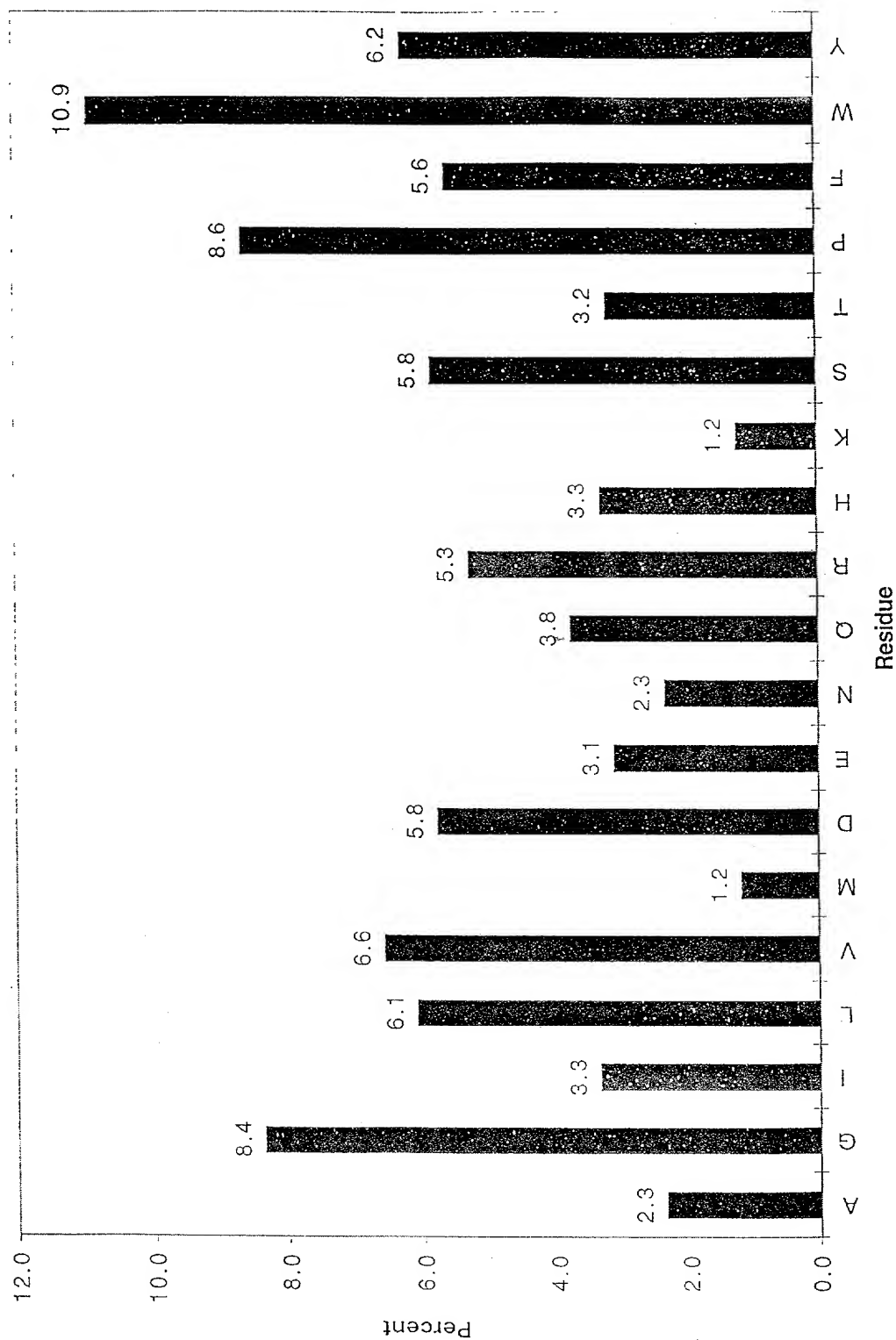
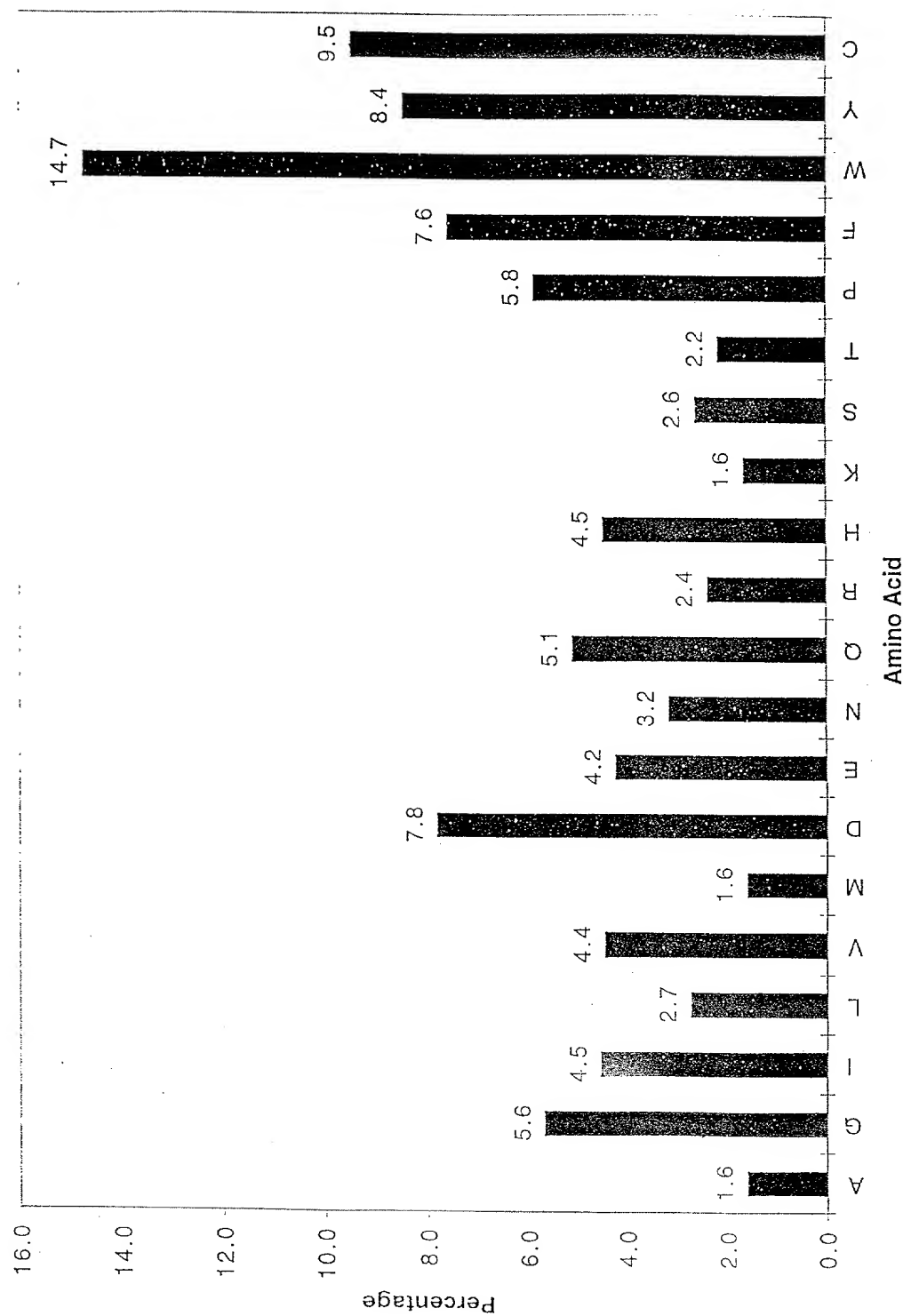


Fig.4 Occurrence of Amino Acids (AA) in Binding Peptides Identified by screening from Phage Display Libraries (Corrected for codon usage)



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Enrichment of Phage Binding to CMV UL44

Fig. 5

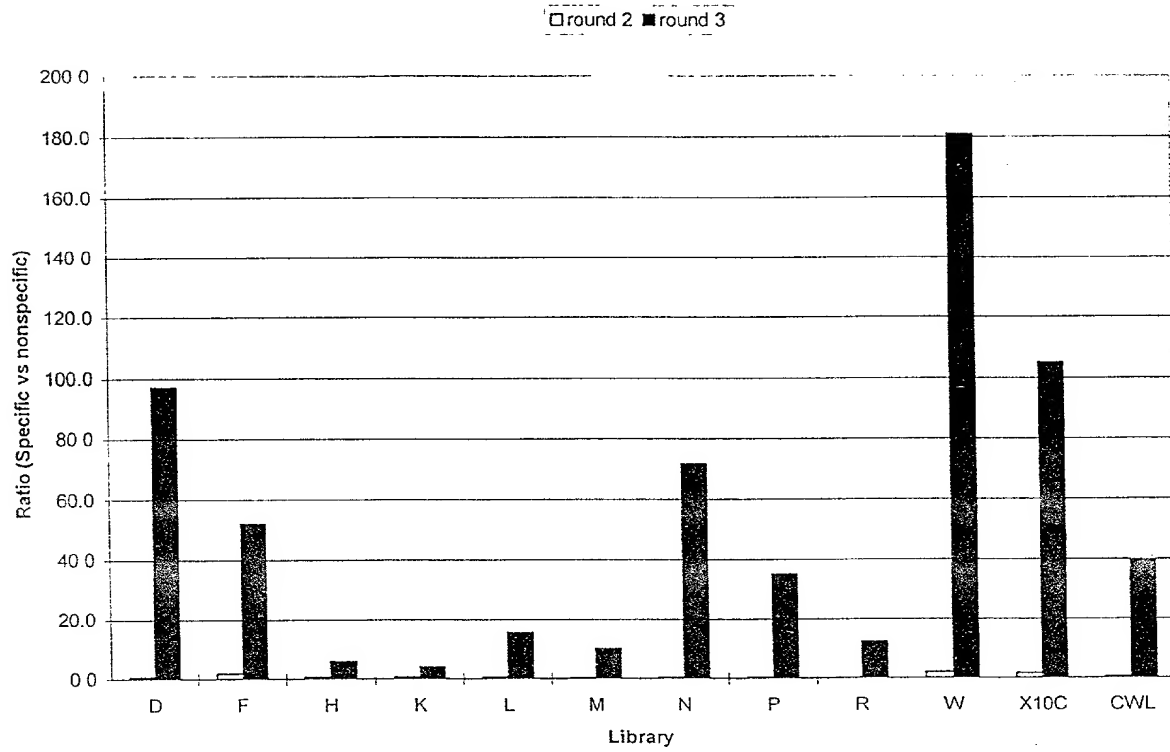


Fig. 6

Individual Phage ELISAs for CMV UL44

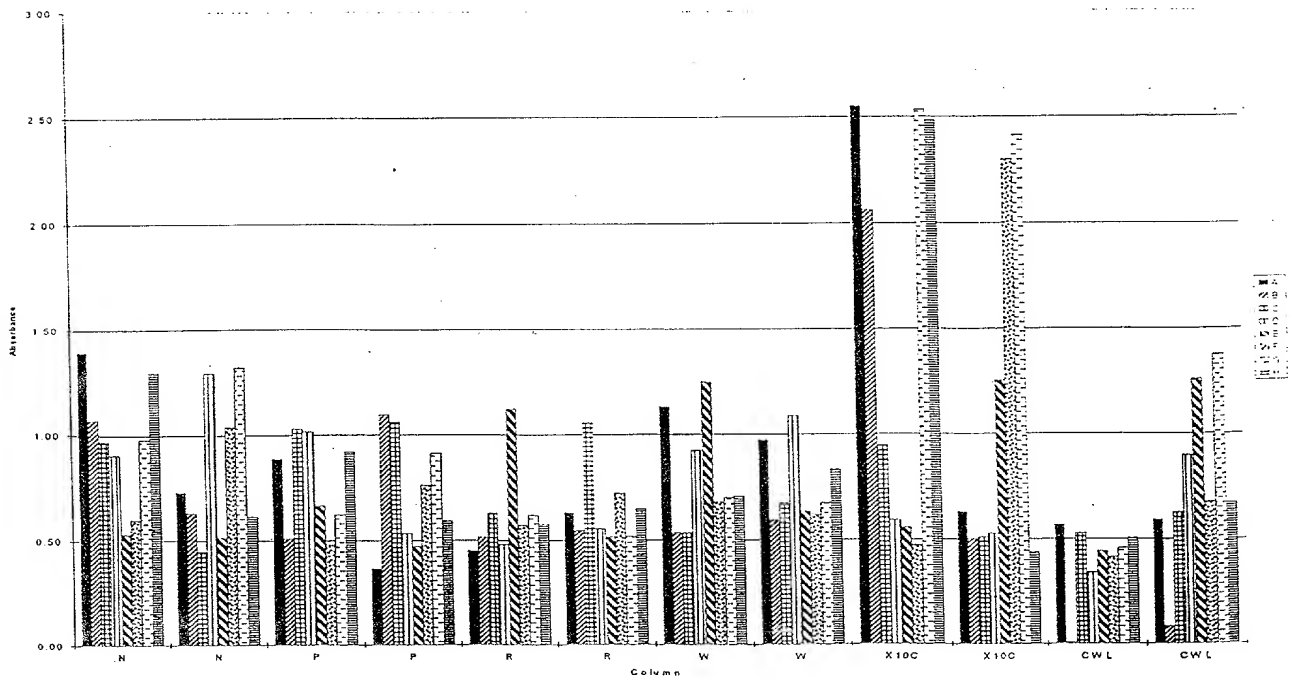


Fig. 7a

Competition for binding between phage and substrates

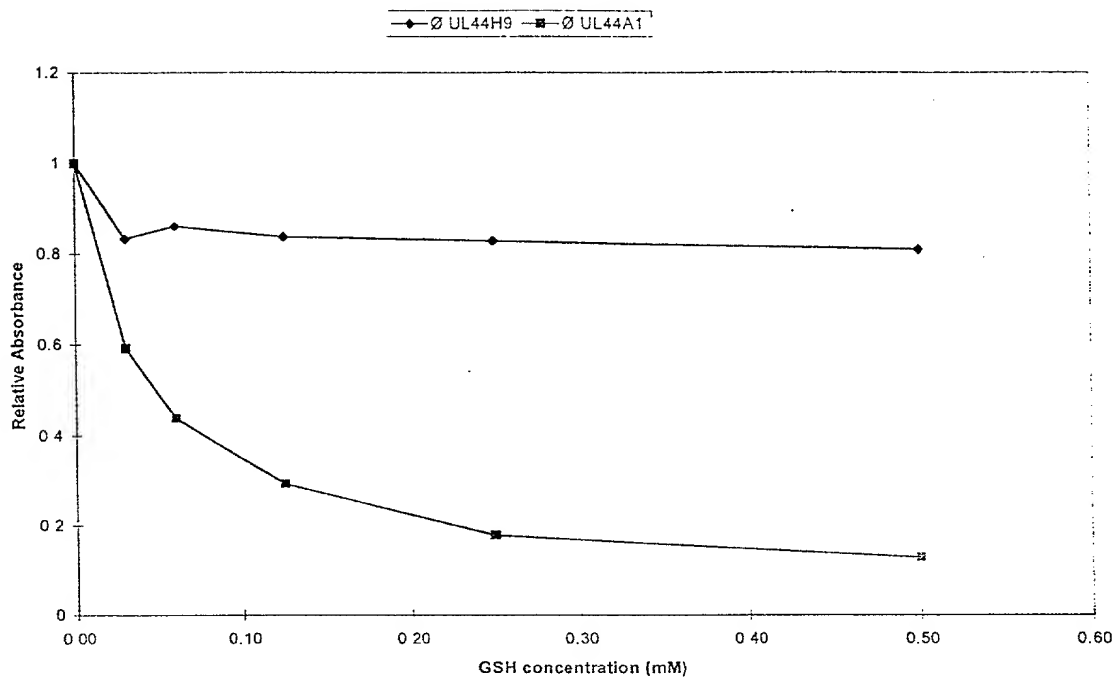
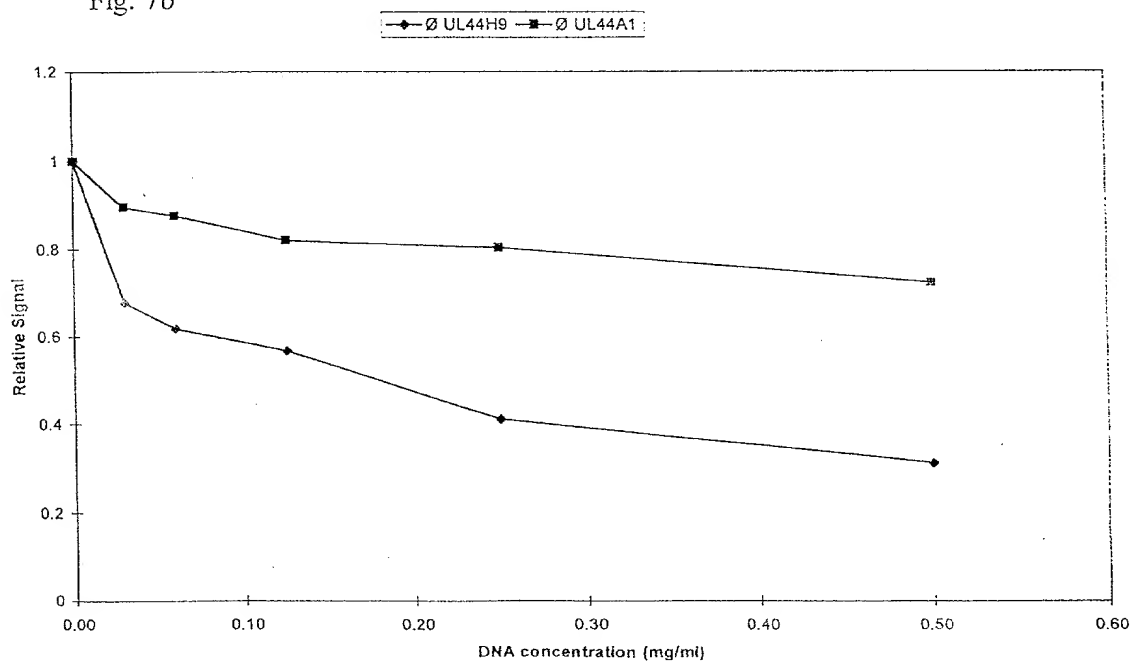


Fig. 7b



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Fig. 8 ELS Assay of UL44 BioKey

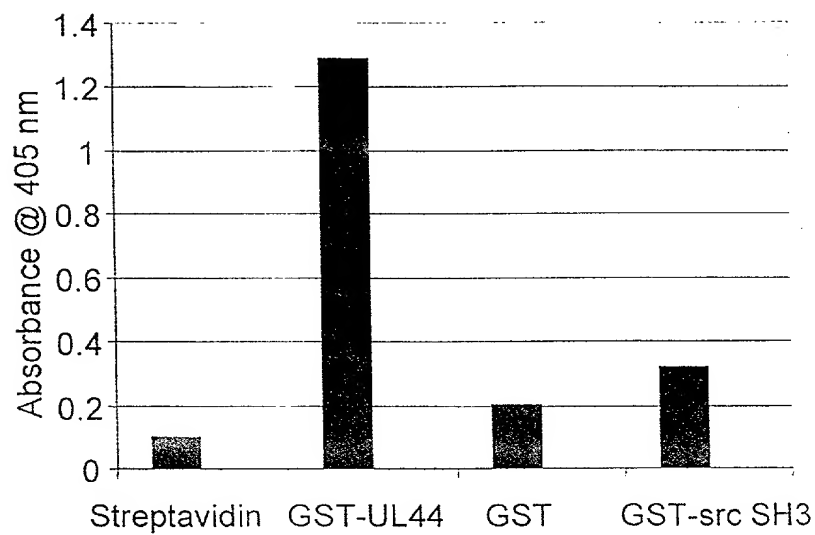


Fig. 9

Time Course of 50 pmol UL44-BioKey Binding to GST-UL44

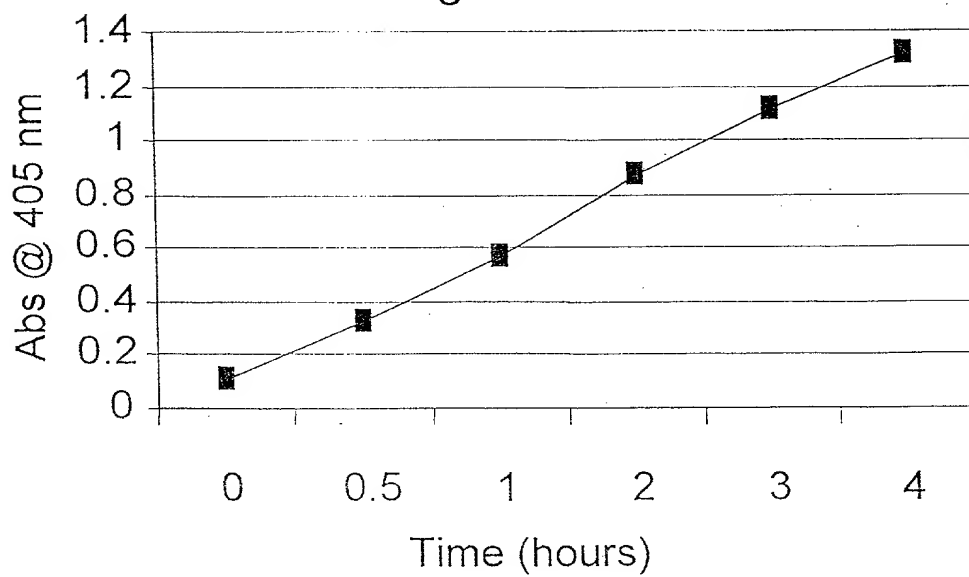


Fig. 10

Titration Curves for Binding of UL44 BioKey to GST-UL44

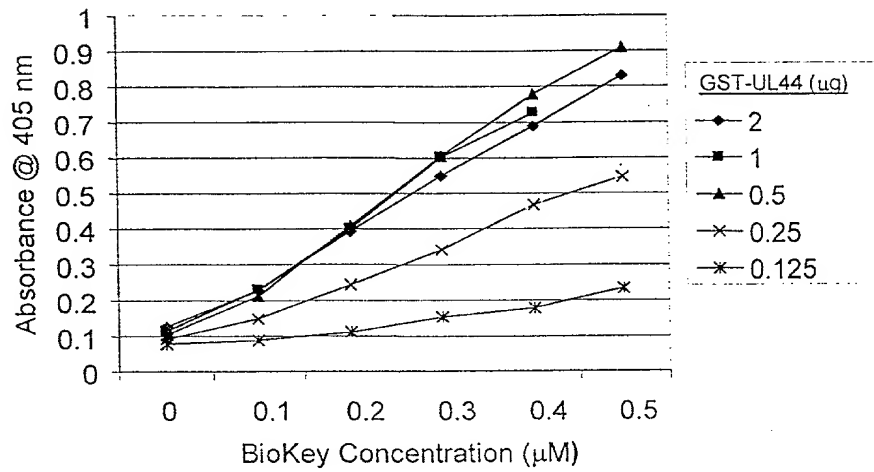


Fig. 11

UL44 BioKey: Self Competition

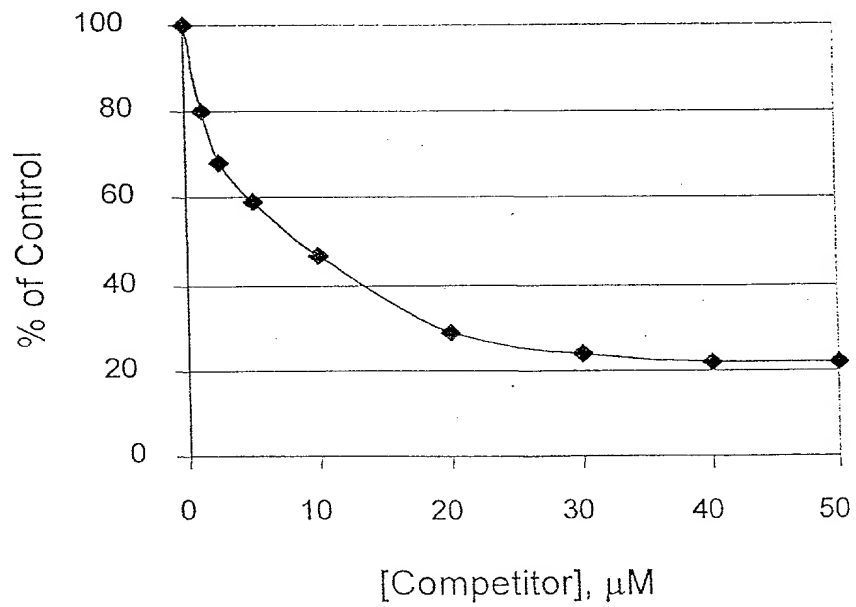
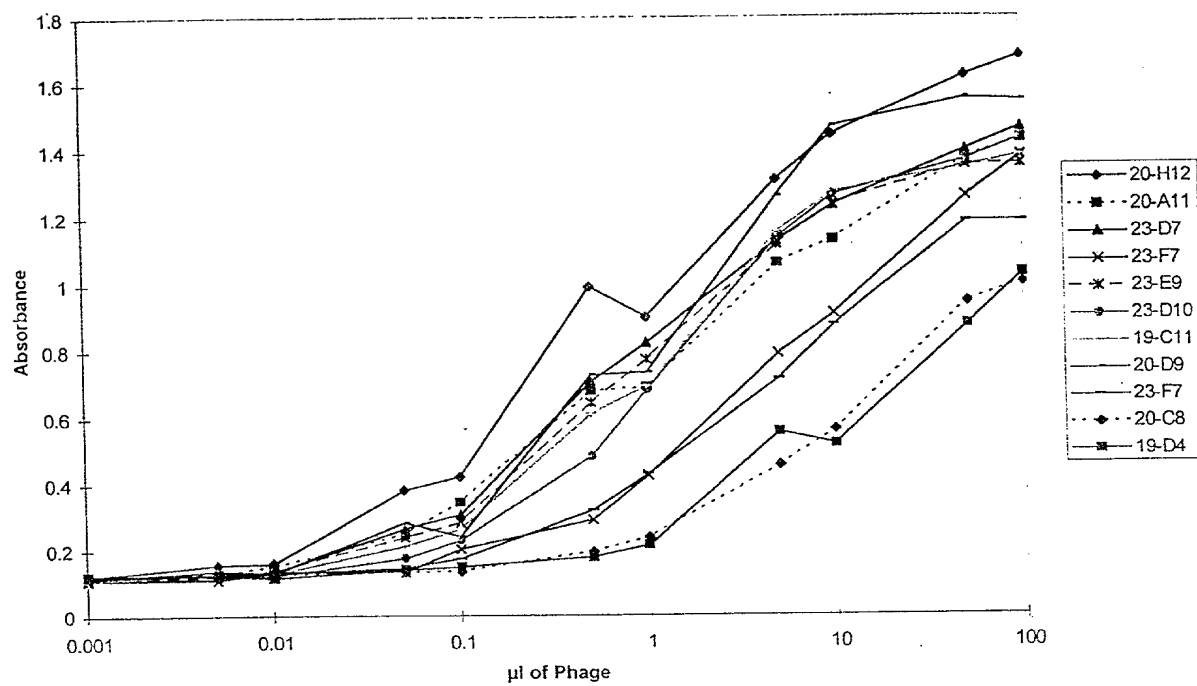


Fig. 12

Titration of PKC Binding Phage



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Fig. 13a

Specific Binding of the ProRS BioKey

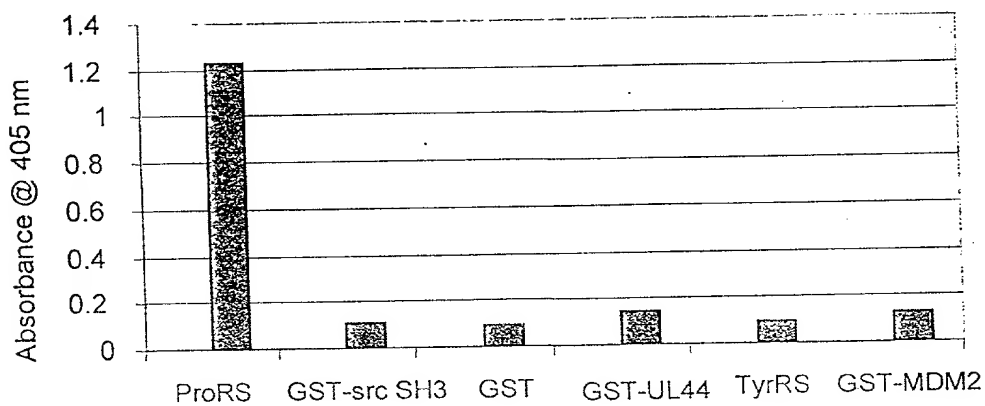
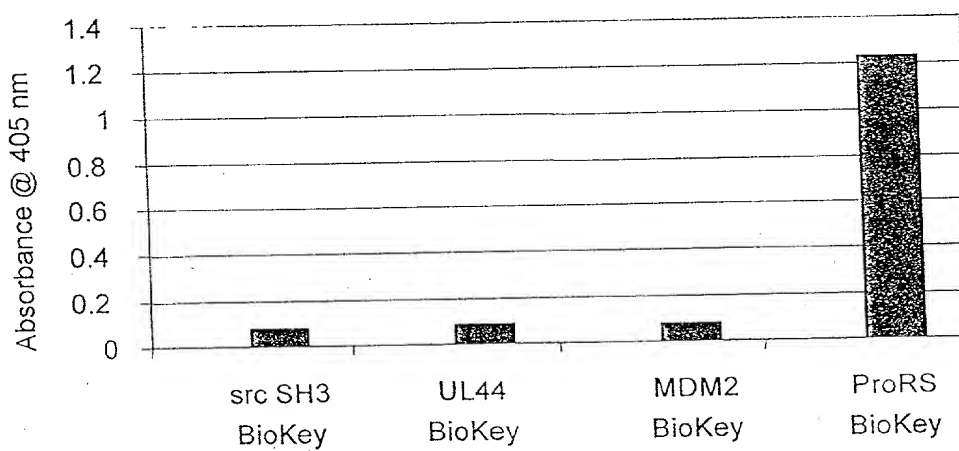


Fig. 13b

BioKeys to Other Targets do not Bind to ProRS



All incubations: 50 pmol BioKey for 2 hours, 5 min development

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Fig. 14a

Self Competition for ProRS BioKey 5 min development

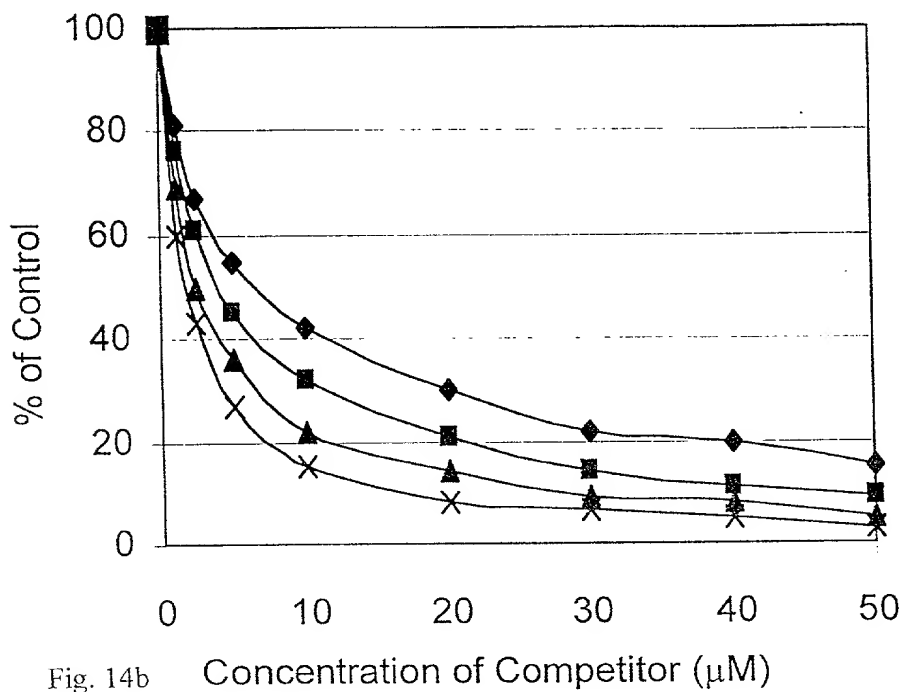
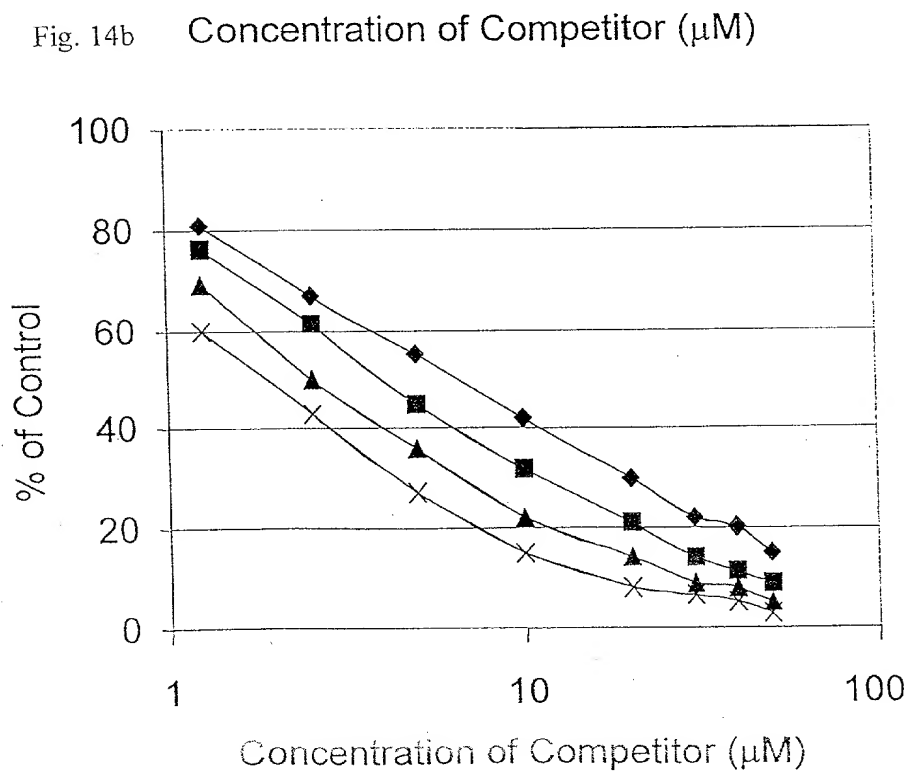


Fig. 14b



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Fig. 15a

ProRS BioKey Concentration Curve
2 h incubation, 5 min. development

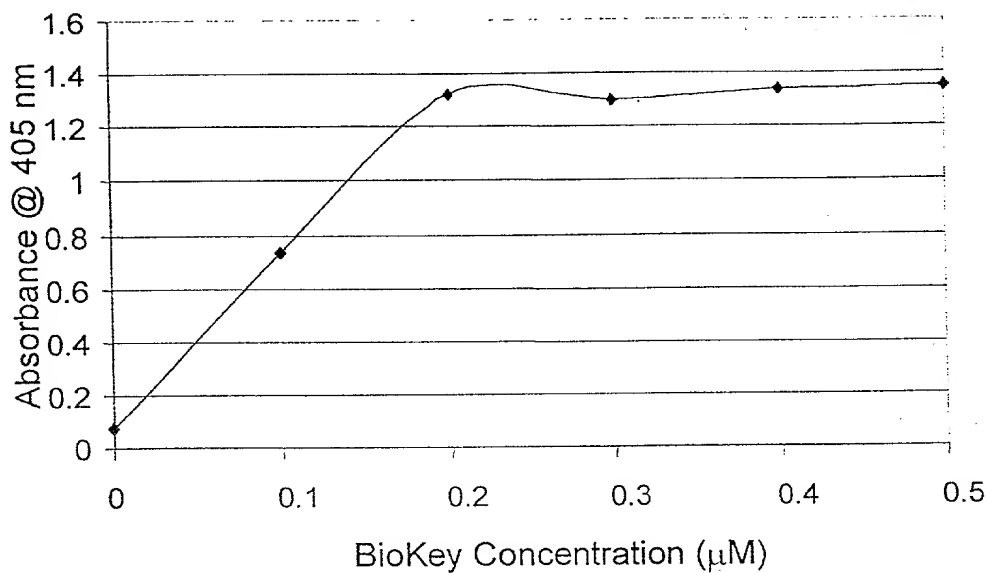


Fig. 15b

ProRS BioKey Binding Time Course
0.5 μM BioKey, 5 min. development

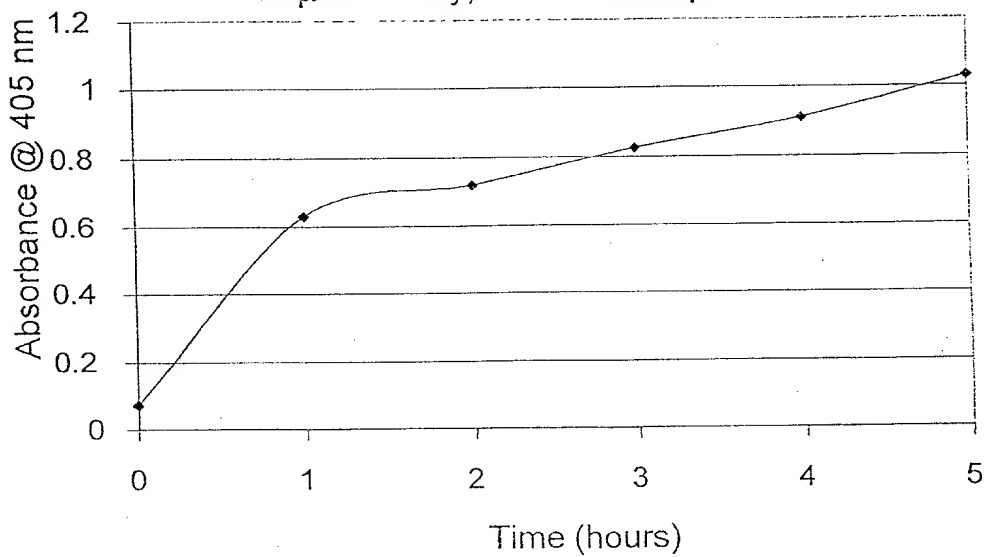


Fig. 16

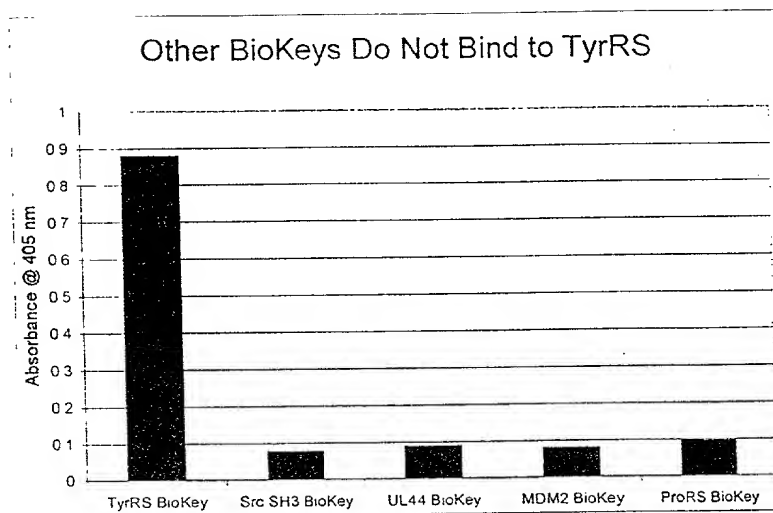


Fig. 17

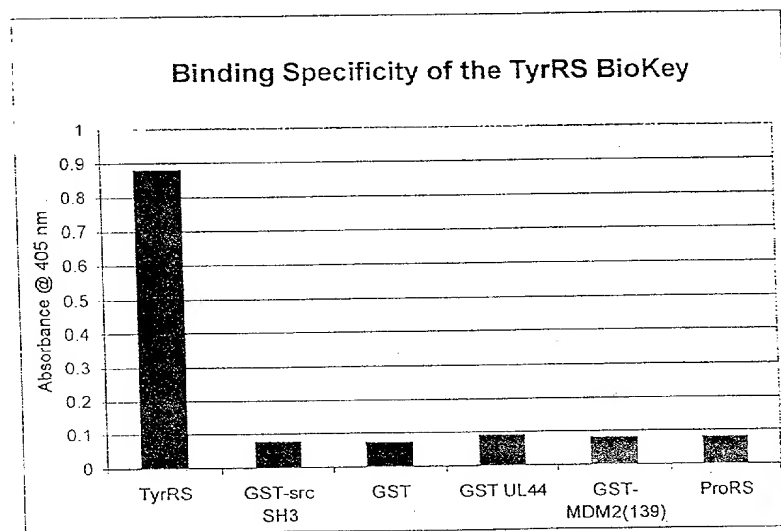


Fig. 18

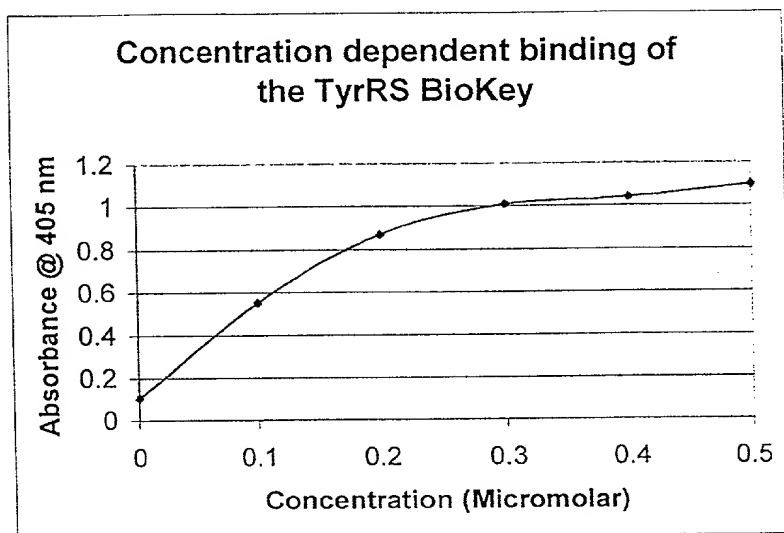


Fig. 19

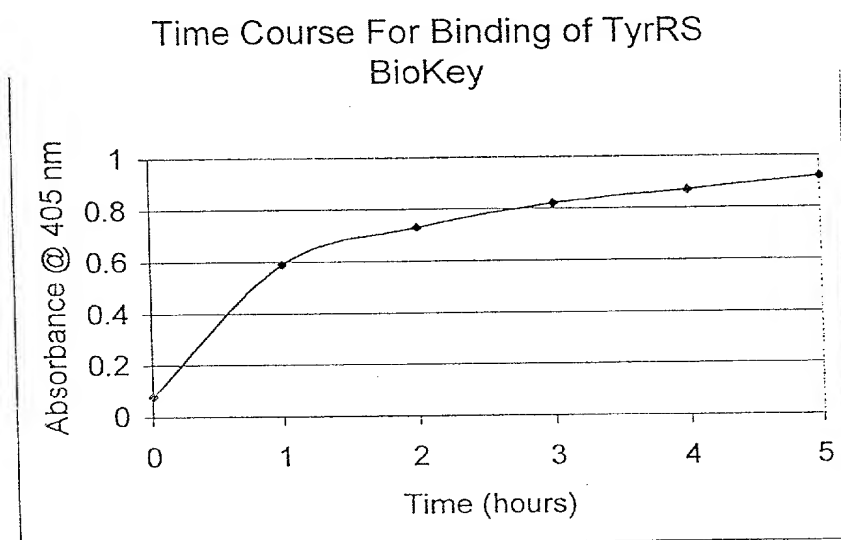


Fig. 20

Competition between HiTryRS BioKey and CB239 and CB16914

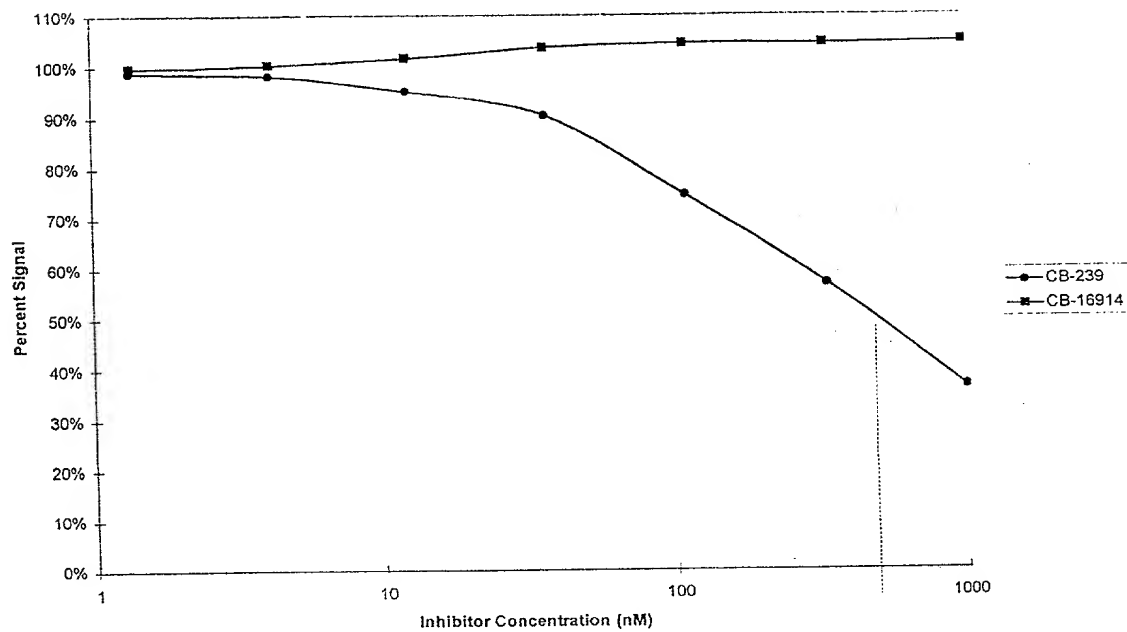
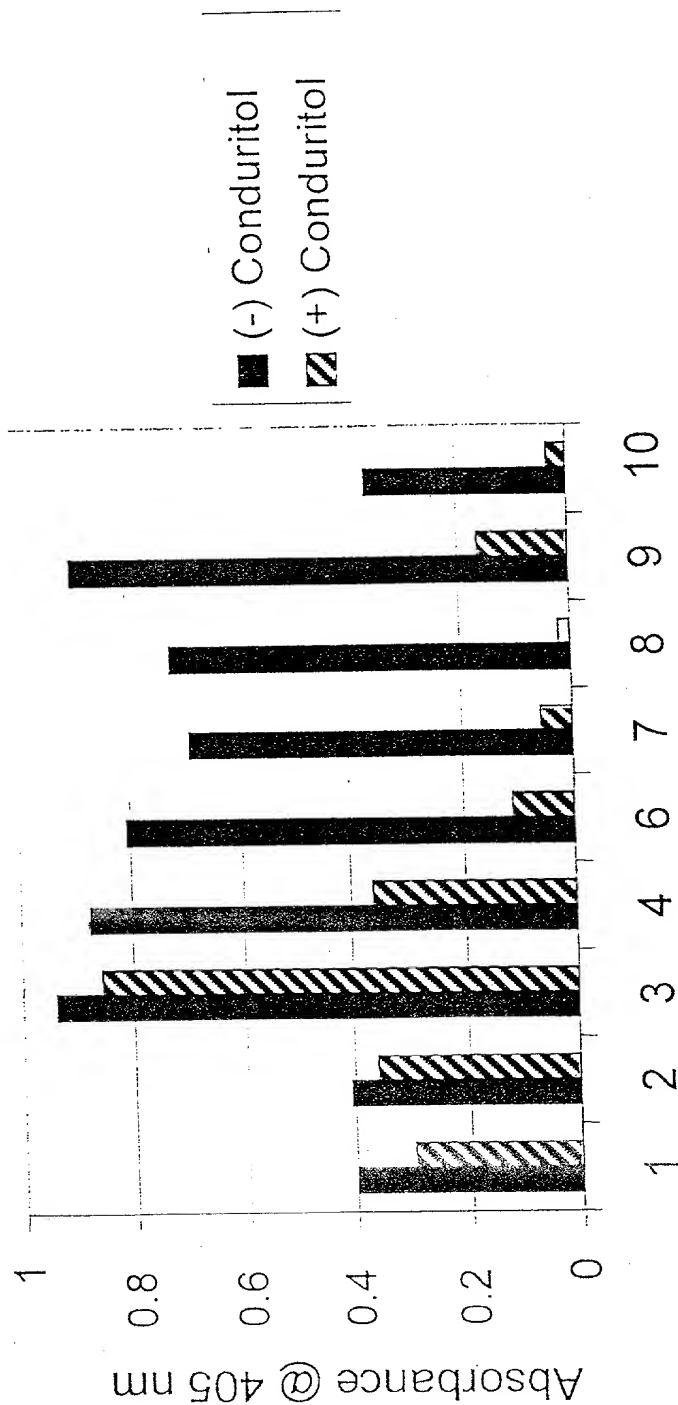


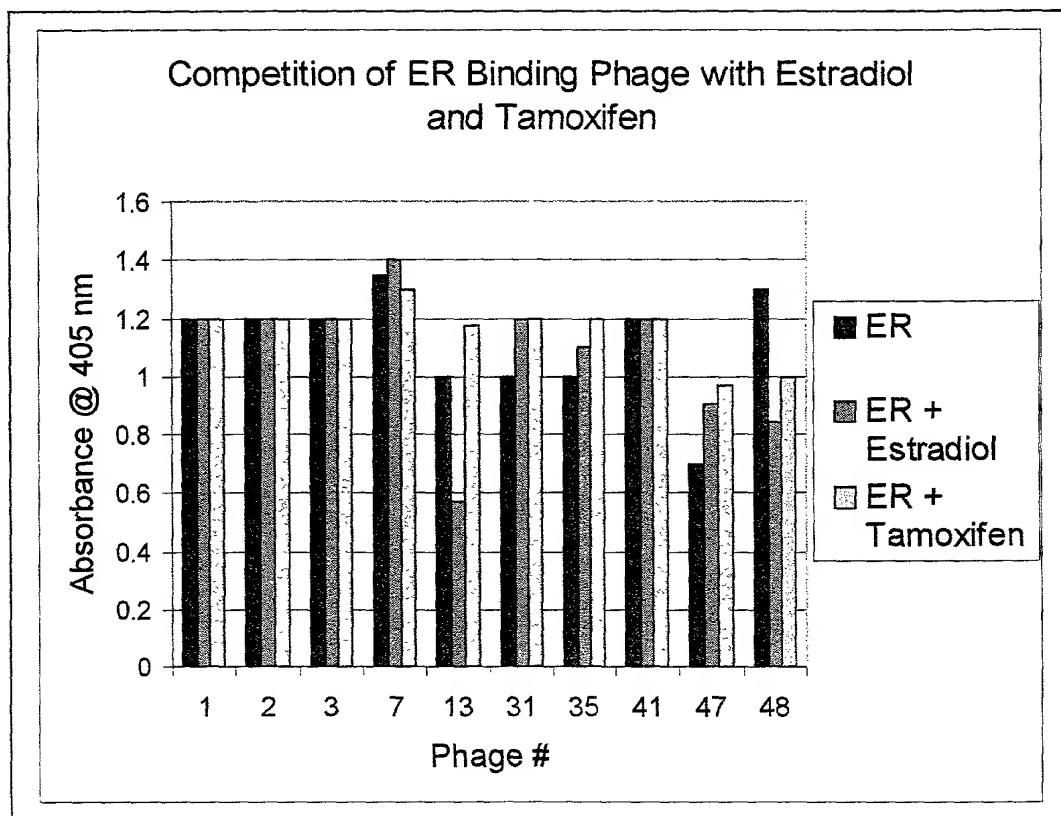
Fig. 21

Inhibition of B-Glucosidase Phage ELISA with Conduritol B Epoxide

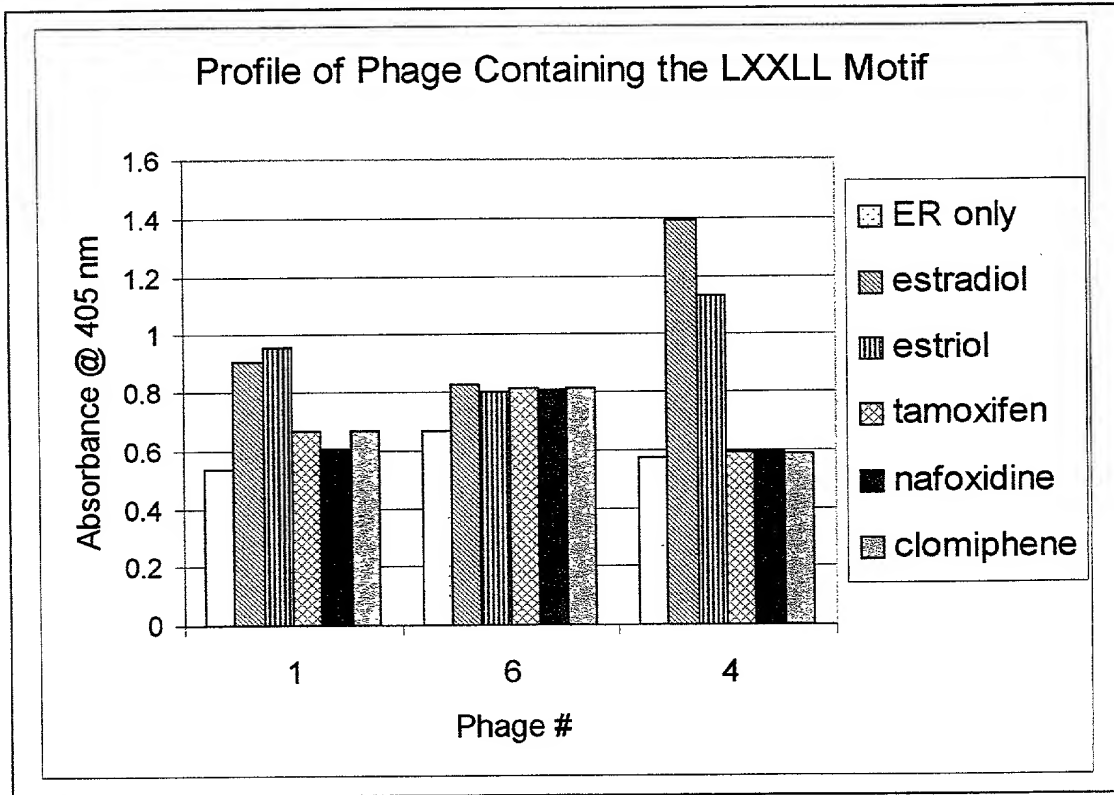


Estrogen Receptor Structural and Functional Domains





24A



24B

